

## RELATIONSHIP BETWEEN GAS EXCHANGES IN INTACT ROOTS AND WATER UPTAKE IN RESPONSE TO LEAF TRANSPIRATION IN HYDROPONICS

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## RELATIONSHIP BETWEEN GAS EXCHANGES IN INTACT ROOTS AND WATER UPTAKE IN RESPONSE TO LEAF TRANSPIRATION IN HYDROPONICS

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YOSHIDA S. and EGUCHI H. *Relationship between gas exchanges in intact roots and water uptake in response to leaf transpiration in hydroponics*. BIOTRONICS 17, 59–68, 1988. A hydroponic system was developed for intact measurements of water uptake rate, dissolved O<sub>2</sub> and CO<sub>2</sub> concentrations in nutrient solution under controlled environment. The relationship between water uptake rate and gas exchange in intact roots of cucumber plants was analyzed in response to stomatal transpiration in the treatments that leaf transpiration was promoted by lighting and was inhibited by applying ABA and microcrystalline wax to the leaves. The decrease rate of O<sub>2</sub> was not necessarily associated with water uptake rate. On the other hand, increase rate of CO<sub>2</sub> was higher in the lighted plants in which leaf transpiration and water uptake were inhibited. The CO<sub>2</sub> concentration continued to increase even in O<sub>2</sub> deficit solution in the transpiring plants. From the fact that water uptake is caused by leaf transpiration and gas exchange in leaf is promoted in transpiring leaf, gas exchange in leaf was considered to be responsible for gas exchange in roots, and it is possible that O<sub>2</sub> transported from leaves to root system may be used for root respiration.

**Key words:** *Cucumis sativus* L.; cucumber plant; hydroponics; gas exchange; dissolved O<sub>2</sub> concentration; dissolved CO<sub>2</sub> concentration; water uptake rate; leaf transpiration; root respiration.

### INTRODUCTION

Plant growth is inhibited by O<sub>2</sub> deficiency or toxicity of accumulated CO<sub>2</sub> in flooded soil (4, 8, 16, 24, 25) and in poor aerated root medium (6, 14, 20, 31–33). The gas exchange in roots is responsible for water uptake, water movement (1, 13, 23, 30), and the plant growth (3, 7, 9–12, 15, 17, 18, 27–29, 34). For better understanding of root system, it is further necessary to examine environmental effects on root function.

The present paper deals with analysis of gas exchange in roots of whole plant in hydroponics.

## MATERIAL AND METHODS

*Hydroponic system*

A hydroponic system was designed and constructed to measure water uptake rate in intact roots, pH,  $O_2$  and  $CO_2$  concentrations in nutrient solution under controlled environment. Figure 1 shows the hydroponic system. Ambient air temperature and relative humidity were controlled at  $25^\circ C$  and 40% in a growth chamber where light intensity was  $200 \mu mol m^{-2} s^{-1}$  (fluorescent lamps; FLR110-EHW/A, Toshiba Corporation). Temperature of nutrient solution in a stainless steel pot was controlled at  $25 \pm 0.1^\circ C$  by a water bath where water ( $15 l min^{-1}$ ) was circulated from water temperature controller (constant temperature circulator; CL-300, Taiyo Scientific Industrial Co., Ltd.). The water bath was covered with styrofoam for insulating heat.

*Measurements*

Polarographic dissolved  $O_2$  meter (UD-1, Central Kagaku Co., Ltd.),  $CO_2$  meter (CGP-1, Toa Electronics Ltd.) and pH meter (HM-7E, Toa Electronics Ltd.) were employed for the measurements in nutrient solution. The stainless steel pot was filled with nutrient solution (3.7 litre) saturated with air, and roots of a material plant was submerged in the solution. The plant and sensors were fixed at the pot surface by using rubber corks where silicone grease was applied to airproof the system. The solution was continuously stirred by a magnetic stirrer to homogenize

