

# Diversification and Evolution of the Signal Transduction System : Rapid Divergence of Tissue Specific Genes in Early Evolution of Chordates

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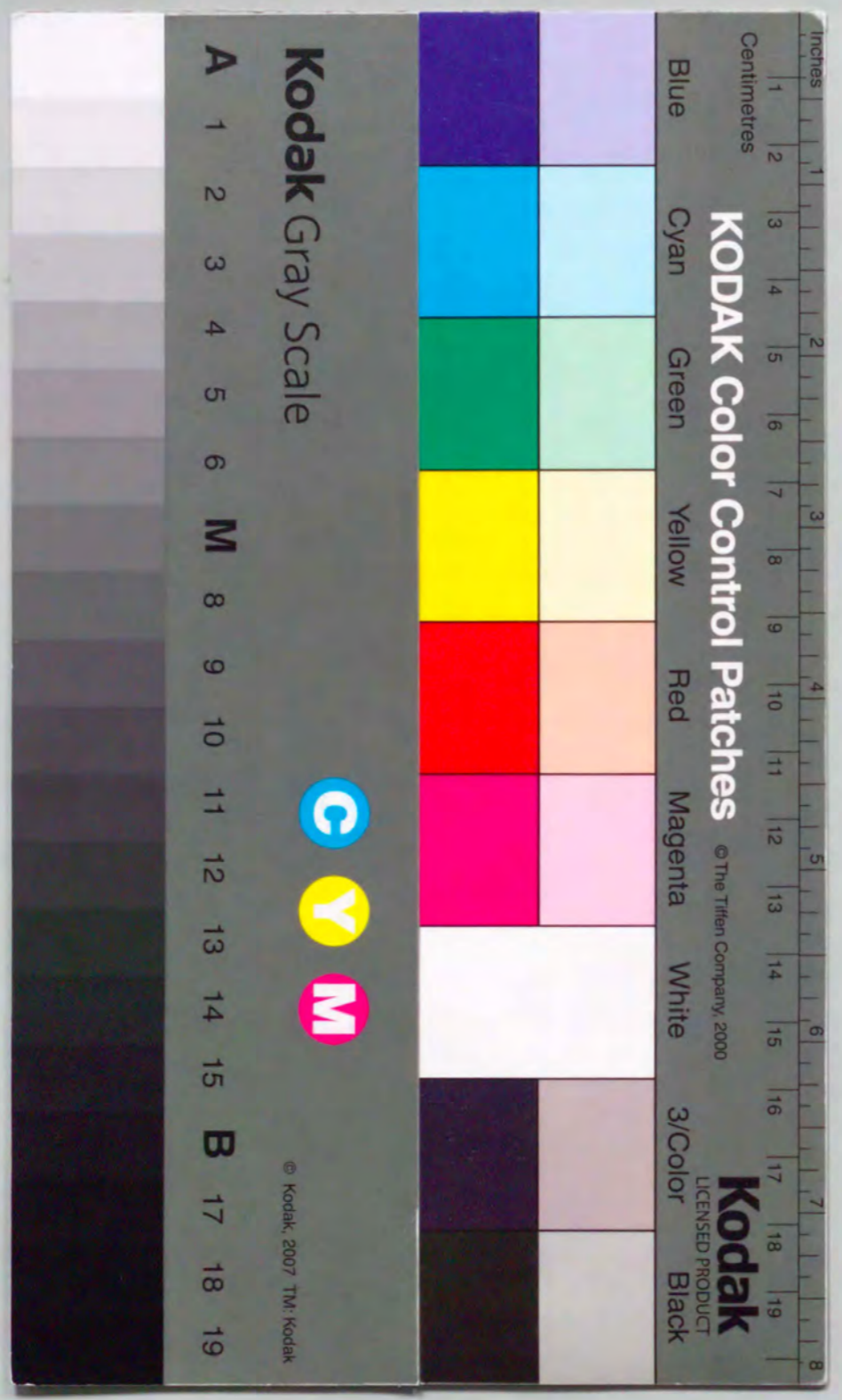
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Diversification and Evolution of the Signal Transduction System:  
Rapid Divergence of Tissue Specific Genes  
in Early Evolution of Chordates

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Diversification and Evolution of the Signal Transduction System:  
Rapid Divergence of Tissue Specific Genes  
in Early Evolution of Chordates

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Summary

The summary section discusses the main findings of the report, including the definition of economic growth, the review of current knowledge on rapidly growing economies, and the prospects for growth and development in the Third World. It also includes conclusions and recommendations.

