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The principle and the potential approach to ROS-dependent cytotoxicity by

non-pharmaceutical therapies: optimal use of medical gases with antioxidant

properties

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#### **Abstract**

Regulation of cellular redox balances is important for the homeostasis of human health. Thus, many important human diseases, such as inflammation, diabetes, glaucoma, cancers, ischemia and neurodegenerative diseases, have been investigated in the field of reactive oxygen species (ROS) and oxidative stress. To overcome the harmful effect of oxidative stress and ROS, one can directly eliminate them by medical gases such as carbon monoxide (CO), hydrogen sulphide (H<sub>2</sub>S), and molecular hydrogen (H<sub>2</sub>), or one can induce ROS-resistant proteins and antioxidant enzymes to antagonize oxidative stresses. This article reviews the molecular mechanisms how these medical gasses work as antioxidants, and how ROS resistant proteins are produced in the physiological context. Targeted therapeutic modalities to scavenge or prevent ROS might be applied in the prevention and treatment of ROS-related diseases in the near future.

#### Introduction

Reactive oxygen species (ROS) play an important role in aging and life-style dependent diseases such as cancer, stroke and diabetes. Recently hydrogen gas, C60 fullerene, and functional nano-particles were acknowledged as selective anti-oxidants with tremendous application potential [1, 2]. In addition, electro-magnetic waves including different spectrum of light, modulate ROS production for the treatment of several diseases. Further, the most attractive new concept in the field of oxidative stress is the artificial production of ROS-resistant proteins, which was found in long-lived (28 years) naked mole rats [3], suggesting how ROS-related redox signaling could be modified for medical purposes [4]. In the following sections, how ROS are produced, how ROS affect cellular functions, and how ROS are eliminated in living cells will be discussed. Targeting pathways of redox signaling, by ROS in living cells is one of the most rational options to overcome or prevent aging-related diseases.

In order to elucidate ROS-related diseases and problems, three sections will be shown as followings,

- I. How ROS are generated in living cells.
- II. How ROS are removed from living cells.
- III. How ROS resistant proteins are made artificially.

In part I, the detail of the mechanism how light and radiation can generate ROS are discussed. Also, we discuss several metal ions which are involved in several steps of redox signaling in the generation of ROS from oxygen molecule. In part II, we propose several medical gases, including carbon monoxide (CO), hydrogen sulphide (H<sub>2</sub>S), and molecular hydrogen (H<sub>2</sub>), to counteract ROS. Of note, these gases share some common

factors and signaling molecules in ROS buffering, such as superoxide dismutase (SOD), heme oxidase-1 (HO-1). At first, we discuss the role of key antioxidant enzymes such as SOD, catalase, and glutathione peroxidase in this process. In the part III, ROS-resistant proteins and one of the key redox signaling cascade in living cells will be discussed.

#### I. Production of ROS in living tissue by light and radiation

ROS and a related term "oxidative stress" were known as a paradigm to describe as an imbalance between oxidants and antioxidants leading to a disruption of redox signaling, resulting in molecular and cellular damages [5]. Accordingly, generation of ROS may contribute to a condition of "oxidative stress" [6]. ROS may also indicate the generic name of singlet oxygen ( ${}^{1}O_{2}$ ), super oxide ( ${}^{4}O_{2}$ ), hydrogen peroxide ( ${}^{4}O_{2}$ ) and hydroxyl radicals ( ${}^{4}O_{1}$ ).

When a photosensitizer (PS), either exogenous or endogenous, is exposed to light, PS will absorb the specific wave length of light and will be excited to singlet state from the ground state. Due to the energy flow, the singlet state will return to the ground state shortly after heat production. Alternatively, it may shift to triplet state, which is more slowly than to singlet state, via the intersystem crossing [7]. The triplet state is more

stable than the singlet state. Eventually, the energy stored at the triple state can decay gradually to ground state via two independent pathways; type I reaction and type II reaction (Fig. 1).

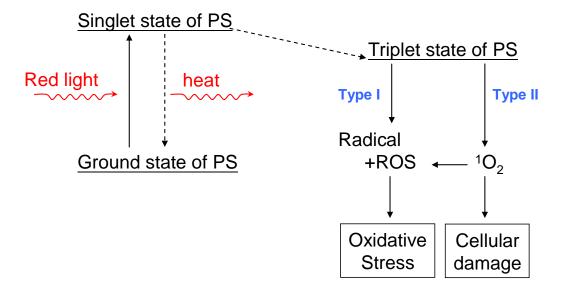


Fig. 1 Excitation of photosensitizer (PS) by red light to develop ROS and singlet oxygen

Type I reaction generates free radicals, which make oxidative stress together with ROS derived from singlet oxygen produced by Type II reaction [8]. Type II reaction is predominant when cells are irradiated with red light [9, 10]. Oxidative stress can result from several other biochemical reactions, including nitrosation [10]. We can classify effect of oxidative stress as a result of modification, such as

nitrosation [11], carbonylation [12], disulfide bond formation [13] and glutathiolation [14]. In addition to those, the effect of oxidative stress on biochemical and physiological effect for cellular dysfunction is collapse of stem cell regulation [15], promotion of cyclophilin A from vascular smooth muscle, activation of transient receptor potential (TRP) channel [16], involvement in amyotrophic lateral sclerosis (ALS) [17]. Also effect of oxidative stress on biological function related to apoptosis is induction of carcinogenesis, activation of MAP kinase, pathway [19], externalization of phosphatidylserine (PS) [20, 21]. Suppression of caspase activity [22], DNA damage [23], induction of mitochondrial membrane permeation [24]

# I-1. Photosensitizer as ROS generator

As depicted in Fig. 1, when photo-sensitizer is illuminated by light, the sensitizer can shift to high density triplet state if it has heavy metal ion. This triple state relaxed to lower state with the emission of phosphorescence and making radicals. Generally it is called photochemicals. The most popular reaction is cis-trans isomerization, which is caused when light is illuminated to C-C double bond. This reaction has been established in the retinal rhodopsin [25], β-carotenoide [26], chlorophyl and hemoglobin [27, 28].

Among them, hemoglobin, especially heme (Fe), is an important component of cytochrome. Heme is also required for the synthesis of enzymes other than cytochromes, as well as myoglobin and hemoglobin. Eventually, iron can play an important role in ROS development and in the increase of cytotoxicity, both of which is shown in a cell culture study where cells were incubated with iron either before or after light irradiation [29]. As expected, iron given before light irradiation resulted in less lethality but adding iron after irradiation increased lethality [30]. This is because chelating iron before light illumination results in less photo-sensitizer accumulation [31].

Interaction of human cells with ultra fine (nano-) particles can cause the formation of ROS, including super oxide, hydrogen peroxide, hydroxyl radicals, and singlet oxygen [32]. While some types of nano- particle made by carbon, metal and metal oxide act as PS, causing production of both singlet oxygen and super oxide from ground state of oxygen gas molecules under the influence of light, the photochemical reactivity of nanoparticles is regarded of little relevance for the interaction with tissues that are not exposed to daylight. Also transition metal ions may be released in the vicinity of nanomaterials, partly derived from particle impurities, catalyzing Fenton type reactions [33, 34]. In addition, the physical interactions of

particles with subcellular structures involved in the catalysis of redox reactions may modulate generation of ROS. Hypothetical targets of the nanoparticles might involve plasma membrane, mitochondria and endoplasmic reticulum. An interaction of nanoparticle with mitochondrial structure may affect directly to electron flow and leakage from the inner membrane of mitochondria. The interaction of nanoparticle with endoplasmic reticulum (ER) may cause dysregulation of intracellular Ca<sup>2+</sup> ion level, which in terms activates Ca<sup>2+</sup>/calmodulin-dependent protein kinases II (CaMKII) and eNOS and nNOS, resulting in production of nitric oxide.

#### I-2. Effect of UV light for redox signaling

UV radiation is divided into three defined regions of wavelength, including UVA (320-400 nm), UVB (280-320 nm) and UVC (less than 280 nm). Among them UVA penetrate skin most deeply and reach to subepidermal layers [35, 36]. It is well known that UVA is attributed to photosensitized generation of singlet oxygen. The singlet oxygen represents an electrically excited state of O<sub>2</sub>, singlet oxygen [37, 38]. Although it is not a radical, singlet oxygen is more reactive than ground state of molecular oxygen due to the "spin-forbidden" rule. Example of such UV photosensitizer includesporphyrins, flavins and certain quinines [39, 40]. This principle of the sensitized

generation of singlet oxygen is exploited medically in photodynamic therapy (PDT), where the sensitizers are administrated and accumulated in target areas prior to irradiation in order to kill the respective cells [41]. How important the singlet oxygen is? Singlet oxygen is electrically excited state of dioxygen, and it has higher energy than the triplet state of O<sub>2</sub>, which is generally found in air. The singlet oxygen has high affinity with double bond of fatty acid. As a consequence, it has higher reactivity than triplet O<sub>2</sub>. The singlet oxygen can also react with a wide range of biomolecules, including DNA, proteins and lipids.