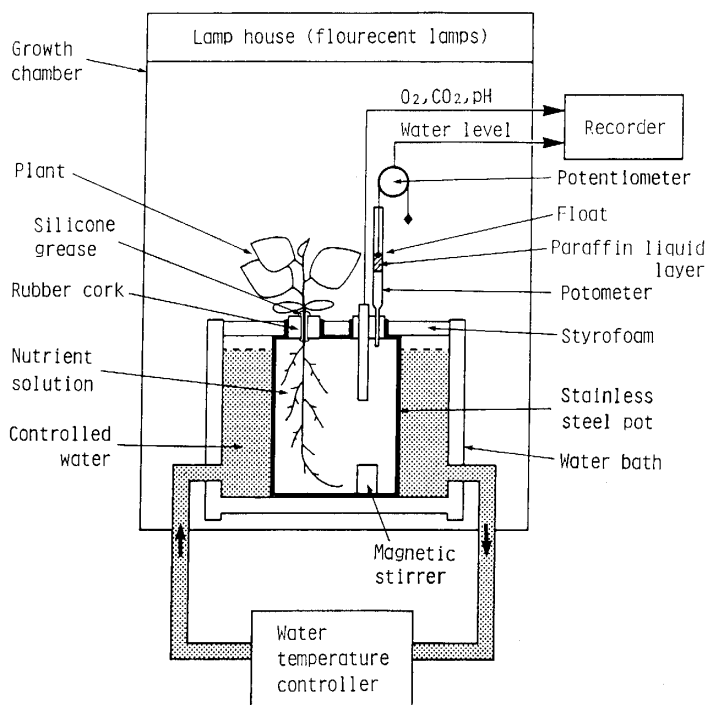


Fig. 1. Schematic diagram of hydroponic system for on-line measurements of pH, water uptake rate, dissolved  $O_2$  and dissolved  $CO_2$  concentrations in controlled temperature of nutrient solution.

the solution. Water uptake rate in roots was measured automatically by a potometer: Decrease of the solution in the potometer was detected by a float which was connected to a potentiometer. Surface of the solution in the potometer is sealed with paraffin liquid layer to prevent diffusion of air into the solution. The sensor signals of the level of the solution in the potometer, pH, O<sub>2</sub> and CO<sub>2</sub> concentrations in the solution were recorded in course of time.

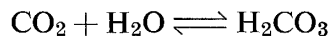
#### *Plant material*

Cucumber plants (*Cucumis sativus* L. "Chojitsu-Ochiai") were used in this experiment. Plants were grown at air temperature of 23°C, relative humidity of 70%, and light intensity of 250  $\mu\text{mol m}^{-2} \text{s}^{-1}$  (metal halide lamps; Yoko lamp, DR400, Toshiba Corporation) in photoperiod of 12 h. The seeds were sown in Vermiculite moistened with tap water. The seedlings were transplanted at the cotyledonary stage to hydroponic system and were grown in fully aerated nutrient solution at about 0.25 mmol l<sup>-1</sup> of dissolved O<sub>2</sub>. The 3 leaf stage plant was kept under the experimental condition for 18 h in the growth chamber. Thereafter, measurements were started, as soon as the plant was transplanted to the pot of the hydroponic system shown in Fig. 1.

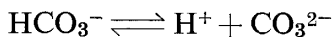
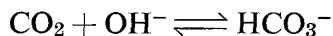
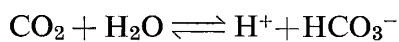
Total leaf area, fresh and dry weight of whole root system in the cucumber plant used were  $577 \pm 99 \text{ cm}^2$ ,  $6.0 \pm 1.3 \text{ g}$  and  $0.22 \pm 0.06 \text{ g}$ , respectively.

#### *Analysis of CO<sub>2</sub> concentration*

Dissolved CO<sub>2</sub> in water is hydrated to form H<sub>2</sub>CO<sub>3</sub> and unhydrated CO<sub>2</sub> is in equilibrium with H<sub>2</sub>CO<sub>3</sub> as expressed by



Then unhydrated and hydrated CO<sub>2</sub> are ionized to form HCO<sub>3</sub><sup>-</sup> and CO<sub>3</sub><sup>2-</sup> as follows;



For analysis of increase of dissolved CO<sub>2</sub> related to gas exchange in roots in hydroponics, it is necessary to evaluate concentration of total inorganic carbon in nutrient solution.