Recently, many genes of key molecules playing important roles in the signal transduction systems have been cloned and sequenced. These key components, including rhodopsin family receptors (G protein coupled receptors), G proteins, adenylyl cyclases, phosphodiesterases, and phospholipase Cs, comprise large families whose genes were derived from each ancestral precursor by gene duplications. In order to clarify how the genes in the signal transduction systems diverged during the evolution of organisms, especially of animals, molecular phylogenetic trees of the receptors, G protein  $\alpha$  subunits, adenylyl cyclases, phosphodiesterases, and phospholipase Cs were reconstructed. On the phylogenetic tree of 124 rhodopsin family receptors, those with similar ligand binding specificities made clusters. The phylogenetic tree of 45 G protein  $\alpha$  subunits was also inferred. On this tree, G protein  $\alpha$  subunits regulating a specific type of amplifier also made a cluster. On both receptor and G protein  $\alpha$  subunit trees, genes with different functions are shown to have diverged before the separation of vertebrate (or deuterostomia) and insect (or protostomia) lineages, while genes duplicated after the divergence of the two animal lineages seems to have same function but different tissue distributions. The receptors binding with same ligand not necessarily interact with similar G proteins. For example, one subtype of muscarinic acetylcholine receptor (mAChR), m1, activating G protein subtype q and then stimulating PLC  $\beta$  subtypes in the next signal transducing step, whereas the other mAChR subtype, m2, activating G protein subtype i and then inhibiting adenylyl cyclases. The two receptors are closely related on the phylogenetic tree but the two G protein subtypes are distantly related. The duplicated genes of receptors binding with same ligand but coupling with different types of G proteins seem to guarantee the development of various types of cells and tissues in both vertebrate and insect lineages. The possible combinations of the signal pathways mediated by the receptors, G protein subunits, and

amplifiers known at present are enormous number even though many of them have never been realized. In the case of the animal amplifiers of G proteins, it becomes clear from the molecular phylogenetic analyses that subtypes of the amplifiers regulated by G proteins, like adenylyl cyclases, cGMP phosphodiesterases, and phospholipase C  $\beta$  subtype, were diverged from non-G protein coupled types, like guanylyl cyclase, calmodulin-dependent phosphodiesterase, and phospholipase C  $\gamma$  subtype, respectively, before the two animal lineages were separated.

It is generally thought that evolutionary changes at phenotypic and molecular levels are caused by distinct mechanisms, Darwinian selection of selectively positive mutants on one hand and random fixation of selectively neutral mutants on the other hand (Kimura, 1983). It is of particular importance to understand evolution at two levels in a unified way, but no data suggesting a link between them has been provided to date. The explosive diversification of metazoa at the early Cambrian would provide a unique opportunity to find a clue for the problem: It is widely accepted that the invasion of new vacant niches followed by relaxed selective constraints at the phenotypic level is a prerequisite for the rapid evolutionary burst at high taxonomic levels (Simpson, 1967; Kimura, 1991; Valentine, 1977). If this hypothesis is correct, the liberation from selective constraints at the phenotypic level is expected to reduce functional constraints at the molecular level, giving rise to rapid accumulations of genetic variations, because molecules that are expressed tissue specifically are constrained not only from functional requirements of individual molecules, but also from higher levels like tissues or organs (Kuma et al., 1993), and a partial lack of tissue function results in the elevated rate of molecular evolution (Hendriks et al., 1987). From a phylogenetic analysis of tissue specific genes we report here that gene duplications as well as amino acid substitutions had occurred with rapid rate during the evolution of echinoderm/chordate common ancestor and the early evolution of chordates,

but these genetic changes had remarkably reduced in the later stages. Thus these results are supporting evidence for the above hypothesis from molecular data and suggests a link between evolution at phenotypic and molecular levels.

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## Introduction

Recently, the signal transduction systems converting extracellular signals through G proteins to intracellular second messengers have been enthusiastically studied in molecular level. The proteins participating in the systems were identified and classified into four groups, receptors, transducers, amplifiers, and effectors. As the primary structures of these proteins were revealed more and more, it was found that there were several gene families. Rhodopsin, a photo receptor having seven transmembrane regions, is homologous to the receptors of neurotransmitters, peptide hormones, and odorant substances (for review see Birnbaumer et al., 1990; Iyengar and Birnbaumer, 1990). Heterotrimeric guanine nucleotide binding proteins (or G proteins), consist of  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits, each make gene families (for review see Iyengar and Birnbaumer, 1990; Kaziro et al., 1990; Kaziro et al., 1991; Simon et al., 1991; Wilkie et al., 1992; Hepler and Gilman, 1992; Birnbaumer, 1992). G proteins regulates amplifiers such as adenylyl cyclase (for review see Tang and Gilman, 1992), phosphodiesterase (for review see Beavo and Reifsnyder, 1990), phospholipase C (for review see Rhee and Choi, 1992), which also make their own large gene families, independently. Second messengers, like cyclic nucleotides (cAMP and cGMP), inositol 1,4,5-trisphosphate, and diacylglycerol, regulates a great number of effectors including protein kinases, channels, calmodulin, etc. Because the members of each gene family of signal transduction systems were diversified by gene duplications, the combinations of these factors became larger and larger, which ensured multicellular organisms to develop many types of cells and tissues. It is very important to study the diversification process of each factor and the interaction of one factor to the other in the cell.

On this study, we paid much attention to the diverging points of vertebrates and insects (and mollusks) on the phylogenetic trees because the divergence time of the two animal lineages seemed to be good measure to

divide the gene duplications into two types. The gene duplications before the divergence time produced the genes of different functions, whereas those after the time generated the genes of same function but different tissue expressions.

## Material and Methods

**Sequence Data Sources** Sequence data were from GenBank Release 77.0 and NBRF (National biochemical Research Foundation) Release 36.0. Accession numbers of the data were listed on tables and figure legends. Data not from the data bases were referred.