Higher energy UVB and UVC are not available as UVA for clinical use. UVB is effective to decomposition of ozone (O<sub>3</sub>), while UVC is effective for formation of ozone from atomic and molecular oxygen. Without doubt, ozone is another highly reactive form of oxygen and plays a role as oxidative stressor but is probably not formed in human body. Some recent reports claimed that ozone is formed in neutrophils [42-44], but no reports were followed subsequently. Lipid peroxidation is also an important factor to cause damage to living cells. When UV is irradiated to lipid molecules, hydrogen molecule in double bond in LH (unsaturated fatty acids) is cut off and L• (lipid radical) is formed. When O<sub>2</sub> molecules are exist nearby, L• will be LOO•, which will make peroxide radicals (LOOH), activating membrane enzyme such as PLC and

PLA2 [45-48].

#### I-3. ROS Production by radiation

The major component of living cell is  $H_2O$ . When low energy radiation enters into the living cell, G values obtained by low LET radiation is listed as shown in Table 1. LET is defined as energy loss/ cm path length in the water. Gamma ray and  $\beta$ -ray has low LET, while  $\alpha$ -ray, proton and heavy ion has high LET [49-51].

<b>Table 1</b> G values of 1st productions from H <sub>2</sub> O by radiation	
Hydroxy radical (• OH)	2.8
Hydrated electron (e <sub>aq</sub> )	2.8
Hydrogen (H <sub>2</sub> )	0.45
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )	0.75

In Table 1, G value is defined as number of product when material absorbed 100 eV of radiation.

As shown in Table 1,  $\bar{e_{aq}}$  is very specific for ROS production by radiation. The  $\bar{e_{aq}}$  is formed by;

$$e- + nH_2O = \bar{e_{aq}}$$
 (hydrated electron)

and,

$$H_2O \rightarrow H_2O \bullet \rightarrow H^- + \bullet OH$$

When  $O_2$  is present,  $H^{-}$  and  $e_{aq}$  make  $\overline{\bullet}O_2$  (super oxide) and hydro-per-oxyradical

(HO₂•). Thus radiation can produce super oxide very effectively in the living tissue.
 This is called "oxygen effect". Accordingly, enhancement of radiation effect with oxygen is defined as,

OER (Oxygen enhancement ratio) = Effect without  $O_2$ / Effect with  $O_2$  Usually, observed value of OER is about 2.5 ~ 3.0.

Subcellular localization of ROS production is crucial in the biological processes. It is well known that mitochondria generate ROS largely within the cell. If we introduce artificial lipophilic photosensitizer [52], such as rose bengal, riboflavin, methylene blue and various porphrins, into the cell, it will be preferentially accumulated in the lysosomes [53], mitochondrial membrane [54], Golgi apparatus [55], endo-plasmic reticulum, and plasma membrane.

#### I-4. Redox signaling and metal ions in the living tissue

Oxygen molecules present in living tissues will become super oxide after several steps of redox signaling eventually. The super oxide will be eventually transformed to  $H_2O$  via  $H_2O_2$  and  $\bullet OH$ . This signaling pathway of ROS production is shown in Fig. 2.

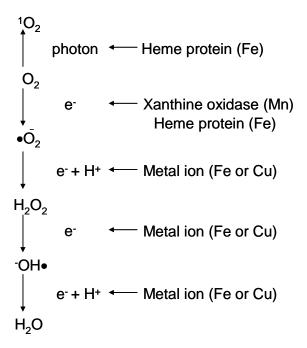


Fig. 2 Redox signaling pathway in the cells in the presence of metal ions.

There are a lot of metals or metallic ions in the human body. In the order of amount in the human body is; Fe > Zn> Cu> Mn> Se> As> Cr> Mo> Co > V> Ni complexes. Many of these complexes contain transition metal ions and are redox active such as cytochrome c. Others take advantage of ligand exchange properties of metal complex (e.g. hemoglobin, myoglobin). For ROS production, Mn (manganese), Fe (ion) and Cu (copper) are major [56]. For ROS removal, Zn (zinc) is important in addition to Mn, Fe and Cu for SOD activation [57]. Se (selenium) is also important for elimination of  $H_2O_2$  by activation of peroxidase with Fe and Mn [58].

Here we discuss As (arsenic) in this paragraph especially, because it is not well understand in the redox signaling. Chronic intake of As was associated with increased risk of cancer, diabetes, developmental vascular diseases and reproduction, all which may result from ROS imbalances. Furthermore, direct evidence that As can cause oxidative damages to DNA and protein was reported recently [59, 60]. Dermatologic toxicities due to As exposure are well documented and include arsenical keratosis, carcinoma in situ, invasive skin cancers, and pigmentary abnormalities [61-63]. In addition to general ROS, nitric oxide (•NO) and peroxynitrite (ONOO) are referred to as reactive nitrogen species (RNS). Exposure of human keratinocytes to As leads to formation of •O2 and H<sub>2</sub>O<sub>2</sub> [63]. Different from other metal ions listed above, As is a semi-metal or a metalloid with two biologically important chemical state, As (V) and As (III), as the oxyacids  $(H_3AsO_4)$  or arsenic trioxide  $(As_2O_3)$ . The other aspect of As chemistry relevant to biological activity is that As (III) behaves as a soft metal ion, forming strong bonds with thiolates of cysteine residues [64] and the imidazolium nitrogen of histidine residues [65]. It is also not clearly determined yet how As enters into mammalian cells. Recently we have discovered that As selectively reduced IP<sub>3</sub>-induced Ca release (unpublished data). It is reasonable to assume that As can enter into the cell via aquaporin channel and react with cystein residues, although cellular

damage by ROS was expected.

#### II. How ROS are removed from living cells: the elimination of ROS

Next, we discuss how to eliminate ROS in the living tissues by several kinds of enzyme and coenzyme. We will also discuss the minerals that act as activators of the enzymes.

#### II-1. Super oxide dismutase for superoxide anion radicals

As described so far ROS is collective term of superoxide radical anion ( $\bullet$ O<sup>2</sup>-), the hydroxyl radical (OH $\bullet$ ) and peroxyl radicals (ROO $\bullet$ ), the most important molecule in the redox signaling, is H<sub>2</sub>O<sub>2</sub> as shown in Fig. 2. The fate of most ROS is dismutation to H<sub>2</sub>O<sub>2</sub> by both enzymatic and non-enzymatic reactions. Thus it is not surprising that proteins known to react with super oxide are guanylate cyclase [66], ribonucleotide reductase [67], phosphor-protein phosphatase 2B [68].

Catalyzed by SOD, super oxide radical anion makes H<sub>2</sub>O<sub>2</sub>.

$$\bullet O_2^- + \bullet O_2^- = H_2O_2 + O_2$$

This is an important reaction to protect against cell damages, because half life of super oxide is very short. In humans, SOD occurs in three forms; two of them, SOD1 and SOD3, contain Zn and Cu, while SOD2 contains Mn. SOD1 is present in cytosol, SOD2

is in mitochondria and SOD3 exist outside the cell. However,  $H_2O_2$  is also toxic, because  $H_2O_2$  act as a redox signal eitherthrough redox coupling directly with amino acid residues or indirectly through cellular mediators, such as the thioredoxin and glutathione couples [69]. In some cases, the products of these reactions inhibit the protein functional [70, 71]. Therefore  $H_2O_2$  must be further decomposed by catalase as;

$$2H_2O_2 \rightarrow 2H_2O + O_2$$

and by glutathione peroxidase as;

$$H_2O_2 + 2GSH \rightarrow 2H_2O + GSSG$$

, where GSH is oxidized to its disulfide GSSG in the reaction.

### II-2. Reducing ROS by medical gases

The diverse physiological actions of the "medical gases" such as CO, NO, and  $H_2S$ , as well as  $H_2$ , have drown attentions. These gases play important roles as cellular signaling molecules. The difficulty in measuring local gas concentrations obscures detailed mechanisms whereby gases exert their actions. Although many questions remain unanswered, some potential mechanisms are summarized as below.

#### Carbon Monoxide (CO)

CO is a diatomic molecule, and is soluble in aqueous media and organic solvents [72]. CO is relatively stable and has no unpaired electron whereas nitric oxide (•NO)

has one unpaired electron. Not only from exogenous environmental exposure but also from endogenous production during the heme metabolism are major source of CO from primitive prokaryotes to humans [72, 73]. Recent studies have revealed that CO serves as an intrinsic signaling molecule and shows anti-inflammatory and anti-apoptotic effects (Fig. 3). Therapeutic effect of CO is observed in several pathological disease models such as sepsis [74, 75], asthma [76], oxidative-lung injury [77, 78], bleomycin-induced pulmonary fibrosis [79], myocardial infarction [80, 81], hypertension [82-85], atherosclerosis [86, 87], ischemia/reperfusion [88-91], and liver/lung transplantation [92, 93].