The equilibrium constants are defined at constant water temperature on respective reactions in dissociation and ionization of inorganic carbon. However, K<sub>a1</sub> is commonly applied to the first ionization of unhydrated and hydrated CO<sub>2</sub> molecules;

$$K_{a1} = \frac{[\text{H}^+][\text{HCO}_3^-]}{[\text{CO}_2 + \text{H}_2\text{CO}_3]}$$

The second ionization constant is defined as

$$K_{a2} = \frac{[H^+][CO_3^{2-}]}{[HCO_3^-]}$$

Molarity of respective components of inorganic carbon at constant temperature is related to pH of solution as represented by Henderson-Hasselbach equations;

$$pH = pK_{a1} + \log \frac{[HCO_3^-]}{[CO_2 + H_2CO_3]}$$

$$pH = pK_{a2} + \log \frac{[CO_3^{2-}]}{[HCO_3^-]}$$

where  $pK_{a1} = -\log K_{a1}$ , and  $pK_{a2} = -\log K_{a2}$

From the fact that pH of nutrient solution was 5.0 to 6.0 in this system, it was estimated that the components of inorganic carbon in the solution are  $CO_2$ ,  $H_2CO_3$  and  $HCO_3^-$ , and total inorganic carbon molarity is total molarity of these components of inorganic carbon.

The  $CO_2$  sensor employed can detect both  $CO_2$  and  $H_2CO_3$ , and the sum of them was used as  $CO_2$  concentration. So, the molarity of  $HCO_3^-$  was calculated from measured values of  $CO_2$  concentration and pH as follows;

$$\log[HCO_3^-] = \log[CO_2 + H_2CO_3] + pH - pK_{a1}$$

where  $K_{a1} = 4.45 \times 10^{-7}$  (mol  $l^{-1}$ ) at solution temperature of  $25^\circ C$  as given by Helder (19). Thus,  $CO_2$  concentration of the solution in the hydroponic system was evaluated by the molarity of total inorganic carbon.

## RESULTS AND DISCUSSION

For examination of gas exchange in roots, dissolved  $O_2$  and  $CO_2$  concentrations in nutrient solution of hydroponics were measured. The concentrations of  $O_2$  and  $CO_2$  in fully aerated solution were  $0.24 \pm 0.01$  mmol  $l^{-1}$  and  $0.05 \pm 0.02$  mmol  $l^{-1}$ , respectively. Figure 2 shows water uptake rate,  $O_2$  and  $CO_2$  concentrations in the hydroponics where plants were cultured in photoperiod of 12 h. The concentration of  $O_2$  decreased to about 0.1 mmol  $l^{-1}$  at 30 h after the start of measurements, and thereafter became constant. The concentration of  $CO_2$  continued to increase even after the time when  $O_2$  concentration was the lowest, and it reached steady-state at about 3.7 mmol  $l^{-1}$  at 130 h after the start of measurements. Water uptake rate oscillated, synchronizing with the photoperiod: It increased in the light and decreased in the dark. This oscillation gradually damped, and water uptake rate became lower in course of time. Figure 3 shows water uptake rate,  $O_2$  and  $CO_2$  concentrations in the solution in which the plants were cultured in the continuous light. The concentration of  $O_2$  decreased to about 0.1 mmol  $l^{-1}$  at 20 h after the start of measurements, and the concentration of  $CO_2$  increased to about 3.5 mmol  $l^{-1}$  at 90 h after the start of measurements in the light. Even in the continuous light, periodical change of water uptake rate was observed during 2 days and was considered to be caused by habituation of the photoperiod during seedling culture. Thereafter, water uptake rate gradually decreased. Thus, it was found that  $CO_2$  release in intact roots