**Phylogenetic Tree Inference** Amino acid sequence data of each gene family were aligned for their homologous regions (Miyata et al., 1985). On the basis of the alignments, the evolutionary distance  $k$  was calculated between each pair of the sequence, excluding amino acid positions where gaps existed in any one of the aligned sequences; the distance  $k$  was measured by calculating the amino acid difference  $K$  (per site) between sequences compared and correcting multiple substitutions as  $k = -\ln(1 - K)$  (Jukes and Cantor, 1969; Kimura, 1983). Based on the evolutionary distances, phylogenetic trees were inferred by the neighbor-joining (NJ) method (Saitou and Nei, 1987). The reliability of the inferred tree was evaluated by bootstrap probability (Felsenstein, 1985).

**Calculation of Nonsynonymous and Synonymous Substitutions** Corrected nucleotide differences at the nonsynonymous ( $k_A$ ) and synonymous ( $k_S$ ) were calculated (Miyata and Yasunaga, 1980) for the homologous regions of the gene families.

## Results and Discussion

**Rhodopsin Family Receptors** Extracellular signals like neurotransmitters, peptide hormones, odorant molecules, and photons are primarily recognized by the receptors specifically identifying them. After complete amino acid sequences of bovine rhodopsin, the visual pigment, was determined in 1982 (Ovchinnikov et al., 1982), many receptors interacting with G proteins to send the information of the extracellular signals to intracellular second messengers has been identified. Although all these receptors have common structure of seven hydrophobic membrane spanning  $\alpha$ -helices, several types of such receptors have no amino acid sequence homology with rhodopsin. Two groups of genes, metabotropic glutamate receptors (Masu et al., 1991) and secretin receptor (Ishihara et al., 1991) and its related genes (Lin et al., 1991; Jüppner et al., 1991; Abou-Samra et al., 1992; Thorens, 1992), are thought to be the G protein coupled receptors having no sequence homology with rhodopsin family receptors and with the members of the other group. Yeast *STE2* and *STE3* (Nakayama et al., 1985) also have seven hydrophobic membrane spanning regions and interact with yeast G protein, *GPA1* (for review see Marsh et al., 1991), but have no homology with the members of other receptor groups and with each other. Bacteriorhodopsin (Ovchinnikov et al., 1979) and halorhodopsin (Blanck and Oesterhelt, 1987) of halophilic archaeobacteria (Archaea) also have seven transmembrane regions but no homology with the eukaryotic receptors. *Dictyostelium* cAMP receptor (Klein et al., 1988), which also contains seven transmembrane regions and interacts with G protein, have weak homology with rhodopsin family receptors. In any case, there is no evidence that all the receptors with seven transmembrane structures are the descendants of a common ancestral gene.

On this analysis, we aligned 124 amino acid sequences of opsins and receptors (Table 1) showing apparent homologies with rhodopsin (Fig. 1; Appendix A). *Dictyostelium* cAMP receptor was not included in this alignment

because of its weak homology with rhodopsin. From the alignment figure of consensus sequences of closely related groups (Fig. 1), highly homologous regions of the sequences were shown to be restricted to the seven transmembranes and the nearby regions.

On the basis of the alignment, every gene pair of evolutionary distances was calculated (Jukes and Cantor, 1969; Kimura, 1983) and the phylogenetic tree of the receptors (Fig. 2) was inferred by the neighbor-joining (NJ) method (Saitou and Nei, 1987). Because there was no information about the oldest gene duplication, the tree should be expressed as an unrooted one (Fig. 2(a)). From the phylogenetic tree, we found three important points.

First, receptors for the same ligand or structurally related ligands made clusters on the tree (Iwabe et al., 1989). For example, opsins (or visual pigments), olfactory receptors, and biogenic amine receptors made single clusters on the tree (Fig. 2(b)), respectively. The receptors for low molecular weight neurotransmitters including adenosine, cannabinoid, and biogenic amines are more closely related to each other than to neuropeptide receptors or peptide hormone receptors. These facts indicate that ligand candidates for so-called "orphan" receptors can be assumed when those receptors are included in the phylogenetic tree. Murphy et al. (1992) also pointed out the relationship between clusters of the receptors on their phylogenetic tree and the ligand specificities. The alignment of the consensus sequences of the receptors (Fig. 1), whose grouping of the members was based on the clusters of the phylogenetic tree (Fig. 2(b)), suggests the existence of subgroup specific amino acid sequences, which can be helpful to search the new member of the subgroup and to elucidate their functional specificities. In some cases, the receptors for same ligand consisting a cluster on the tree couple to different effector systems. For example, m1 and m2 muscarinic acetylcholine receptor subtypes make a cluster on the tree (Fig. 2(b)) but couple to different G protein subtypes, q and i, respectively. Opsins of

vertebrates couple to transducins (or G protein  $\alpha$  subtypes) but those of insects to  $\alpha$ q subtype (dgq). It seems that after the gene duplications or separation of species, changing the ligand specificities of the receptors was more difficult than changing the specificities for G protein coupling.

