### 九州大学学術情報リポジトリ Kyushu University Institutional Repository

## Complete Genome Sequences of Ralstonia solanacearum Strains Isolated from Zingiberaceae Plants in Japan

Iiyama, Kazuhiro

Laboratory of Plant Pathology, Faculty of Agriculture, Graduate School, Kyushu University

Kodama, Sawa

Laboratory of Plant Pathology, Faculty of Agriculture, Graduate School, Kyushu University

Kusakabe, Honoka

Laboratory of Plant Pathology, Faculty of Agriculture, Graduate School, Kyushu University

Sakai, Yoriko

Institute for Agro-Environmental Sciences, National Agriculture and Food Research Organization (NARO)

他

https://hdl.handle.net/2324/4774264

出版情報: Microbiology Resource Announcements. 10 (4), pp.e01303-20-, 2021-01-28. American

Society for Microbiology

バージョン:

権利関係: Creative Commons Attribution International









# Complete Genome Sequences of *Ralstonia solanacearum* Strains Isolated from Zingiberaceae Plants in Japan

© Kazuhiro liyama, a Sawa Kodama, a Honoka Kusakabe, a Yoriko Sakai, b Mitsuo Horita, b Kazutaka Yano, c Htet Wai Wai Kyaw, a Kenichi Tsuchiya, a Naruto Furuya

<sup>a</sup>Laboratory of Plant Pathology, Faculty of Agriculture, Graduate School, Kyushu University, Fukuoka, Japan <sup>b</sup>Institute for Agro-Environmental Sciences, National Agriculture and Food Research Organization (NARO), Ibaraki, Japan <sup>c</sup>Kochi Agricultural Research Center, Kochi, Japan

**ABSTRACT** Here, we report the complete genome sequences of three *Ralstonia solanacearum* strains isolated from Zingiberaceae plants in Japan. The total genome sizes of these strains ranged from 5.87 to 6.05 Mb. Strains MAFF 211472, MAFF 211479, and MAFF 311693 each carried one chromosome and one megaplasmid. MAFF 311693 contained an additional 71.9-kb plasmid.

alstonia solanacearum is a soilborne plant-pathogenic bacterium. In Japan, R. solanacearum strains infecting Curcuma alismatifolia (Zingiberaceae) were first reported in 1995 (1). Thereafter, the bacterium was isolated from other Zingiberaceae plants, including ginger (Zingiber officinale), myoga (Z. mioga), turmeric (C. longa), wild turmeric (C. aromatica), and zedoary (C. zedoaria) (2–4). Repetitive sequence-based PCR (rep-PCR) assays identified invasive strains from foreign countries (5).

Since not all *R. solanacearum* strains are pathogenic to Zingiberaceae plants (6), the genomes of bacterial isolates from Zingiberaceae plants were sequenced to determine their pathological potential. The *R. solanacearum* strains (phylotype I, race 4, biovar 4) used in this study were MAFF 211472, MAFF 211479, and MAFF 311693. MAFF 211472 was isolated from ginger in Nakamura, Kochi Prefecture, Japan, in 1997. MAFF 211479 was isolated from ginger in Kahoku, Kochi Prefecture, Japan, in 1997. MAFF 311693 was isolated from wild turmeric in Nago, Okinawa Prefecture, Japan, in 2016. The strains were preserved as freeze-dried cultures at Genebank in the National Agriculture and Food Research Organization (NARO, Tsukuba, Ibaraki, Japan). The freeze-dried cultures were rehydrated in the Laboratory of Plant Pathology (Kyushu University). The cultures were stored as freeze-dried cells and 20% glycerol suspensions at  $-70^{\circ}$ C for long-term preservation and routine work, respectively.

The glycerol suspension was streaked onto Casamino Acid-peptone-glucose (CPG) medium (7), and the single colony isolation was repeated with CPG medium two to three times for genome extraction. Single colonies of *R. solanacearum* strains were cultured in CPG broth overnight at 30°C. Genomic DNA was extracted using the cetyltrimethylammonium bromide protocol (8). The extracted DNA was used on long-read and short-read sequencing platforms to generate hybrid assemblies using single-molecule real-time (SMRT) (PacBio, CA, USA) and Illumina (San Diego, CA, USA) sequencing, respectively. Library preparation and sequencing were performed by Novogene Co., Ltd. (Beijing, China).