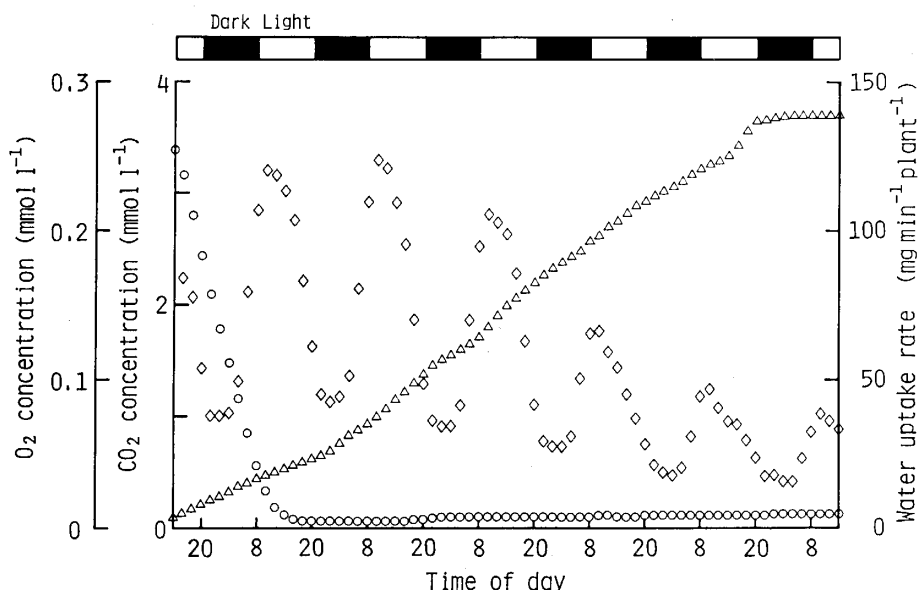


Fig. 2. Time course patterns of water uptake rate ( $\diamond$ ), dissolved  $O_2$  ( $\circ$ ) and dissolved  $CO_2$  ( $\triangle$ ) concentrations in hydroponics in photoperiod of 12 h (8:00–20:00).

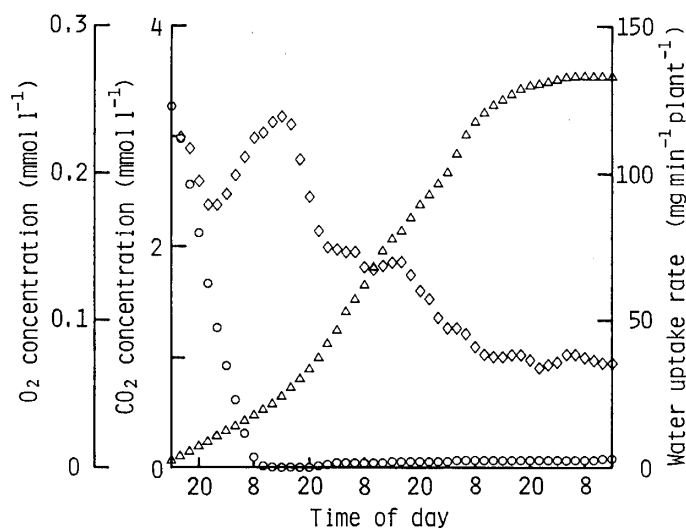


Fig. 3. Time course patterns of water uptake rate ( $\diamond$ ), dissolved  $O_2$  ( $\circ$ ) and dissolved  $CO_2$  ( $\triangle$ ) concentrations in hydroponics under continuous light condition.

was caused even in the  $O_2$  deficit solution.

Figure 4 shows water uptake rate (a),  $O_2$  (b) and  $CO_2$  (c) concentrations in the solution of hydroponics where the plants were lighted and were kept in the dark. Water uptake rate in the light was about two times as much as that in the dark. Decrease rate of  $O_2$  and increase rate of  $CO_2$  in the light were higher than those in the dark. The  $CO_2$  concentration in the light was about four times as much as that