Second, on the phylogenetic tree, gene duplications producing the receptors for different ligand specificities occurred before the divergence of vertebrates and insects (and mollusks). For example, gene duplications of muscarinic acetylcholine receptors and histamine H-1 receptor and of opsins and the other receptors preceded the divergence of the two animal lineages (Fig. 2(b)). We will call these receptors of different ligand specificities "anisoforms". Mammalian tachykinin peptide receptor case, however, is an exceptional one because the duplication of substance K, substance P, and neuromedin K receptors occurred after the divergence of vertebrates and insects (Fig. 2(b)). The fact that these receptors can be cross-activated at reduced potency by the other neuropeptide (Nakanishi, 1991) indicates that they are now on the way to accepting the rigid ligand specificities. Because neuropeptides and peptide hormones might have frequently duplicated and diverged after the separation of vertebrates and insects, it seemed appropriate that some subtypes of the receptors became to get modified ligand specificities.

Third, gene duplications producing the receptors for same ligand specificity occurred independently in each lineage of vertebrates and insects (and mollusks). The genes diverged by these duplications have their own tissue or cell specific expressions. For example, color opsins and rhodopsins of vertebrates and opsin Rh1, 2, 3, and 4 of insects diverged independently after the divergence of the two animal lineages (Fig. 2(b)), indicating the independent acquisition of color visions of vertebrates and insects (Zuker et al., 1987; Iwabe et al., 1989; Fryxell and Meyerowitz, 1991) and possibly mollusks (Iwabe et al., 1989). These opsins are expressed in different visual

cells in both vertebrates and insects (Table 1). Vertebrates' muscarinic acetylcholine receptor subtypes m1, 2, 3, 4, and 5 also diverged after the separation of the two animal lineages and the tissue expressions of those subtypes are different to each other (Table 1; Bonner et al., 1987). We will call these receptor subtypes of same ligand specificity but of different tissue (or cell) distributions "tissue specific isoforms". In some cases, like m1 and m2 muscarinic acetylcholine receptor subtypes, G protein coupling specificity of one receptor subtype is different to that of others, as previously discussed. In vertebrate lineage, many of the gene duplications of tissue specific isoforms occurred before the divergence of fish and amphibians (Iwabe et al., 1993; Part 2). In the vertebrate opsin lineage, the evolutionary rate of amino acid substitutions before the separation of lamprey and amphibians (extending 600 myrs ago to 500 myrs ago (Dickerson, 1971)) is apparently higher than that after the separation of the two lineages (Fig. 2(b)). The tendency of high evolutionary rates of the first period is not restricted to opsins but can be seen in many gene families (Iwabe et al., 1993; Part 2).

In any case, the common ancestor of vertebrates and insects possessed a variety of "anisoform" receptors, although it had only one or a few "tissue specific isoform" receptors per one ligand.

Recently, Okano et al. reported that vertebrate rhodopsins have diverged from cone visual pigments (Okano et al., 1992). The phylogenetic tree inferred in this study also suggest the same result (Fig. 2(b)). Previously, Iwabe et al. reported that human blue opsin was closely related with rhodopsin than red and green opsins (Iwabe et al., 1989), while Yokoyama and Yokoyama suggested that blue and red/green opsins made a cluster (Yokoyama and Yokoyama, 1989). The genes of chicken green opsin (Okano et al., 1992) and gecko opsin P467 (Kojima et al., 1992) showed sufficient evidences that the genes for scotopic vision (rhodopsin) was evolved from that for photopic vision (color pigments).

Corrected nucleotide differences at the nonsynonymous and synonymous positions (Miyata and Yasunaga, 1980) of the receptor genes compared between human and mouse (or rat) were calculated for their common homologous regions (Table 2). The nonsynonymous substitution values ( $k_A$ ) were ranging from 0.00 to 0.14, whereas synonymous substitution values ( $k_S$ ) were from 0.24 to 0.93. The  $k_A$  values of thromboxane-A2 receptor, olfactory receptor I15, interleukin-8 receptor, thrombin receptor, and BK-2 bradykinin receptor were seemed to be larger than those of the receptors expressed mainly at brain (Table 1 and 2). Kuma et al. (1993) previously showed that the evolutionary rates of protein kinase genes expressed at immune systems are higher than those expressed at brain, which suggested that tissue specific genes are constrained against amino acid alternations not only from the structural or functional requirements of individual molecules but also from higher level as tissues or organs, where they are expressed. The fact that the receptors with higher  $k_A$  values are expressed in immune systems, blood, or peripheral nervous systems may be consistent with the results of Kuma et al. (1993) although it must be considered that the receptors with different ligand specificities have different functional constraints. On the receptors for same ligand, the  $k_A$  values varied to some extent. For example,  $k_A$  value of  $\beta_3$  adrenergic receptor, 0.052, is about 6 times higher than that of  $\alpha_2B$  adrenergic receptor, 0.0082. Such differences are expected as a consequence of the constraint from the tissues where receptors are expressed (Kuma et al., 1993), but the tendency that  $k_A$  values of the receptors coupling to G protein subtype i are smaller than that of the isoforms coupling to G protein subtype s or q may explain some part of the differences.

*G protein  $\alpha$  subunits* Heterotrimeric guanine nucleotide binding proteins (G proteins), consisting of  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits, transduce extracellular signals

received by the rhodopsin family receptors to intracellular signal amplifiers. The  $\alpha$  subunits, which catalyze GTP to GDP and mainly interact with the receptors and amplifiers, contain GTP binding domains homologous to those of other GTP binding proteins including ras, rab, rho, ADP ribosylation factor, polypeptide chain elongation factor Tu/1 $\alpha$  and G/2, and initiation factor 2.