Endogenous production of CO is highly associated with heme oxygenases (HO) activity which induces enzymatic degradation of heme. HO breaks the alpha-methene carbon bond of the porphyrin ring with the help from NADPH and molecular O<sub>2</sub> in a reaction that releases equimolar amount of biliverdin, iron, and CO [94, 95]. Of two major isoforms HO-1 and HO-2, HO-1 is inducible whereas HO-2 is constitutively expressed [96, 97]. In an oxidative stress condition, HO-1, which is also named as HSP 32, is induced by several transcription factors including NF-E2-related factor-2 (Nrf2), which binds antioxidant response elements (ARE) [98, 99]. Other transcription elements are also associated with the transcriptional induction of HO-1, including cAMP

response element, TPA response element, and Maf binding site, which are similar to stress-response element [100]. The expression level of HO-1 itself is also induced by CO exposure via nitric oxide (•NO) in bovine pulmonary artery endothelial cells culture [101].

HO can catalyze the first and rate-limiting step in the oxidative degradation of heme to form the open-chain tetrapyrrole biliverdin-IXa (BV) [102, 103]. BV is subsequently converted to bilirubin-IXa (BR) via an NAD(P)H-dependent biliverdin reductases [104]. Both BV and BR can act as potent antioxidants [105-107]. Both BV and BR serve as physiological antioxidants. BR acts as an electron donor, and thereby inhibits lipid peroxidation by chain breaking antioxidant effect [108]. On the other hands, BV reacts with peroxyl radicals, and form carbon-centered radical (or radicals) by the addition of an LOO- to the pigment (LOO• + BV → LOO-BV•) [108]. This radical-trapping activity of BV is relatively higher than BR.

Endogenous CO, which is produced by heme degradation, induces ROS-dependent signaling transduction in the antioxidant defense enzyme systems, including the mitochondrial superoxide dismutase (SOD2) and HO-1 itself [101]. The source of ROS, especially super oxide, is mainly mitochondrial complex III [109]. CO binds at cytochrome c oxidase, which leads to the slowing the transfer rate of electrons to

molecular O<sub>2</sub>. This delay in electron transfer make the upstream carriers to more reduced state and fill the Q cycle electron pool, leading to increases in O<sub>2</sub> concentration. MnSOD converts •O<sub>2</sub> to H<sub>2</sub>O<sub>2</sub>, and CO-mediated mitochondrial H<sub>2</sub>O<sub>2</sub> serves as a signaling molecule to mitochondrial biogenesis. The key transcription factor of mitochondrial biogenesis is nuclear respiratory factor-1 (NRF-1), which is functionally regulated by Akt [110]. Mitochondrial H<sub>2</sub>O<sub>2</sub> inactivates counter-regulatory phosphatases such as PTEN and PTP1B, which leads to PI3K-induced unopposed activity of Akt [111-114]. NRF-1 binds to mitochondrial transcription factor-A promoter (*Tfam* promoter). Transcriptional induction of Tfam is essential for mitochondrial DNA transcription and replication [95, 115]. The mitochondrial biogenesis can maintain the mitochondrial number, volume density, and function, which allows the cells to show their adaptive response against mitochondrial damages.

The anti-inflammatory and anti-apoptotic effects of CO are mediated by p38 mitogen-activated protein kinase (MAPK) signaling, which acts in response to physical and chemical stress inducers including oxidative stress, UV, ischemia, and pro-inflammatory cytokines [116]. Activation of p38 also mediates the induction of heat shock protein 70 via its transcriptional factor, heat shock factor-1, leading to the cytoprotective effects [117]. (See Fig. 3)

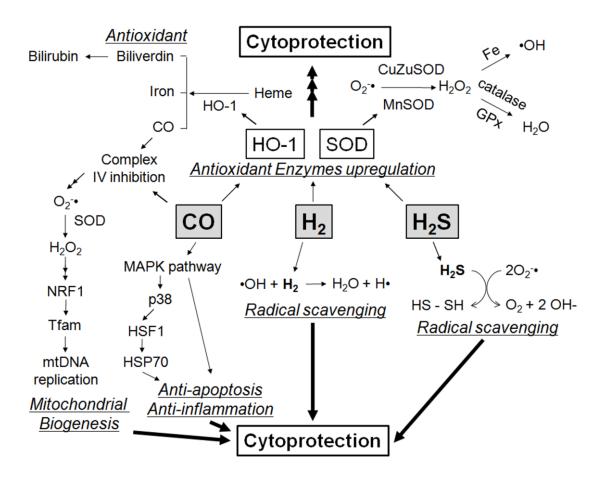


Fig. 3 Schematic mechanisms for cytoprotection induced by medical gases

#### *Hydrogen sulphide* (*H*<sub>2</sub>*S*)

Hydrogen sulphide  $(H_2S)$  is a flammable, water-soluble gas with a smell of rotten eggs. Although it is known as a toxic gas and as an environmental hazard, the role of  $H_2S$  in tissue has gained an interest with its cytoprotective effects. $H_2S$  can be

synthesized by endogenous two enzymes, cystathionine β-synthase (CBS), and cystathionine γ-lyase (CSE). These two enzymes are pyridozal-5'-phosphate-dependent, and their substrate, L-cysteine, is derived from nutritional supports or liberated from endogenous proteins or peptides. It is also synthesized from L-methionine, and during this synthesis, homocysteine is produced as its intermediate [118-120]. Although higher level (millimolar) H<sub>2</sub>S exposure is cytotoxic (free radical generation, glutathione depletion, intracellular iron increase, mitochondrial cell death signal), lower level (micromolar) H<sub>2</sub>S shows cytoprotective (anti-necrotic and anti-apoptotic) effects.

Biochemical analysis have revealed that sulphide shows the direct antioxidant reaction with one- or two-electron molecules (one-electron molecules: •NO<sub>2</sub>, •OH, CO<sub>3</sub>•, two-electron molecules: peroxynitrite, hydrogen peroxide, hypochlorite, taurine chloramine) as well as other low-molecular-weight thiol molecules such as cysteine and glutathione. Although sulphide is not a preferential target for radicals or oxidants due to its low concentration *in vivo*, it can serve as a direct antioxidant [121, 122].

H<sub>2</sub>S induces upregulation of anti-inflammatory and cytoprotective genes, including HO-1 [120, 123]. As mentioned above, enhanced HO-1 activity produces CO, which shows cytoprotective effects such as anti-apoptosis, anti-inflammation, and antioxidant effects. This indicates that H<sub>2</sub>S might display cytoprotective effects together with the

induction of CO. In brain endothelial cells, H<sub>2</sub>S donor, sodium hydrosulfide (NaHS), protects against methionine-induced oxidative stress via induction of redox-regulated enzymes and proteins such as superoxide dismutase, catalase, apocynin, N-acetyl-l-cysteine, and reduced glutathione [124].

 $H_2S$  can open the potassium-operated ATP channel ( $K_{ATP}$  channel), which is associated with the distinct pharmacological effects of  $H_2S$  [125].  $K_{ATP}$  channel opening leads to the vasodilatory and myocardial preconditioning effects [126-128]. In the isolated organ system,  $H_2S$  results in smooth muscle relaxation possibly by endothelium-dependent hyperpolarizing factor-like effects [120]. Because the genetic deletion or pharmacological inhibitor of  $K_{ATP}$  channel increases myocardial infarct size even in  $H_2S$ -treated animals, precise role of  $K_{ATP}$  channel in tissue ischemia remains unsettled. [129-131].

H<sub>2</sub>S or its donors, NaHS and sodium sulphide provide cytoprotective actions in several disease models, such as myocardial ischemia, carrageenan-induced paw edema, air pouch inflammation, gastric ulcer, colorectal distension, non-steroidal anti-inflammatory drugs induced gastropathy model, and so on [128, 132-135]. On the other hand, H<sub>2</sub>S donor can exacerbate pathological processes in several disease models such as sepsis and stroke. Inhalation of H<sub>2</sub>S at high levels causes toxic effects such as a

transient loss of olfaction, pulmonary inflammation, cardiovascular events, neurological toxicity [136-138]. Cardioprotective effects mediated by H<sub>2</sub>S is bell-shaped, therefore, exquisite balance must be required in either endogenous or exogenous H<sub>2</sub>S derived cytoprotective actions [128]. (See also Fig. 3)

## $\underline{Molecular\ hydrogen\ (H_2)}$

Since the first striking evidence indicating that molecular hydrogen ( $H_2$ ) acts as an antioxidant and inhalation of  $H_2$ -containing gas reduces ischemic injury in brain [1], there have been increasing reports which support therapeutic properties of  $H_2$  against oxidative stress-related diseases and damages in the brain [139, 140], liver [141], intestinal graft [142], myocardial injury [143, 144], and atherosclerosis [145].  $H_2$  can be taken up by inhalation of  $H_2$ -containing gas ( $H_2$  gas) or drinking  $H_2$ -containing water ( $H_2$  water). One hour after the start of inhalation of  $H_2$  gas,  $H_2$  can be detectable in blood, at levels of 10  $\mu$ M in arterial blood [1]. The content of  $H_2$  can be measured even after intake of  $H_2$  water by a catheter, which shows 5  $\mu$ M in artery calculated after 3 min of  $H_2$  water ingestion [139].