For PacBio sequencing, SMRTbell libraries were generated using sheared template DNA, and SMRT sequencing was carried out on a PacBio Sequel platform, creating 178,820 (MAFF 211472), 137,360 (MAFF 211479), and 187,585 (MAFF 311693) subreads with average lengths of 9,231 bp, 9,513 bp, and 9,912 bp, respectively; the  $N_{50}$  values were 10,416 bp, 10,783 bp, and 11,237 bp, respectively.

Citation liyama K, Kodama S, Kusakabe H, Sakai Y, Horita M, Yano K, Kyaw HWW, Tsuchiya K, Furuya N. 2021. Complete genome sequences of *Ralstonia solanacearum* strains isolated from Zingiberaceae plants in Japan. Microbiol Resour Announc 10:e01303-20. https://doi.org/10.1128/MRA.01303-20.

Editor David A. Baltrus, University of Arizona

Copyright © 2021 liyama et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Kazuhiro liyama, iiyama@grt.kyushu-u.ac.jp.

Received 24 November 2020 Accepted 7 January 2021 Published 28 January 2021

TABLE 1 Accession numbers, assembly metrics, and annotated features of the sequenced Ralstonia solanacearum strains

|                     | GenBank       | Genome assembly |                | No. of:          |           |           |          |       |         |        |
|---------------------|---------------|-----------------|----------------|------------------|-----------|-----------|----------|-------|---------|--------|
| Strain <sup>a</sup> | accession no. | size (bp)       | GC content (%) | CDS <sup>b</sup> | 16S rRNAs | 23S rRNAs | 5S rRNAs | tRNAs | tmRNAsc | CRISPR |
| MAFF 211472         |               |                 |                |                  |           |           |          |       |         |        |
| Chromosome          | AP024157      | 3,914,553       | 66.7           | 3,673            | 3         | 3         | 3        | 59    | 1       | 1      |
| Megaplasmid         | AP024158      | 2,139,380       | 66.6           | 1,726            | 1         | 1         | 1        | 7     | 0       | 0      |
| Total               |               | 6,053,933       | 66.7           | 5,399            | 4         | 4         | 4        | 66    | 1       | 1      |
| MAFF 211479         |               |                 |                |                  |           |           |          |       |         |        |
| Chromosome          | AP024159      | 3,774,488       | 66.8           | 3,514            | 3         | 3         | 3        | 63    | 1       | 0      |
| Megaplasmid         | AP024160      | 2,131,116       | 66.8           | 1,738            | 1         | 1         | 1        | 8     | 0       | 0      |
| Total               |               | 5,905,604       | 66.8           | 5,252            | 4         | 4         | 4        | 71    | 1       | 0      |
| MAFF 311693         |               |                 |                |                  |           |           |          |       |         |        |
| Chromosome          | AP024161      | 3,680,093       | 67.1           | 3,441            | 3         | 3         | 3        | 59    | 1       | 0      |
| Megaplasmid         | AP024162      | 2,117,975       | 66.7           | 1,741            | 1         | 1         | 1        | 8     | 0       | 0      |
| Plasmid             | AP024163      | 71,852          | 61.4           | 89               | 0         | 0         | 0        | 0     | 0       | 0      |
| Total               |               | 5,869,920       | 66.9           | 5,271            | 4         | 4         | 4        | 67    | 1       | 0      |

Detailed classification: MAFF 211472, phylotype I, sequevar 16, race 4, biovar 4; MAFF 211479 and MAFF 311693, phylotype I, sequevar 30, race 4, biovar 4.

For Illumina sequencing, we generated paired-end libraries (fragment size, ~350 bp) using the NEBNext Ultra DNA library prep kit. Libraries were sequenced on an Illumina HiSeq 4000 sequencing system. After filtering with Trimmomatic v.0.32 (9), 5.24, 4.71, and 4.95 million 150-bp paired-end reads were used to generate hybrid assemblies for MAFF 211472, MAFF 211479, and MAFF 311693, respectively. Hybrid de novo assemblies were generated using Unicycler v.0.4.7 (10) with PacBio long reads and Illumina short reads. Default parameters were used for all software. The assemblies of MAFF 211472 and MAFF 211479 had total lengths of 6,053,933 bp with 66.7% GC content and 5,905,604 bp with 66.8% GC content, respectively. Both comprised two circular contigs, a chromosome and a megaplasmid. In MAFF 311693, the genome size was 5,869,920 bp with 66.9% GC content, and three circular contigs were generated, indicating that this strain carries an additional 71,852-bp plasmid.