Forty-five amino acid sequences of G protein  $\alpha$  subunits from various eukaryotes including yeast, plants, *Dictyostelium*, and animals (Table 3) were aligned for their almost whole regions (Fig. 3). On the basis of the alignment, the distant matrix was calculated and the molecular phylogenetic tree of the  $\alpha$  subunits (Fig. 4) was inferred by NJ method (Saitou and Nei, 1987). Because there was no information about the oldest gene duplication or separation of the species, the tree should be expressed as an unrooted one (Fig. 4(a)). From the phylogenetic tree, we found three important points about animal  $\alpha$  subunits, which is consistent with the three ideas about the rhodopsin family receptors, as is stated above. Recently, Yokoyama and Starmer (1992) also inferred phylogenetic trees of animal G protein  $\alpha$  subunits.

First, animal  $\alpha$  subunits interacting the same type of amplifiers made clusters on the tree. For example,  $\alpha_s$  and  $\alpha_{olf}$ , activating adenylyl cyclases, made a single cluster on the tree, whereas  $\alpha_{i1}$ , 2, and 3, inhibiting adenylyl cyclases, made another cluster (Fig. 4(b)).  $\alpha_q$ , y, 14, 15, and 16, recently probed to activate phospholipase C  $\beta$  subtypes (for review see Sternweis and Smrcka, 1992; Birnbaumer, 1992), also made a cluster (Fig. 4(b)). Two types of transducin  $\alpha$  subunits,  $\alpha_{t1}$  and  $\alpha_{t2}$ , activating phosphodiesterases on vertebrate photoreceptors, also made a cluster on the tree (Fig. 4(b)). These facts indicate that gustducin, which thought to transduce the signal of taste molecules to unknown effectors on taste receptor cells (McLaughlin et al., 1992), activates some type of phosphodiesterase because it belongs to the cluster of transducins (Fig. 4(b)). As *Caenorhabditis gpa* genes make a single independent cluster on the tree (Fig. 4(b)), there is a possibility that they

interact with an unknown amplifier protein. The alignment of the consensus sequences of the  $\alpha$  subunits (Fig. 3), whose grouping of the members was based on the clusters of the phylogenetic tree (Fig. 4(b)), suggests the existence of specific amino acid sequences for the subgroups which can be helpful to search the new member of the subgroup and to elucidate their functional specificities. There is no relationship between the clusters of the  $\alpha$  subunits and the relatedness of the receptors that interact them, which will be discussed later.

Second, gene duplications of the  $\alpha$  subunits of different amplifier interactions occurred before the divergence of vertebrates and insects (and mollusks). For example, gene duplications making  $\alpha_i$ , o, z, and t subtypes preceded the separation of the two animal lineages (Fig. 4(b)). The  $\alpha_s$  and  $\alpha_q$  subtypes also duplicated before the divergence. Thus these  $\alpha$  subunit subtypes of different amplifier interactions are "anisoforms", a term defined in the above discussion of the receptors. Although  $\alpha_q$  and 16, both activating phospholipase C  $\beta$  subtypes (Sternweis and Smrcka, 1992; Birnbaumer, 1992), made a cluster on the tree, the divergence of the two genes was before the separation of vertebrates and insects (Fig. 4(b)). This suggests that the two  $\alpha$  subtypes comprise some different functions.

Third, gene duplications of the  $\alpha$  subunits interacting the same amplifier occurred in vertebrate lineage and each duplicated gene has different tissue or cell specific expression. For example, the duplication of  $\alpha_s$ , expressed in many tissues, and  $\alpha_{olf}$ , expressed in olfactory sensory neurons, occurred after the divergence of vertebrates and insects (and mollusks) (Fig. 4(b)). Thus these  $\alpha$  subunits are "tissue specific isoforms". Furthermore, in vertebrate lineage, the gene duplications of tissue specific isoforms, like  $\alpha_s$  and  $\alpha_{olf}$  or  $\alpha_{i1}$ , 2, and 3 (Fig. 4(b)), occurred before the divergence of amphibians and mammals. In the vertebrate  $\alpha_s$  lineage, the evolutionary rate of amino acid substitutions before the separation of amphibians and reptiles (extending 600

myrs ago (Dickerson, 1971) to 350 myrs ago (Dayhoff, 1978)) is apparently higher than that after the separation of the two lineages (Fig. 4(b)). The tendency of higher evolutionary rates before the divergence of fish and amphibians than those after it can be seen also in many other genes (Iwabe et al., 1993; Part 2).

In the case of the G protein  $\alpha$  subunits, the common ancestor of vertebrates and insects possessed a variety of "anisoforms" but had only one (or a few) "tissue specific isoforms" (Fig. 4(b)), like the case of the receptors (Fig. 2(b)).