H<sub>2</sub> in drinking water rescued the loss of dopaminergic neurons in MPTP-treated mice [146]. The therapeutic effects of H<sub>2</sub> water against Parkinson's disease model have also been confirmed in another animal model, 6-OHDA-treated rats [147]. 6-OHDA can

cause 8-oxoG, an oxidized form of guanine into DNA and RNA by •OH, leading to mitochondrial dysfunction through oxidative stress [148].

H<sub>2</sub> reduces cytotoxic •OH selectively, whereas the production of other radicals such as superoxide, hydrogen peroxide, nitric oxide are not altered by H<sub>2</sub> [1]. This selectivity was proved by cell-free system, and in particular, the preference of scavenging of •OH rather than superoxide was confirmed in PC12 cell culture system [1]. According to Setsukinai et al. [149], both •OH and peroxynitrite (ONOO<sup>-</sup>) were much more reactive than other ROS. This might address why H<sub>2</sub> shows selective reaction with only the strongest radicals both in the cell-free system and in PC12 cells.

Notably, •OH overproduction in oxidative and neurotoxic reaction by MPTP leads to lipid peroxidation observed by 4-hydroxynonenal (4-HNE) immunostaining in nigral dopaminergic neurons prior to cellular death. Immunoreactivity of 4-HNE in MPTP-treated mice is increased by three times as much as that in saline-treated mice [146], which is similar to the previous report of 4-HNE protein levels in substantia nigra observed at the same period after MPTP administration using HPLC [150]. H<sub>2</sub> water significantly reduces the formation of 4-HNE in dopaminergic neurons in the substantia nigra to the level of control [146]. On the other hand, the increase in superoxide, which is detectable by administration of dihydroethidine (DHE) intravenously, was not

significantly reduced by H<sub>2</sub> water [146]. Although H<sub>2</sub> reduces the production of superoxide in brain slices in hypoxia/reperfusion injury [151], H<sub>2</sub> water might show a preferential reduction of •OH during the protection of dopaminergic neurons.

H<sub>2</sub> water significantly reduces the accumulation of 8-oxoG in striatum after MPTP administration [146]. As mentioned above, 8-oxoG, an oxidized form of guanine into DNA and RNA by •OH, accumulates both in mitochondria and in nucleus; their nomenclature are mt8oxoG and nu8oxoG, respectively. Mt8oxoG accumulates in striatum which are rich in mitochondria in nerve terminal of dopaminergic neurons projected from the substantia nigra. Although nu8oxoG was not detected in nigral cell nucleus, H<sub>2</sub> water might prevent the mt8oxoG-induced cellular apoptotic signals, not just reduce •OH in dopaminergic nerve terminal.

H<sub>2</sub> was effective when it was inhaled during reperfusion; when H<sub>2</sub> was inhaled just during ischemia (not in the reperfusion stage), infarct volume was not significantly decreased [1]. It was shown that H<sub>2</sub> in the brain decreased immediately after stopping inhalation and completely disappeared within 10 minutes [146], indicating that the effect of H<sub>2</sub> can be observed only during the period when the oxidative insults occur. According to the previous report [139], H<sub>2</sub> could be detected in the blood 3 minutes after administration of H<sub>2</sub> water into the stomach. However, our unpublished data

showed that the half-life of H<sub>2</sub> in the muscle in rats was approximately 20 minutes after the administration of H<sub>2</sub> gas. Taking these reports into consideration, H<sub>2</sub> in the brain and other tissues does not stay long enough to exert its ability as an antioxidant to ROS directly. Therefore, it is unlikely that direct reaction of H<sub>2</sub> itself with ROS plays a major role in the neuroprotection, especially by H<sub>2</sub> in drinking water, even though H<sub>2</sub> itself has the ability to reduce •OH preferentially. In accordance with this hypothesis, drinking H<sub>2</sub> water increases urinary antioxidant enzyme, superoxide dismutase (SOD) [152], like polyphenols, as an endogenous defensive system against ROS (especially superoxide)-mediated cellular damage. Although it takes eight weeks for significant increase of SOD in human, H<sub>2</sub> is able to alter the expression level of urinary antioxidant enzyme. It was also reported that H<sub>2</sub> water increases total bilirubin for four to eight weeks when compared to baseline. Bilirubin is produced by the catalytic reaction of HO-1, and degradation of heme generates bilirubin as well as carbon monoxide and free iron. The increase of HO-1 expression may result from the response to oxidative stress, which is also characterized as a phase II antioxidant and positively regulated by several stress-responsive transcriptional factors [100]. Therefore, taking these observations into consideration, there seems to be other mechanisms for protective effect of H<sub>2</sub> in drinking water apart from inhalation. It is possible that drinking of H<sub>2</sub> water has not

only the ability to reduce cytotoxic radicals, but also novel mechanisms which are related to anti-oxidative defense system. These cytoprotective mechanisms of  $H_2$  are quite similar to those of other medical gases (gasotransmitters), CO and  $H_2S$ , as we have discussed above. In conclusion,  $H_2$  could be another gas mediator which shows therapeutic effects like CO and  $H_2S$  (Fig. 3).

# III. How ROS resistant proteins are made artificially: a different way to overcome ROS toxicity

As was described above, ROS toxicity was overcome by activation of SOD or by medical gases. Another efficient way to overcome ROS would be producing ROS-resistant proteins. There was a cohort study in Taiwan showing that low-dose ionizing irradiation for many years was turned out to treat and cure many illnesses, including cancer [153]. It is serendipitous application of low-dose radiation. Another interest comes from the fact obtained from "naked mole rat", which is well adapted for the limited availability with oxygen due to high affinity for oxygen of its hemoglobin. It has very low respiration and metabolic rate for animal of its size. It is extraordinarily long lived for a rodent of its size (up to 28 years) due to prevention of oxidative damage. According to Perez, et al. [3], the longevity of the rat may attribute to a very stable

structure of protein. This stability may be ascribed to high level of cysteine in their proteins for shielding them from oxidation. Unexpectedly, Western blot analysis of Hsp70 and Hsc70 levels in liver homogenates showed no age-related difference in the rat. This is because heat shock protein is produced by differential heat stimulation or transient ROS stimulations, while ROS resistant proteins produced in the rats without any kinds of transient stimulations. Therefore it is reasonable to assume that inherent cysteine (Cys) content may serve as a determination of longevity in the rat, while ROS-resistant protein may be also produced by transient ROS exposure repeatedly.

#### III-1. ROS resistant protein formation by low dose radiation

As described in I-1, radiation including X-ray is most effective means to produce ROS. Therefore it is likely that ROS resistant protein can be produced with high efficiency. In the case of heat shock protein, as a good example, it is already known that transient change in the external media from regular extracellular temperature to  $40^{\circ}\text{C}-43^{\circ}\text{C}$  will be a trigger for whole process.

The level of structural organization of globular proteins is;

Primary (amino acid) sequence → secondary sequence →

 $\rightarrow$  supersecondary sequence  $\rightarrow$  domain  $\rightarrow$  globular protein  $\rightarrow$  aggregation

The constitution determines the system of covalent bonds in the protein macromolecule

and includes information about disulfide bridges that form spontaneously during protein syntheses on ribosome. The secondary structure is a regular geometric figure of chain created by H bonding between the C-O and N-H group of peptide bonds. Further folding of polypeptide chains result in the formation of tertiary structures. They are produced by long range contacts within the chain and stabilized quaternary structure is an organization of protein subunits or two or more independent polypeptide chains.

The spontaneous act of protein folding is remarkable in that the complex motion of a protein structural element. This process results from thermally activated Brownian motion and resembles a phase transition, such as coupling. Consequently the net stabilization of the native state of protein conformation results from a balance of large force that favors both folding and unfolding. The net free energy of folding is 10 kcal/mol for hypothetical protein. By the combined effect of hydrophobicity of some side chains of amino acids and the structure of water side chain of non-polar amino acid residues is to locate in the interior of a protein. The order of hydrophobicity and hydrophilicity for the amino acids is:

Hydrophobic: Phe> Ala > Val > Leu > Cys

Hydrophilic: Tyr > Ser > Asp > Glu > Asn > Gln > Arg

Consequently, hydrophobic side chain coalesces in the interior of proteins. Much of the

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free energy of protein folding is entropic. It is possible protein unfolding will be happened when conformational entropy was increased. This process was well confirmed by us recently using deuteron as a probe [155]. This can be denominated as two stable state theory. Therefore it is natural that heat shock signal is triggered by unfolding of proteins which is formed by hydrophobic collapse, intramolecular H bonding and van der Waals interaction.

When ROS attacks inside the cells, hydrophilic amino acids are released in the cytosol then tertial structure of the functional protein are destroyed and Ser, Asp, Glu and Arg (well known neuro-transmitter) can move just like second messengers and activate synthesis of ROS-resistant proteins.

III-2. Overview of non-drug therapy according to the thermodynamic principle for redox signaling

We are living by consumption of Gibbs free energy (F), which is expressed as,

$$dF = dU - TdS$$

, where dU is the change in the internal energy of the system (e.g. energy owing to the formation of ATP from ADP, or growth, etc), T is absolute temperature, dS is the entropy change (e.g. the rate of heat flow to the environment, passive diffusion and heat production), dF is sum of the free energy introduced into the system (e.g. radiation,

oxidation and energy influx) and the work that may have to be done (e.g. muscular work, diffusion against concentration gradient).