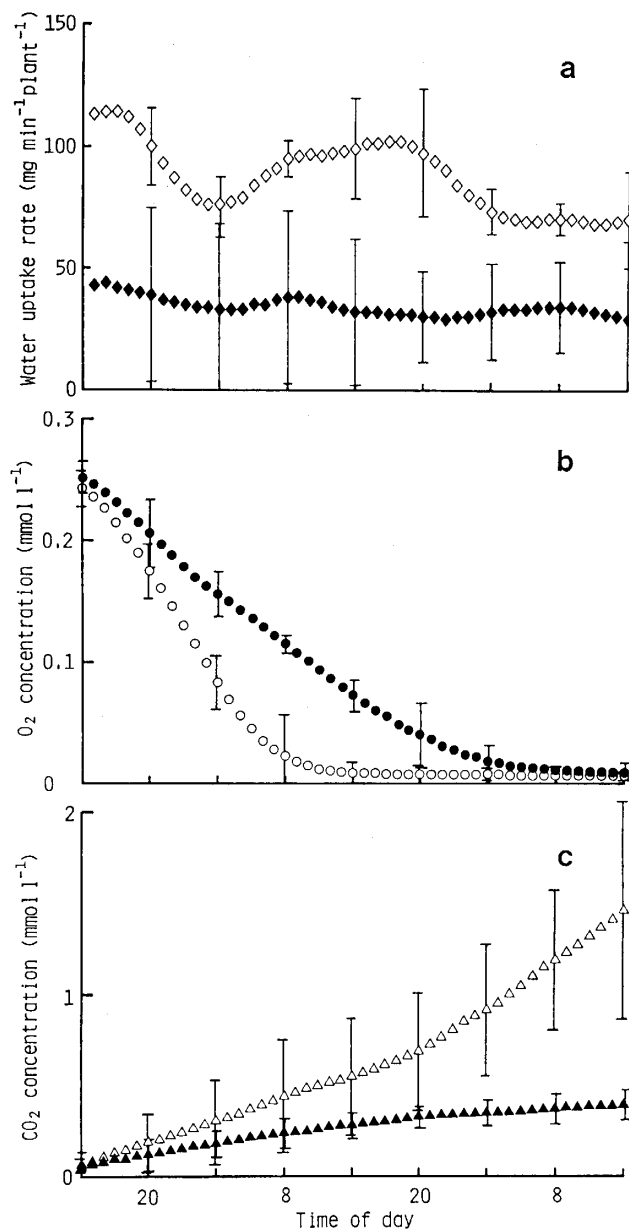


Fig. 4. Time course patterns of water uptake rate (a), dissolved O<sub>2</sub> (b) and dissolved CO<sub>2</sub> (c) concentrations in hydroponics of intact plants under light and dark conditions, where means of measured values in 3 plants are plotted with 95% confidence intervals: Open symbols, continuous light condition; closed symbols, dark condition.

in the dark at 48 h after the start of measurements. The difference in CO<sub>2</sub> concentration between the light and the dark was significant at 5% level. Thus, lighting to leaves appeared to be responsible for gas exchange in roots as well as water uptake rate, and it was suggested that stomatal movement is related to gas exchange in roots.