The mammalian gene of G protein  $\alpha$  subunit subtype o ( $\alpha_o$ ) produces two alternatively spliced mRNA encoding  $\alpha_o1$  and  $\alpha_o2$  that contain different last two exons (exons 7 and 8) (Tsukamoto et al., 1991). As the amino acid sequences of the exons 7 and 8 regions of  $\alpha_o1$  show strong homology with those of  $\alpha_o2$  and to other  $\alpha$  subunit subtypes, the sequences of these  $\alpha$  subunits were aligned (Appendix B'(a)). The phylogenetic tree of the  $\alpha_o$  subunit exons 7 and 8 regions inferred on the basis of the alignment (Appendix B'(b)) indicated that duplication of the two exons of  $\alpha_o1$  and  $\alpha_o2$  was occurred before the divergence of vertebrates and insects (and mollusks). Because the exons 7 and 8 of  $\alpha_o1$  were paralogous to the other animal  $\alpha_o$  subunits, we included human  $\alpha_o2$  in the phylogenetic tree (Fig. 4(b)). Yokoyama and Starmer (1992) included both  $\alpha_o1$  and  $\alpha_o2$  in their phylogenetic trees, which misled their results. There is a possibility that some functions of  $\alpha_o1$  and  $\alpha_o2$  are different because the duplication was before the separation of vertebrates and insects.

Although the root of the phylogenetic tree was not determined, no G protein  $\alpha$  subunit of fungi, plants, and *Dictyostelium* was closely related to animal  $\alpha_s$ ,  $\alpha_q$ ,  $\alpha_i$ ,  $\alpha_o$ ,  $\alpha_z$ , or  $\alpha_t$  (Fig. 4(b)). There is a possibility that  $\alpha$  subunit "anisoforms" regulating adenylyl cyclases, phosphodiesterases, and

phospholipase Cs diverged only in animal lineages. The evolutionary relationships of G proteins and the amplifiers will be discussed later.

Corrected nucleotide differences at the nonsynonymous and synonymous positions (Miyata and Yasunaga, 1980) of the G protein  $\alpha$  subunit genes compared between human and mouse (or rat) were calculated for their common homologous regions (Table 4). The  $k_A$  values were ranging from 0.00 to 0.065, whereas  $k_S$  values were from 0.29 to 0.97. Apparently the  $k_A$  value of  $\alpha_{16}$ , expressed specifically in myeloid cells (Table 3; Hepler and Gilman, 1992), was larger than those of the other  $\alpha$  subunit genes. This may be consistent with the results of Kuma et al. (1993) that the evolutionary rates of protein kinase genes expressed at immune systems are higher than those expressed at brain.

**Amplifiers** The extracellular signals received by the receptors are finally converted into the intracellular second messengers by signal amplifiers. There has been identified and sequenced several types of amplifiers: Adenylyl cyclase (AC), producing cAMP and regulating many effector proteins like A kinases or cAMP-gated channels; phosphodiesterase (PDE), hydrolyzing cGMP to GMP and closing the cGMP-gated channels; phospholipase C (PLC), producing two types of second messengers, inositol 1,4,5-trisphosphate (IP<sub>3</sub>) and diacylglycerol (DG) and regulating calmodulins and C-kinases. We inferred the molecular phylogenetic trees of these three amplifier families (Fig. 5, 6, and 7) by NJ method (Saitou and Nei, 1987).

Animal ACs comprise common homologous structure of two repeats of [6 transmembrane regions plus one catalytic domain]-structure and are regulated by G proteins. The phylogenetic tree of animal ACs (Fig. 5) was inferred by using *Dictyostelium* AC-A as an outgroup, which have overall homology with them. The guanylyl cyclases and yeast, *Trypanosoma*, and *Rhizobium* ACs also contain the homologous catalytic domains but are not

regulated by G proteins (for review see Tang and Gilman, 1992). They were not included in the tree (Fig. 5) because their structures except for the catalytic domains are completely different from those of animal ACs. The branching patterns of the phylogenetic tree of animal ACs, which was also inferred by maximum likelihood method (data not shown) (Kishino et al., 1990), indicates that the common ancestor of vertebrates and insects possessed at least four types of ACs, I and rutabaga, II and IV, III, and V and VI (Fig. 5). The results of rhodopsin family receptors and G protein  $\alpha$  subunits that the gene duplications before the divergence of vertebrates and insects (and mollusks) usually produced "anisoforms" suggest the functional differences of those four types of ACs. The fact that the effects of G protein  $\beta\gamma$  subunit complex and  $\text{Ca}^{2+}$ -calmodulin to these animal four types are different to each other (Tang and Gilman, 1992) is consistent with the above suggestion.

Figure 6 is the phylogenetic tree of PDE catalytic subunits. Mouse photoreceptor cGMP PDE  $\alpha$  and  $\beta$  subunits and bovine photoreceptor cGMP PDE are regulated by G protein  $\alpha$  subunits (or transducins), whereas other PDEs are regulated by other factors like cGMP or calmodulin (for review see Beavo and Reifsnyder, 1990). These PDEs regulated by different factors seems to have generated by gene duplications before the divergence of vertebrates and insects (Fig. 6). They can be defined as "anisoforms". On the other hand, four types of mammalian cAMP PDEs, whose functions are same but expressions are different to each other (Swinnen et al., 1989), had generated by gene duplications after the divergence of vertebrates and insects (Fig. 6). They can be defined as "tissue specific isoforms".