The assemblies were annotated using DFAST (11), and the results are summarized in Table 1.

Data availability. The complete sequences of the R. solanacearum strains have been deposited in the DNA Data Bank of Japan (DDBJ) under BioProject accession number PRJDB9507. The BioSample accession numbers are SAMD00256261, SAMD00256262, and SAMD00256263. The DRA accession numbers are DRX245399/DRX245396, DRX245400/ DRX245397, and DRX245401/DRX245398. The GenBank assembly accession numbers are GCA\_015698345.1, GCA\_015698365.1, and GCA\_015698385.1. The genome sequence accession numbers are listed in Table 1. The raw sequencing reads (PacBio and Illumina) were deposited in the DDBJ Sequence Read Archive database under accession number DRA011112.

#### **ACKNOWLEDGMENTS**

This research was supported by grants from a project of the NARO Bio-oriented Technology Research Advancement Institution (Research Program on Development of Innovative Technology).

We thank Novogene Co., Ltd. for sequencing. We thank Editage for English language editing.

### **REFERENCES**

- 1. Morita Y, Yano K, Tsuchiya K, Kawada Y. 1996. Bacterial wilt of Curcuma alismatifolia caused by Pseudomonas solanacearum. Proc Assoc Plant Protec Shikoku 31:1-6. (In Japanese.)
- 2. Tsuchiya K, Yano K, Horita M, Morita Y, Kawada Y, D'Ursel CM. 1999. Occurrence of bacterial wilt of ginger in Japan, Jpn J Phytopathol 65:363. (In Japanese.) https://doi.org/10.3186/jjphytopath.65.321.

<sup>&</sup>lt;sup>b</sup> CDS, coding DNA sequences.

<sup>&</sup>lt;sup>c</sup> tmRNAs, transfer-messenger RNAs.

Downloaded from https://journals.asm.org/journal/mra on 07 March 2022 by 133.5.78.159.

- 3. Yano K, Kawada Y, Tsuchiya K, Horita M. 2005. First report of bacterial wilt of mioga (Zingiber mioga) caused by Ralstonia solanacearum in Japan. Jpn J Phytopathol 71:179–182. (In Japanese.) https://doi.org/10.3186/ iiphytopath.71.179.
- 4. Ajitomi A, Inoue Y, Horita M, Nakaho K. 2015. Bacterial wilt of three Curcuma species, C. longa (turmeric), C. aromatica (wild turmeric) and C. zedoaria (zedoary) caused by Ralstonia solanacearum in Japan. J Gen Plant Pathol 81:315-319. https://doi.org/10.1007/s10327-015 -0596-9.
- 5. Tsuchiya K, Yano K, Horita M, Morita Y, Kawada Y, D'Ursel CM. 2005. Occurrence and epidemic adaptation of new strains of Ralstonia solanacearum associated with Zingiberaceae plants under agro-ecosystem in Japan, p 463-469. In Allen C, Prior P, Hayward AC (ed), Bacterial wilt disease and the Ralstonia solanacearum species complex. American Phytopathological Society, St. Paul, MN.
- Yano K, Kawada Y, Horita M, Hikichi Y, Tsuchiya K. 2011. Phylogenetic discrimination and host ranges of Ralstonia solanacearum isolates from

- Zingiberaceae plants. Bull Jpn Soc Plant Pathol 77:88-95. (In Japanese.) https://doi.org/10.3186/jjphytopath.77.88.
- 7. Kelman A. 1954. The relationship of pathogenicity of *Pseudomonas sola*nacearum to colony appearance on a tetrazolium medium. Phytopathology 44:693-695.
- 8. Wilson K. 2001. Preparation of genomic DNA from bacteria. Curr Protoc Mol Biol 56:2.4.1-2.4.5. https://doi.org/10.1002/0471142727.mb0204s56.
- 9. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- 10. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- 11. Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34:1037-1039. https://doi.org/10.1093/bioinformatics/btx713.