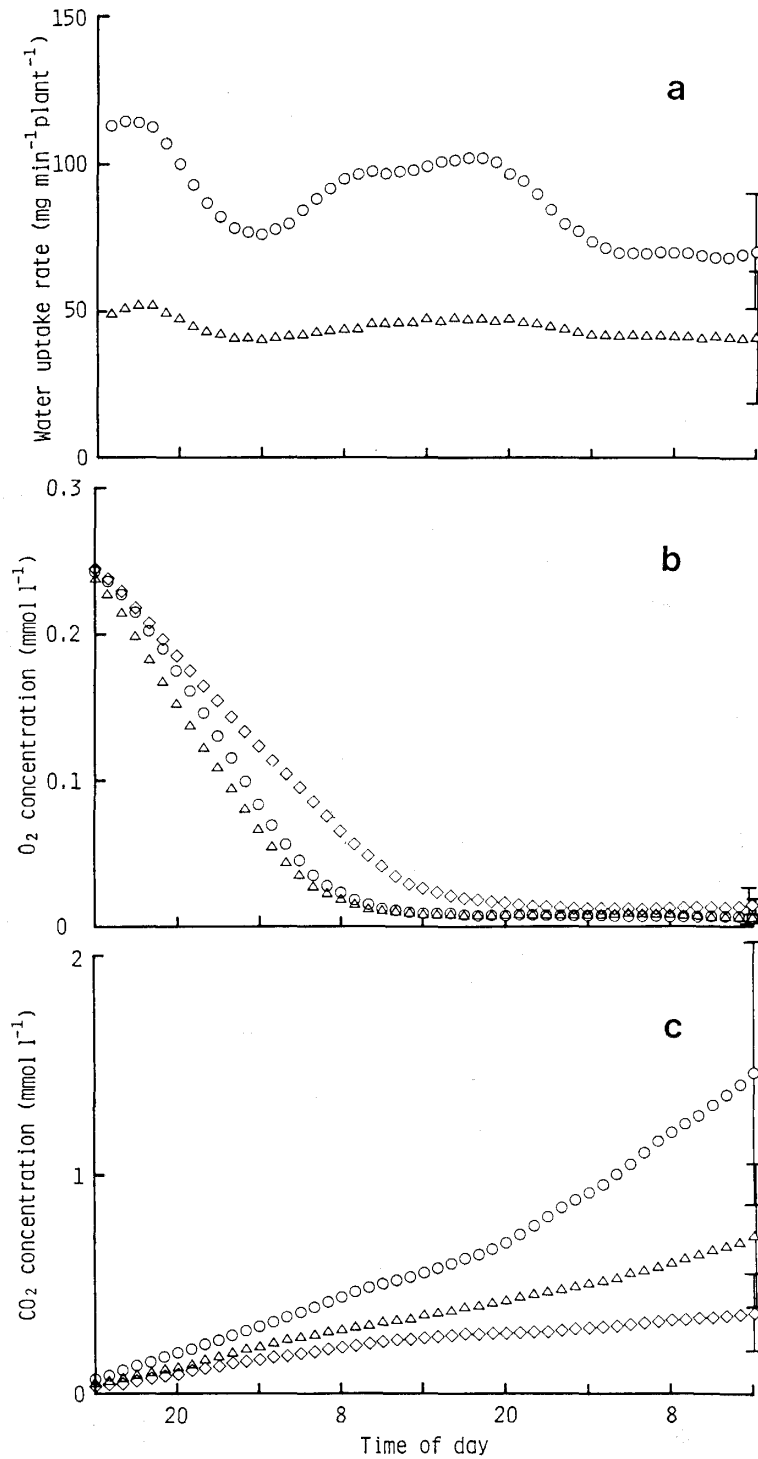


Fig. 5. Time course patterns of water uptake rate (a), dissolved O<sub>2</sub> (b) and dissolved CO<sub>2</sub> (c) concentrations in hydroponics under continuous light condition, where means of measured values in 3 plants are plotted with 95% confidence intervals at 48 h after the start of measurements: ○, untreated plants; △, plants treated with ABA and microcrystalline wax; ◇, detached roots.



For inhibition of transpiration and the gas exchange in leaves, abscisic acid ( $10^{-3}$  mol  $l^{-1}$ ) and microcrystalline wax were applied to leaves. Furthermore, detached root system was used to examine influence of leaves on the gas exchange in roots. Figure 5 shows water uptake rate (a),  $O_2$  (b) and  $CO_2$  (c) concentrations in hydroponics where plants were continuously lighted. Water uptake rate in plants treated with ABA and the wax was half as much as that in untreated plants. Decrease rate of  $O_2$  in detached roots was lower than that in untreated whole plants. The difference in  $O_2$  decrease was very small between treated and untreated plants. On the other hand, increase rate of  $CO_2$  in the treated plants was lower than that in untreated plants. At 48 h after the start of measurements, difference in  $CO_2$  concentration between the treated and untreated plants was significant at 5% level. Thus, increase in  $CO_2$  concentration was inhibited in plants treated with ABA and the wax and in detached roots.

Water uptake rate, decrease rate of  $O_2$ , and increase rate of  $CO_2$  are listed in Table 1. In the light, water uptake rate in plants treated with ABA and the wax was lower than that in untreated plants. Although decrease rate of  $O_2$  in the treated plants was similar to that in untreated plants, increase rate of  $CO_2$  was lower in the treated plant as compared with untreated plants. On the other hand, water uptake rate in the dark was lower than that in the light, and decrease rate of  $O_2$  and increase rate of  $CO_2$  in the dark were similar to those in detached roots. Thus, decrease in  $O_2$  concentration was not necessarily associated with water uptake, but increase in  $CO_2$  concentration was related to water uptake.

Water uptake in roots is caused by leaf transpiration, and gas exchange is caused through stomata opening in transpiring leaf (2, 22, 26). Internal  $O_2$  transport from the leaf to roots has been reported in several plant species (5, 21). In this experiment, increase rate of dissolved  $CO_2$  concentration was higher in the case of higher water uptake rate, and the  $CO_2$  concentration continued to increase even in the  $O_2$  deficit solution. From these facts, it is possible that  $O_2$  transported from leaf to root system may be used for root respiration.

Table 1. Water uptake rate, decrease rate of dissolved  $O_2$  concentration, and increase rate of dissolved  $CO_2$  concentration, where means of measured values in 3 plants with respective 95% confidence limits are listed

Treatments	Light condition	Water uptake rate (mg min <sup>-1</sup> plant <sup>-1</sup> )	$O_2$ decrease rate ( $\mu$ mol l <sup>-1</sup> min <sup>-1</sup> )	$CO_2$ increase rate ( $\mu$ mol l <sup>-1</sup> min <sup>-1</sup> )
Applying ABA + micro-crystalline wax to leaves	Light	44.1* ( $\pm 20.0$ )	0.24 ( $\pm 0.10$ )	0.24* ( $\pm 0.10$ )
Detaching roots	—	0.0* ( $\pm 0.0$ )	0.16 ( $\pm 0.08$ )	0.12* ( $\pm 0.05$ )
Untreated	Dark	34.2* ( $\pm 26.7$ )	0.14 ( $\pm 0.11$ )	0.12* ( $\pm 0.01$ )
Untreated	Light	87.2 ( $\pm 6.6$ )	0.23 ( $\pm 0.11$ )	0.49 ( $\pm 0.22$ )

\*; significant difference at 5% level in comparison with untreated plant in the light.

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