Three types of animal PLC,  $\beta$ ,  $\gamma$ , and  $\delta$ , apparently diverged before vertebrates and insects separated (Fig. 7). PLC $\beta$ ,  $\gamma$ , and  $\delta$  can be defined as "anisoforms" because they are regulated by G proteins, receptor type protein kinases, and unknown factors, respectively (for review see Rhee and Choi, 1992). Furthermore, because three PLC  $\beta$  subtypes, [mammalian  $\beta$ 1, 2, and

3], [*Drosophila plc-21*], and [bovine retina  $\beta$  and *Drosophila norpA*], also diverged before the separation of the two animal lineages (Fig. 7), some functions of the three subtypes are possibly different to each other, like the case of animal ACs.

There are four points that can be seen from the three phylogenetic trees of the amplifiers (Fig. 5, 6, and 7). First, genes with same or similar functions made clusters on the trees. Second, gene duplications generating "anisoforms", gene products of different functions, occurred before the divergence of vertebrates and insects. Third, gene duplications producing "tissue specific isoforms" occurred after the separation of the two animal lineages. Fourth, G protein coupled types of amplifiers are possibly generated by gene duplications of G protein non-coupled types before the separation of the two animal lineages (Fig. 6 and 7). Although the root of the trees should be determined to clarify the relationships of the G protein coupled and non-coupled types of amplifiers, the phylogenetic trees of G protein  $\alpha$  subunits (Fig. 4) and the amplifiers (Fig. 6 and 7) strongly suggest that the G protein coupled type amplifiers and G protein  $\alpha$  subunits regulating them were generated by the gene duplications in early animal lineages. If plants and fungi also have G protein coupled type amplifiers, it is possible that they produced the subtypes by gene duplications in their own lineages. In any case, to understand when these systems were established, the G protein  $\alpha$  subunit and amplifier genes of primitive animals like sponge, jellyfish, and planarian and of many plants and fungi will be very important.

#### *Diversification of the Signal Transduction System*

Figure 8 is the schematic phylogenetic relationships of the receptors and G protein  $\alpha$  subunits extracted from the trees of Figures 2(b) and 4(b). The figure indicates that there is little relationship between the phylogeny of receptors and that of G protein  $\alpha$  subunits from the view point of the coupling of the two factors.

Closely related receptors not necessarily couple to the same or closely related G protein  $\alpha$  subunits. Furthermore, in some cases, a receptor couples to more than two G proteins (Ross, 1989; Birnbaumer et al., 1990). After gene duplication, one of the duplicated receptors changed its coupling affinity to the G protein, which guaranteed the development of complex tissues or cell types that differently respond to the same ligand.

Gene duplications not only of the receptors but also of G protein  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits, adenylyl cyclases, phosphodiesterases, and phospholipases contributed the diversification of the signaling pathways (Ross, 1989; Birnbaumer et al., 1990). The possible combination of the signal pathways of the receptors, G protein subunits, and amplifiers known at present is more than hundred thousands although many of them have never been realized. Some signal pathways are listed on Figure 9. Many ligands and the receptors had been acquired when the unicellular animal ancestor became multicellular organism with many cell types, whereas the second messengers like cAMP (and/or cGMP), IP<sub>3</sub>, and DG were already used by the common ancestor of animals, fungi, and plants (Fig. 6 and 7). The G proteins combined the newly synthesized ligand perception systems with the preexisting second messenger regulating systems. The G proteins also directly regulate many ion channels, which were not discussed in this study (for review see Hepler and Gilman, 1992; Birnbaumer, 1992). The diversification of signal pathways by combinations of the duplicated gene products can be also seen in the protein kinase, phosphatase, and other systems.

*Concluding Remarks* As previously discussed, it seems very informative to pay attention to the divergence of vertebrates and insects (or mollusks) on the phylogenetic trees of signal transducing factor families. Gene duplications producing the proteins of different functions occurred before the divergence of the two animal lineages, whereas those producing the proteins of same

functions but different tissue distributions did after the divergence. This tendency is also seen in house keeping genes like peptide elongation factor EF-1 $\alpha$  and aldolase. Gene duplications of the latter case in the vertebrate lineage often happened before the divergence of fish and amphibians. During this period (from about 600 myrs ago to 400 myrs ago), molecular evolutionary rates not only of the genes of signal transduction system but also of EF-1 $\alpha$  and aldolase were higher than those after this time (from 400 myrs ago to now) (Iwabe et al., 1993; Part 2). This was probably related to the development of complex tissues of early chordates, and possibly of arthropods and other animal lineages, at the early Cambrian period (Iwabe et al., 1993; Part 2).

To clarify the detail processes of the diversification of signal transduction systems of eukaryotes, gene data from primitive animals, fungi, plants, and protista are very important.