Most of the processes in our living body system are non-voluntary biochemical reactions, such as protein synthesis from amino acids. Energy source in living cells to synthesize protein is supplied from ATP. When ATP is converted into ADP, Gibbs energy (G) is released by ATP decomposition, associated with increased entropy (S). The released G is used to connect amino acids to make protein. For example connection of 2 amino acids, decomposition of 3 ATP molecules is needed.

Attention must be paid that  $dS_1 < 0$  and  $dS_d > 0$ , where subscript "I" means living organism and "d" means death of living organism.  $S_d > 0$  coincides with the halt of feedback loop. Accordingly aging or senescence means  $S_1$  will increase gradually to positive values. What is the trigger signal for changing  $S_1$  from negative to positive? It is well known that irradiation, local heating and oxidation reaction resulted sequential rearrangements as described in section I. These changes lead to the degeneration of bio-molecules which then undergo further replication. Tertiary and quaternary structures of proteins, for example, are easily altered by thermal vibration. The denaturation of protein is observed at temperature above 42°C. UV and X-ray irradiation kill cells or make errors in replication and translation, which may lead to cancer. Ionizing radiation

can penetrate into the tissue much deeper and cause the decomposition of molecules. Finally, we concluded that removing toxic ROS and promotion of damage repair by producing chaperon are the most rational ways to survive and live a longer time. In Fig. 4, bird's-eye view of non-drug therapy for aging and health medical science is shown.

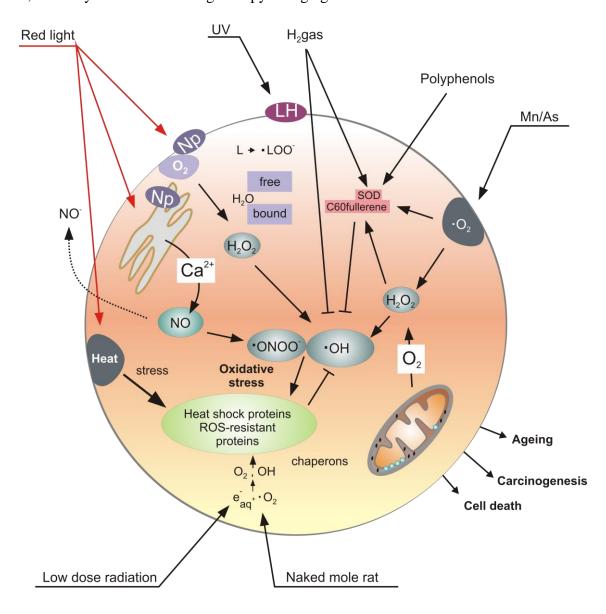


Fig. 4 Bird's-eye view of redox signaling in the living cell

Clockwise from the left bottom, low dose radiation, red light and UV are for ROS production; Hydrogen gas, minerals such as manganese and arsenic are for elimination of ROS-induced damage. Naked mole rat model may give us a novel idea how to make ROS-resistant proteins for longevity. Low dose ionizing radiation may also be a serendipity to produce chaperons as ROS-resistant proteins. LH: unsaturated fatty acids, Np: nanoparticle.

#### References

- [1] Ohsawa I, Ishikawa M, Takahashi K, et al. Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. Nat Med 2007; 13: 688-94.
- [2] Baggott J. Perfect Symmetry: The Accidental Discovery of Buckminsterfullerene, Oxford University Press: Oxford 1994.
- [3] Perez VI, Buffenstein R, Masamsetti V, et al. Protein stability and resistance to oxidative stress are determinants of longevity in the longest-living rodent, the naked mole-rat. *Proc Natl Acad Sci USA* 2009; 106: 3059-64.
- [4] Jacob C, Winyard PG. Redox Signaling and Regulation in Biology and Medicine, WILEY-VCH Verlag GmbH & Co. KGaA: Weinheim 2009.
- [5] Sies H, Jones D. Oxidative stress, in Encyclopedia of stress (Fink, G., ed.), San Diego: Elsevier 2007: pp 45-48
- [6] Sies H. Oxidative stress: oxidants and antioxidants. Exp Physiol 1997; 82: 291-5.
- [7] Petersen AB, Gniadecki R, Vicanova J, Thorn T, Wulf HC. Hydrogen peroxide is responsible for UVA-induced DNA damage measured by alkaline comet assay in HaCaT keratinocytes. *J Photochem Photobiol B* 2000; 59: 123-31.
- [8] Henderson BW, Dougherty TJ. How does photodynamic therapy work? *Photochem Photobiol* 1992; 55: 145-57.
- [9] Halliwell B. Oxygen and nitrogen are pro-carcinogens. Damage to DNA by reactive oxygen, chlorine and nitrogen species: measurement, mechanism and the effects of nutrition. *Mutat Res* 1999; 443: 37-52.
- [10] Valko M, Leibfritz D, Moncol J, et al. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 2007; 39: 44-84.
- [11] Eu JP, Liu L, Zeng M, Stamler JS. An apoptotic model for nitrosative stress. *Biochemistry* 2000; 39: 1040-7.
- [12] England K, O'Driscoll C, Cotter TG. Carbonylation of glycolytic proteins is a key response to drug-induced oxidative stress and apoptosis. *Cell Death Differ* 2004; 11: 252-60.
- [13] D'Alessio M, De Nicola M, Coppola S, *et al.* Oxidative Bax dimerization promotes its translocation to mitochondria independently of apoptosis. *FASEB J* 2005; 19: 1504-6.
- [14] Chrestensen CA, Starke DW, Mieyal JJ. Acute cadmium exposure inactivates thioltransferase (Glutaredoxin), inhibits intracellular reduction of protein-glutathionyl-mixed disulfides, and initiates apoptosis. *J Biol Chem* 2000;

- 275: 26556-65.
- [15] Ceccatelli S, Tamm C, Sleeper E, Orrenius S. Neural stem cells and cell death. \*Toxicol Lett 2004; 149: 59-66.
- [16] Miller BA, Zhang W. TRP Channels as Mediators of Oxidative Stress. *Adv Exp Med Biol* 2011; 704: 531-44.
- [17] Rosenfeldt MT, Ryan KM. The multiple roles of autophagy in cancer. Carcinogenesis.
- [18] Antelmann H, Helmann JD. Thiol-based redox switches and gene regulation. Antioxid Redox Signal 2011; 14: 1049-63.
- [19] Katagiri K, Matsuzawa A, Ichijo H. Regulation of apoptosis signal-regulating kinase 1 in redox signaling. *Methods Enzymol* 2010; 474: 277-88.
- [20] Ciesielski-Treska J, Ulrich G, Chasserot-Golaz S, et al. Mechanisms underlying neuronal death induced by chromogranin A-activated microglia. J Biol Chem 2001; 276: 13113-20.
- [21] Adayev T, Estephan R, Meserole S, et al. Externalization of phosphatidylserine may not be an early signal of apoptosis in neuronal cells, but only the phosphatidylserine-displaying apoptotic cells are phagocytosed by microglia. J Neurochem 1998; 71: 1854-64.
- [22] Aloy MT, Hadchity E, Bionda C, et al. Protective role of Hsp27 protein against gamma radiation-induced apoptosis and radiosensitization effects of Hsp27 gene silencing in different human tumor cells. Int J Radiat Oncol Biol Phys 2008; 70: 543-53.
- [23] Che Y, Wang JF, Shao L, Young T. Oxidative damage to RNA but not DNA in the hippocampus of patients with major mental illness. *J Psychiatry Neurosci* 2010; 35: 296-302.
- [24] Montaigne D, Marechal X, Baccouch R, et al. Stabilization of mitochondrial membrane potential prevents doxorubicin-induced cardiotoxicity in isolated rat heart. *Toxicol Appl Pharmacol* 2010; 244: 300-7.
- [25] Stryer L. Rapid motions in protein molecules. Biochem Soc Symp 1981: 39-55.
- [26] Rodriguez-Amaya DB. Food carotenoids: analysis, composition and alterations during storage and processing of foods. *Forum Nutr* 2003; 56: 35-7.
- [27] Grudzinski W, Matula M, Sielewiesiuk J, et al. Effect of 13-cis violaxanthin on organization of light harvesting complex II in monomolecular layers. Biochim Biophys Acta 2001; 1503: 291-302.
- [28] Santiveri CM, Perez-Canadillas JM, Vadivelu MK, et al. NMR structure of the alpha-hemoglobin stabilizing protein: insights into conformational heterogeneity and binding. J Biol Chem 2004; 279: 34963-70.

- [29] Pu H, Sakaguchi M, Kondo T, et al. Effects of oxygen concentrations on human fibroblasts treated with Fe(3+)-NTA. Int J Mol Med 2001; 7: 295-300.
- [30] Lin F, Geiger PG, Korytowski W, Girotti AW. Protoporphyrin IX-sensitized photoinactivation of 5-aminolevulinate-treated leukemia cells: effects of exogenous iron. *Photochem Photobiol* 1999; 69: 375-81.
- [31] Jomova K, Vondrakova D, Lawson M, Valko M. Metals, oxidative stress and neurodegenerative disorders. *Mol Cell Biochem* 2010; 345: 91-104.
- [32] Raji V, Kumar J, Rejiya CS, et al. Selective photothermal efficiency of citrate capped gold nanoparticles for destruction of cancer cells. Exp Cell Res 2011.
- [33] Grimland JL, Wu C, Ramoutar RR, Brumaghim JL, McNeill J. Photosensitizer-doped conjugated polymer nanoparticles with high cross-sections for one- and two-photon excitation. *Nanoscale* 2011; 3: 1451-5.
- [34] Valavanidis A, Vlahoyianni T, Fiotakis K. Comparative study of the formation of oxidative damage marker 8-hydroxy-2'-deoxyguanosine (8-OHdG) adduct from the nucleoside 2'-deoxyguanosine by transition metals and suspensions of particulate matter in relation to metal content and redox reactivity. *Free Radic Res* 2005; 39: 1071-81.
- [35] Ryu A, Arakane K, Koide C, Arai H, Nagano T. Squalene as a target molecule in skin hyperpigmentation caused by singlet oxygen. *Biol Pharm Bull* 2009; 32: 1504-9.
- [36] Kubin A, Alth G, Jindra R, Jessner G, Ebermann R. Wavelength-dependent photoresponse of biological and aqueous model systems using the photodynamic plant pigment hypericin. *J Photochem Photobiol B* 1996; 36: 103-8.
- [37] Sikorski M, Sikorska E, Koziolowa A, *et al.* Photophysical properties of lumichromes in water. *J Photochem Photobiol B* 2001; 60: 114-9.
- [38] Farmilo A, Wilkinson F. On the mechanism of quenching of singlet oxygen in solution. *Photochem Photobiol* 1973; 18: 447-50.
- [39] Kvam E, Tyrrell RM. Induction of oxidative DNA base damage in human skin cells by UV and near visible radiation. *Carcinogenesis* 1997; 18: 2379-84.
- [40] Tyrrell RM, Keyse SM. New trends in photobiology. The interaction of UVA radiation with cultured cells. *J Photochem Photobiol B* 1990; 4: 349-61.
- [41] Ebermann R, Alth G, Kreitner M, Kubin A. Natural products derived from plants as potential drugs for the photodynamic destruction of tumor cells. *J Photochem Photobiol B* 1996; 36: 95-7.
- [42] Wentworth P, Jr., McDunn JE, Wentworth AD, et al. Evidence for antibody-catalyzed ozone formation in bacterial killing and inflammation. Science 2002; 298: 2195-9.
- [43] Lerner RA, Eschenmoser A. Ozone in biology. Proc Natl Acad Sci USA 2003; 100:

- 3013-5.
- [44] Babior BM, Takeuchi C, Ruedi J, Gutierrez A, Wentworth P, Jr. Investigating antibody-catalyzed ozone generation by human neutrophils. *Proc Natl Acad Sci U S A* 2003; 100: 3031-4.
- [45] Schwartsburd PM, Lankin VZ. Lipoproteins in hypoxic tumor cells as traps of free radicals. *Med Oncol* 1994; 11: 101-10.
- [46] Ginsburg I, Sadovnic M, Yedgar S, Kohen R, Hrbac J. Hemolysis of human erythrocytes by hypochlorous acid is modulated by amino acids, antioxidants, oxidants, membrane-perforating agents and by divalent metals. *Free Radic Res* 2002; 36: 607-19.
- [47] Nakajima T, Yukawa O. Mechanism of radiation-induced diacylglycerol production in primary cultured rat hepatocytes. *J Radiat Res (Tokyo)* 1999; 40: 135-44.
- [48] Adibhatla RM, Hatcher JF, Dempsey RJ. Phospholipase A2, hydroxyl radicals, and lipid peroxidation in transient cerebral ischemia. *Antioxid Redox Signal* 2003; 5: 647-54.
- [49] Corre I, Niaudet C, Paris F. Plasma membrane signaling induced by ionizing radiation. *Mutat Res* 2010; 704: 61-7.
- [50] Kim GJ, Chandrasekaran K, Morgan WF. Mitochondrial dysfunction, persistently elevated levels of reactive oxygen species and radiation-induced genomic instability: a review. *Mutagenesis* 2006; 21: 361-7.
- [51] Zhang QJ, Wu K. [Biological oxidative stress induced by electromagnetic irradiation]. Space Med Med Eng (Beijing) 2004; 17: 152-6.
- [52] Sanovic R, Krammer B, Grumboeck S, Verwanger T. Time-resolved gene expression profiling of human squamous cell carcinoma cells during the apoptosis process induced by photodynamic treatment with hypericin. *Int J Oncol* 2009; 35: 921-39.
- [53] Moan J, Berg K, Kvam E, et al. Intracellular localization of photosensitizers. Ciba Found Symp 1989; 146: 95-107; discussion 107-11.
- [54] Cernay T, Zimmermann HW. Selective photosensitization of mitochondria by the lipophilic cationic porphyrin POR10. *J Photochem Photobiol B* 1996; 34: 191-6.
- [55] Delaey EM, Obermueller R, Zupko I, et al. In vitro study of the photocytotoxicity of some hypericin analogs on different cell lines. *Photochem Photobiol* 2001; 74: 164-71.
- [56] Ishisaka R, Kanno T, Kanematsu H, et al. Superoxide dismutase-like activity of metal substituted lactoferrin derivatives. Physiol Chem Phys Med NMR 1998; 30: 1-13.
- [57] Suzuki K, Miyazawa N, Nakata T, et al. High copper and iron levels and expression

- of Mn-superoxide dismutase in mutant rats displaying hereditary hepatitis and hepatoma (LEC rats). *Carcinogenesis* 1993; 14: 1881-4.
- [58] Ozata M, Uckaya G, Aydin A, Isimer A, Ozdemir IC. Defective antioxidant defense system in patients with a human leptin gene mutation. *Horm Metab Res* 2000; 32: 269-72.
- [59] De Vizcaya-Ruiz A, Barbier O, Ruiz-Ramos R, Cebrian ME. Biomarkers of oxidative stress and damage in human populations exposed to arsenic. *Mutat Res* 2009; 674: 85-92.
- [60] Kligerman AD, Tennant AH. Insights into the carcinogenic mode of action of arsenic.

  \*Toxicol Appl Pharmacol\*\* 2007; 222: 281-8.
- [61] Shannon RL, Strayer DS. Arsenic-induced skin toxicity. *Hum Toxicol* 1989; 8: 99-104.
- [62] Schwartz RA. Arsenic and the skin. Int J Dermatol 1997; 36: 241-50.
- [63] Shi H, Hudson LG, Ding W, et al. Arsenite causes DNA damage in keratinocytes via generation of hydroxyl radicals. *Chem Res Toxicol* 2004; 17: 871-8.
- [64] Hilmy AM, el-Domiaty NA, Kamal MA, Mohamed MA, Abou Samra WE. Effect of some arsenic antagonists on the toxicity, distribution and excretion of arsenite and arsenate in rats. *Comp Biochem Physiol C* 1991; 99: 357-62.
- [65] Kimura A, Ishida Y, Wada T, *et al.* MRP-1 expression levels determine strain-specific susceptibility to sodium arsenic-induced renal injury between C57BL/6 and BALB/c mice. *Toxicol Appl Pharmacol* 2005; 203: 53-61.
- [66] Li J, Zhang M, Yang C, et al. Nitroglycerin protects small intestine from ischemia-reperfusion injury via NO-cGMP pathway and upregulation of alpha-CGRP. J Gastrointest Surg 2009; 13: 478-85.
- [67] Cooper KL, Meng Y, Harnan S, et al. Positron emission tomography (PET) and magnetic resonance imaging (MRI) for the assessment of axillary lymph node metastases in early breast cancer: systematic review and economic evaluation. Health Technol Assess 2011; 15: iii-iv, 1-134.
- [68] See V, Loeffler JP. Oxidative stress induces neuronal death by recruiting a protease and phosphatase-gated mechanism. *J Biol Chem* 2001; 276: 35049-59.
- [69] Toledano MB, Delaunay A, Monceau L, Tacnet F. Microbial H2O2 sensors as archetypical redox signaling modules. *Trends Biochem Sci* 2004; 29: 351-7.
- [70] Podesta D, Fernandez Villamil SH, Stoppani AO. [Inhibitory action of Fenton systems on topoisomerase I from Trypanosoma cruzi and Crithidia fasciculata]]. Rev Argent Microbiol 2003; 35: 80-5.
- [71] Monfort V, Pouthas V. Effects of working memory demands on frontal slow waves in

- time-interval reproduction tasks in humans. Neurosci Lett 2003; 343: 195-9.
- [72] Von Burg R. Carbon monoxide. *J Appl Toxicol* 1999; 19: 379-86.
- [73] Sjostrand T. Early studies of CO production. Ann NYAcad Sci 1970; 174: 5-10.
- [74] Otterbein LE, Bach FH, Alam J, et al. Carbon monoxide has anti-inflammatory effects involving the mitogen-activated protein kinase pathway. Nat Med 2000; 6: 422-8.
- [75] Morse D, Pischke SE, Zhou Z, et al. Suppression of inflammatory cytokine production by carbon monoxide involves the JNK pathway and AP-1. J Biol Chem 2003; 278: 36993-8.
- [76] Chapman JT, Otterbein LE, Elias JA, Choi AM. Carbon monoxide attenuates aeroallergen-induced inflammation in mice. *Am J Physiol Lung Cell Mol Physiol* 2001; 281: L209-16.
- [77] Otterbein LE, Mantell LL, Choi AM. Carbon monoxide provides protection against hyperoxic lung injury. *Am J Physiol* 1999; 276: L688-94.
- [78] Otterbein LE, Otterbein SL, Ifedigbo E, et al. MKK3 mitogen-activated protein kinase pathway mediates carbon monoxide-induced protection against oxidant-induced lung injury. Am J Pathol 2003; 163: 2555-63.
- [79] Zhou Z, Song R, Fattman CL, et al. Carbon monoxide suppresses bleomycin-induced lung fibrosis. Am J Pathol 2005; 166: 27-37.
- [80] Guo Y, Stein AB, Wu WJ, et al. Administration of a CO-releasing molecule at the time of reperfusion reduces infarct size in vivo. Am J Physiol Heart Circ Physiol 2004; 286: H1649-53.
- [81] Stein AB, Guo Y, Tan W, et al. Administration of a CO-releasing molecule induces late preconditioning against myocardial infarction. J Mol Cell Cardiol 2005; 38: 127-34.
- [82] Johnson RA, Lavesa M, DeSeyn K, Scholer MJ, Nasjletti A. Heme oxygenase substrates acutely lower blood pressure in hypertensive rats. *Am J Physiol* 1996; 271: H1132-8.
- [83] Motterlini R, Clark JE, Foresti R, et al. Carbon monoxide-releasing molecules: characterization of biochemical and vascular activities. Circ Res 2002; 90: E17-24.
- [84] Li P, Jiang H, Yang L, et al. Angiotensin II induces carbon monoxide production in the perfused kidney: relationship to protein kinase C activation. Am J Physiol Renal Physiol 2004; 287: F914-20.
- [85] Teran FJ, Johnson RA, Stevenson BK, et al. Heme oxygenase-derived carbon monoxide promotes arteriolar endothelial dysfunction and contributes to salt-induced hypertension in Dahl salt-sensitive rats. Am J Physiol Regul Integr

- Comp Physiol 2005; 288: R615-22.
- [86] Juan SH, Lee TS, Tseng KW, et al. Adenovirus-mediated heme oxygenase-1 gene transfer inhibits the development of atherosclerosis in apolipoprotein E-deficient mice. *Circulation* 2001; 104: 1519-25.
- [87] Otterbein LE, Zuckerbraun BS, Haga M, et al. Carbon monoxide suppresses arteriosclerotic lesions associated with chronic graft rejection and with balloon injury. Nat Med 2003; 9: 183-90.
- [88] Nakao A, Kimizuka K, Stolz DB, et al. Protective effect of carbon monoxide inhalation for cold-preserved small intestinal grafts. Surgery 2003; 134: 285-92.
- [89] Amersi F, Shen XD, Anselmo D, et al. Ex vivo exposure to carbon monoxide prevents hepatic ischemia/reperfusion injury through p38 MAP kinase pathway. *Hepatology* 2002; 35: 815-23.
- [90] Zhang X, Shan P, Alam J, et al. Carbon monoxide modulates Fas/Fas ligand, caspases, and Bcl-2 family proteins via the p38alpha mitogen-activated protein kinase pathway during ischemia-reperfusion lung injury. J Biol Chem 2003; 278: 22061-70.
- [91] Zhang X, Shan P, Otterbein LE, et al. Carbon monoxide inhibition of apoptosis during ischemia-reperfusion lung injury is dependent on the p38 mitogen-activated protein kinase pathway and involves caspase 3. J Biol Chem 2003; 278: 1248-58.
- [92] Sato K, Balla J, Otterbein L, *et al.* Carbon monoxide generated by heme oxygenase-1 suppresses the rejection of mouse-to-rat cardiac transplants. *J Immunol* 2001; 166: 4185-94.
- [93] Song R, Kubo M, Morse D, et al. Carbon monoxide induces cytoprotection in rat orthotopic lung transplantation via anti-inflammatory and anti-apoptotic effects.

  Am J Pathol 2003; 163: 231-42.
- [94] Maines MD. The heme oxygenase system: update 2005. *Antioxid Redox Signal* 2005; 7: 1761-6.
- [95] Piantadosi CA. Carbon monoxide, reactive oxygen signaling, and oxidative stress. Free Radic Biol Med 2008; 45: 562-9.
- [96] Maines MD, Trakshel GM, Kutty RK. Characterization of two constitutive forms of rat liver microsomal heme oxygenase. Only one molecular species of the enzyme is inducible. *J Biol Chem* 1986; 261: 411-9.
- [97] Trakshel GM, Kutty RK, Maines MD. Purification and characterization of the major constitutive form of testicular heme oxygenase. The noninducible isoform. J Biol Chem 1986; 261: 11131-7.
- [98] Wakabayashi N, Dinkova-Kostova AT, Holtzclaw WD, et al. Protection against

- electrophile and oxidant stress by induction of the phase 2 response: fate of cysteines of the Keap1 sensor modified by inducers. *Proc Natl Acad Sci U S A* 2004; 101: 2040-5.
- [99] Martin D, Rojo AI, Salinas M, et al. Regulation of heme oxygenase-1 expression through the phosphatidylinositol 3-kinase/Akt pathway and the Nrf2 transcription factor in response to the antioxidant phytochemical carnosol. *J Biol Chem* 2004; 279: 8919-29.
- [100] Alam J, Cook JL. Transcriptional regulation of the heme oxygenase-1 gene via the stress response element pathway. *Curr Pharm Des* 2003; 9: 2499-511.
- [101] Thom SR, Fisher D, Xu YA, Notarfrancesco K, Ischiropoulos H. Adaptive responses and apoptosis in endothelial cells exposed to carbon monoxide. *Proc Natl Acad Sci U* SA 2000; 97: 1305-10.
- [102] Tenhunen R, Marver HS, Schmid R. Microsomal heme oxygenase. Characterization of the enzyme. *J Biol Chem* 1969; 244: 6388-94.
- [103] Ryter SW, Alam J, Choi AM. Heme oxygenase-1/carbon monoxide: from basic science to therapeutic applications. *Physiol Rev* 2006; 86: 583-650.
- [104] Tenhunen R, Ross ME, Marver HS, Schmid R. Reduced nicotinamide-adenine dinucleotide phosphate dependent biliverdin reductase: partial purification and characterization. *Biochemistry* 1970; 9: 298-303.
- [105] Stocker R, Glazer AN, Ames BN. Antioxidant activity of albumin-bound bilirubin. *Proc Natl Acad Sci USA* 1987; 84: 5918-22.
- [106] Neuzil J, Stocker R. Free and albumin-bound bilirubin are efficient co-antioxidants for alpha-tocopherol, inhibiting plasma and low density lipoprotein lipid peroxidation. *J Biol Chem* 1994; 269: 16712-9.
- [107] Stocker R, Ames BN. Potential role of conjugated bilirubin and copper in the metabolism of lipid peroxides in bile. *Proc Natl Acad Sci USA* 1987; 84: 8130-4.
- [108] Stocker R, Yamamoto Y, McDonagh AF, Glazer AN, Ames BN. Bilirubin is an antioxidant of possible physiological importance. *Science* 1987; 235: 1043-6.
- [109] Trumpower BL. The protonmotive Q cycle. Energy transduction by coupling of proton translocation to electron transfer by the cytochrome bc1 complex. *J Biol Chem* 1990; 265: 11409-12.
- [110] Virbasius JV, Scarpulla RC. Activation of the human mitochondrial transcription factor A gene by nuclear respiratory factors: a potential regulatory link between nuclear and mitochondrial gene expression in organelle biogenesis. *Proc Natl Acad Sci USA* 1994; 91: 1309-13.
- [111] Lee SR, Yang KS, Kwon J, et al. Reversible inactivation of the tumor suppressor

- PTEN by H2O2. J Biol Chem 2002; 277: 20336-42.
- [112] Chen K, Thomas SR, Albano A, Murphy MP, Keaney JF, Jr. Mitochondrial function is required for hydrogen peroxide-induced growth factor receptor transactivation and downstream signaling. *J Biol Chem* 2004; 279: 35079-86.
- [113] Leslie NR, Bennett D, Lindsay YE, et al. Redox regulation of PI 3-kinase signalling via inactivation of PTEN. EMBO J 2003; 22: 5501-10.
- [114] Zhang X, Shan P, Alam J, Fu XY, Lee PJ. Carbon monoxide differentially modulates STAT1 and STAT3 and inhibits apoptosis via a phosphatidylinositol 3-kinase/Akt and p38 kinase-dependent STAT3 pathway during anoxia-reoxygenation injury. *J Biol Chem* 2005; 280: 8714-21.
- [115] Piantadosi CA, Suliman HB. Mitochondrial transcription factor A induction by redox activation of nuclear respiratory factor 1. *J Biol Chem* 2006; 281: 324-33.
- [116] Brancho D, Tanaka N, Jaeschke A, et al. Mechanism of p38 MAP kinase activation in vivo. Genes Dev 2003; 17: 1969-78.
- [117] Kim HP, Wang X, Zhang J, et al. Heat shock protein-70 mediates the cytoprotective effect of carbon monoxide: involvement of p38 beta MAPK and heat shock factor-1. J Immunol 2005; 175: 2622-9.
- [118] Wang R. Two's company, three's a crowd: can H2S be the third endogenous gaseous transmitter? *FASEB J* 2002; 16: 1792-8.
- [119] Fiorucci S, Distrutti E, Cirino G, Wallace JL. The emerging roles of hydrogen sulfide in the gastrointestinal tract and liver. *Gastroenterology* 2006; 131: 259-71.
- [120] Szabo C. Hydrogen sulphide and its therapeutic potential. *Nat Rev Drug Discov* 2007; 6: 917-35.
- [121] Whiteman M, Armstrong JS, Chu SH, et al. The novel neuromodulator hydrogen sulfide: an endogenous peroxynitrite 'scavenger'? J Neurochem 2004; 90: 765-8.
- [122] Carballal S, Trujillo M, Cuevasanta E, et al. Reactivity of hydrogen sulfide with peroxynitrite and other oxidants of biological interest. Free Radic Biol Med 2011; 50: 196-205.
- [123] Qingyou Z, Junbao D, Weijin Z, et al. Impact of hydrogen sulfide on carbon monoxide/heme oxygenase pathway in the pathogenesis of hypoxic pulmonary hypertension. Biochem Biophys Res Commun 2004; 317: 30-7.
- [124] Tyagi N, Moshal KS, Sen U, et al. H2S protects against methionine-induced oxidative stress in brain endothelial cells. Antioxid Redox Signal 2009; 11: 25-33.
- [125] Zhao W, Zhang J, Lu Y, Wang R. The vasorelaxant effect of H(2)S as a novel endogenous gaseous K(ATP) channel opener. *EMBO J* 2001; 20: 6008-16.
- [126] Auchampach JA, Maruyama M, Cavero I, Gross GJ. Pharmacological evidence for a

- role of ATP-dependent potassium channels in myocardial stunning. *Circulation* 1992; 86: 311-9.
- [127] Toombs CF, Moore TL, Shebuski RJ. Limitation of infarct size in the rabbit by ischaemic preconditioning is reversible with glibenclamide. *Cardiovasc Res* 1993; 27: 617-22.
- [128] Johansen D, Ytrehus K, Baxter GF. Exogenous hydrogen sulfide (H2S) protects against regional myocardial ischemia-reperfusion injury--Evidence for a role of K ATP channels. *Basic Res Cardiol* 2006; 101: 53-60.
- [129] Miura T, Miki T. ATP-sensitive K+ channel openers: old drugs with new clinical benefits for the heart. *Curr Vasc Pharmacol* 2003; 1: 251-8.
- [130] Yamada S, Kane GC, Behfar A, et al. Protection conferred by myocardial ATP-sensitive K+ channels in pressure overload-induced congestive heart failure revealed in KCNJ11 Kir6.2-null mutant. J Physiol 2006; 577: 1053-65.
- [131] Gumina RJ, O'Cochlain DF, Kurtz CE, et al. KATP channel knockout worsens myocardial calcium stress load in vivo and impairs recovery in stunned heart. Am J Physiol Heart Circ Physiol 2007; 292: H1706-13.
- [132] Zanardo RC, Brancaleone V, Distrutti E, et al. Hydrogen sulfide is an endogenous modulator of leukocyte-mediated inflammation. FASEB J 2006; 20: 2118-20.
- [133] Wallace JL, Dicay M, McKnight W, Martin GR. Hydrogen sulfide enhances ulcer healing in rats. *FASEB J* 2007; 21: 4070-6.
- [134] Yonezawa D, Sekiguchi F, Miyamoto M, et al. A protective role of hydrogen sulfide against oxidative stress in rat gastric mucosal epithelium. *Toxicology* 2007; 241: 11-8.
- [135] Distrutti E, Sediari L, Mencarelli A, et al. Evidence that hydrogen sulfide exerts antinociceptive effects in the gastrointestinal tract by activating KATP channels. J Pharmacol Exp Ther 2006; 316: 325-35.
- [136] Beauchamp RO, Jr., Bus JS, Popp JA, Boreiko CJ, Andjelkovich DA. A critical review of the literature on hydrogen sulfide toxicity. Crit Rev Toxicol 1984; 13: 25-97.
- [137] Woodall GM, Smith RL, Granville GC. Proceedings of the Hydrogen Sulfide Health Research and Risk Assessment Symposium October 31-November 2, 2000. *Inhal Toxicol* 2005; 17: 593-639.
- [138] Reiffenstein RJ, Hulbert WC, Roth SH. Toxicology of hydrogen sulfide. Annu Rev Pharmacol Toxicol 1992; 32: 109-34.
- [139] Nagata K, Nakashima-Kamimura N, Mikami T, Ohsawa I, Ohta S. Consumption of molecular hydrogen prevents the stress-induced impairments in

- hippocampus-dependent learning tasks during chronic physical restraint in mice. Neuropsychopharmacology 2009; 34: 501-8.
- [140] Gu Y, Huang CS, Inoue T, et al. Drinking hydrogen water ameliorated cognitive impairment in senescence-accelerated mice. J Clin Biochem Nutr 2010; 46: 269-76.
- [141] Fukuda K, Asoh S, Ishikawa M, et al. Inhalation of hydrogen gas suppresses hepatic injury caused by ischemia/reperfusion through reducing oxidative stress. Biochem Biophys Res Commun 2007; 361: 670-4.
- [142] Buchholz BM, Kaczorowski DJ, Sugimoto R, et al. Hydrogen inhalation ameliorates oxidative stress in transplantation induced intestinal graft injury. Am J Transplant 2008; 8: 2015-24.
- [143] Nakao A, Kaczorowski DJ, Wang Y, et al. Amelioration of rat cardiac cold ischemia/reperfusion injury with inhaled hydrogen or carbon monoxide, or both. J Heart Lung Transplant 2010; 29: 544-53.
- [144] Hayashida K, Sano M, Ohsawa I, et al. Inhalation of hydrogen gas reduces infarct size in the rat model of myocardial ischemia-reperfusion injury. Biochem Biophys Res Commun 2008; 373: 30-5.
- [145] Ohsawa I, Nishimaki K, Yamagata K, Ishikawa M, Ohta S. Consumption of hydrogen water prevents atherosclerosis in apolipoprotein E knockout mice. Biochem Biophys Res Commun 2008; 377: 1195-8.
- [146] Fujita K, Seike T, Yutsudo N, et al. Hydrogen in drinking water reduces dopaminergic neuronal loss in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. PLoS One 2009; 4: e7247.
- [147] Fu Y, Ito M, Fujita Y, et al. Molecular hydrogen is protective against 6-hydroxydopamine-induced nigrostriatal degeneration in a rat model of Parkinson's disease. *Neurosci Lett* 2009; 453: 81-5.
- [148] Stack EC, Ferro JL, Kim J, et al. Therapeutic attenuation of mitochondrial dysfunction and oxidative stress in neurotoxin models of Parkinson's disease. Biochim Biophys Acta 2008; 1782: 151-62.
- [149] Setsukinai K, Urano Y, Kakinuma K, Majima HJ, Nagano T. Development of novel fluorescence probes that can reliably detect reactive oxygen species and distinguish specific species. *J Biol Chem* 2003; 278: 3170-5.
- [150] Liang LP, Huang J, Fulton R, Day BJ, Patel M. An orally active catalytic metalloporphyrin protects against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity in vivo. *J Neurosci* 2007; 27: 4326-33.
- [151] Sato Y, Kajiyama S, Amano A, et al. Hydrogen-rich pure water prevents superoxide formation in brain slices of vitamin C-depleted SMP30/GNL knockout mice. Biochem

- Biophys Res Commun 2008; 375: 346-50.
- [152] Nakao A, Toyoda Y, Sharma P, Evans M, Guthrie N. Effectiveness of hydrogen rich water on antioxidant status of subjects with potential metabolic syndrome-an open label pilot study. *J Clin Biochem Nutr* 2010; 46: 140-9.
- [153] Chen WL, Luan YC, Shieh MC, et al. Effects of cobalt-60 exposure on health of Taiwan residents suggest new approach needed in radiation protection. Dose Response 2007; 5: 63-75.
- [154] Csiszar A, Labinskyy N, Orosz Z, et al. Vascular aging in the longest-living rodent, the naked mole rat. Am J Physiol Heart Circ Physiol 2007; 293: H919-27.
- [155] Hirakura Y, Sugiyama T, Takeda M, Ikeda M, Yoshioka T. Deuteration as a tool in investigating the role of protons in cell signaling. *Biochim Biophys Acta* 2011; 1810: 218-25.