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Table 1

## Expressions and Accession Numbers of Rhodopsin Family Receptors

No.	Gene	Expression	Accession No.
(A) opsin			
1.	human rhodopsin	rod cell	K02281
2.	chicken rhodopsin	rod cell	D00702
3.	lamprey rhodopsin	short photoreceptor cell	M63632
4.	chicken green	cone cell	M92038, M88178
5.	<i>Gecko</i> P467	rod cell	M92035
6.	chicken blue	cone cell	M92037
7.	human blue	cone cell	M13299
8.	chicken violet	cone cell	M92039
9.	human red	cone cell	M13305
10.	human green	cone cell	K03494
11.	chicken iodopsin (red)	cone cell	X57490
12.	<i>Astyanax</i> R007	(genomic)	M38630
13.	<i>Astyanax</i> G101	(genomic)	M38624
14.	<i>Astyanax</i> G103	(genomic)	M60945
15.	<i>Gecko</i> P521	rod cell	M92036
16.	octopus rhodopsin	visual cell	X07797
17.	<i>Loligo</i> rhodopsin	visual cell	S14332 <sup>a</sup>
18.	<i>Drosophila</i> Rh1	photoreceptor cell R1-6	K02315
19.	<i>Calliphora</i> Rh1	photoreceptor cell R1-6	M58334
20.	<i>Drosophila</i> Rh2	photoreceptor cell R8	M12896
21.	<i>Drosophila</i> Rh3	photoreceptor cell R7	M17718, Y00043
22.	<i>Drosophila</i> Rh4 (ultraviolet)	photoreceptor cell R7	b

Table 1 (continued)

No.	Gene	Expression	Accession No.
(B) olfactory (odorant) receptors (putative)			
23.	rat I3	olfactory sensory cell	M64385
24.	rat I8	olfactory sensory cell	M64387
25.	rat I4	olfactory sensory cell	M64391
26.	rat I9	olfactory sensory cell	M64388
27.	rat I15	olfactory sensory cell	M64392
28.	rat F5	olfactory sensory cell	M64377
29.	rat F12	olfactory sensory cell	M64381
30.	rat F3	olfactory sensory cell	M64376
31.	rat I7	olfactory sensory cell	M64386
32.	rat F6	olfactory sensory cell	M64378
33.	human HGMP07I (ligand unknown)	testis	X64994
34.	human HGMP07J (ligand unknown)	testis	X64995
35.	dog DTMT (ligand unknown)	testis (spermatocytes)	X64996
(C) pituitary glycoprotein hormone receptors			
36.	human luteinizing hormone- choriogonadotropic hormone (LH-CG) R.	testis, ovary <sup>c, d</sup>	M63108
37.	human follicle-stimulating hormone (FSH) R.	ovary	M65085
38.	human thyroid stimulatory hormone (TSH; thyrotropin) R.	thyroid	M32215

Table 1 (continued)

No.	Gene	Expression	Accession No.
(D) tachykinin peptide receptors			
(D-1) substance P receptor and others			
39.	human substance P R. (neurokinin 1 R.)	brain, alimentary canal <sup>e</sup> , lung, spinal cord	M74290, M81797, M84425, X65177
40.	human substance K R. (neurokinin 2 R.)	lung	M57414, M60284
41.	human neuromedin K R. (neurokinin 3 R.)	brain, kidney	M89473
42.	human putative opioid R. (ligand unknown)	placenta, brain	M84605
43.	<i>Drosophila</i> DTKR (tachykinin-like peptide R.)	head	X62711
44.	<i>Drosophila</i> NKD (tachykinin R.)	head > body	M77168
(D-2) glucocorticoid-induced receptor and PR4			
45.	mouse glucocorticoid-induced (GI) R. (ligand unknown)	T-cell	M80610
46.	<i>Drosophila</i> PR4 (neuropeptide R.)	head, body	M81490
(D-3) neuropeptide Y receptor			
47.	human neuropeptide Y R.	brain	M84755
(E) gastrointestinal hormone receptors			
48.	rat cholecystokinin A (CCK-A) R.	pancreas	M88096
49.	rat cholecystokinin B (CCK-B) R.	brain	M99418
50.	dog gastrin R.	gastric parietal cells, pancreas, brain	M87834

Table 1 (continued)

No.	Gene	Expression	Accession No.
(F) endotheline receptors			
51.	human endotheline-1 (ET-1) R.	arota, lung, atrium, colon, placenta	D90348
52.	human endotheline-B (ET-B) R.	brain > placenta, lung, kidney, adrenal, colon, duodenum	D90402, M74921
(G) bombesin-like peptide receptors			
53.	human gastrin-releasing peptide (GRP) R.	brain, colon, pancreas	M73481
54.	human neuromedin B R.	brain, esophagus <sup>f</sup>	M73482
(H) neurotensin receptor			
55.	rat neurotensin R.	brain > heart, duodenum, intestine, liver	JH0164 <sup>a</sup>
(I) posterior pituitary hormone receptors			
56.	human vasopressin V2 R. (antidiuretic hormone R.)	kidney	Z11687
57.	rat V1a arginine vasopressin R.	liver, kidney, spleen	Z11690
58.	human oxytocin R.	myometrium, mammary gland, non- pregnant endometrium, ovary	X64878
59.	mouse gonadotropin-releasing hormone (GNRH) R.	pituitary gland	M93108
(J) thyrotropin-releasing hormone receptor			
60.	mouse thyrotropin-releasing hormone (TRH) R.	pituitary gland	M94384, M59811

Table 1 (continued)

No.	Gene	Expression	Accession No.
(K) cannabinoid receptor related			
61.	human cannabinoid R.	brain	X54937
62.	human edg-1 (ligand unknown)	endothelial cell	M31210
63.	rat R334 (ligand unknown)	brain, testis	X61496
(L) adenosine receptors			
64.	rat A1 adenosine R.	brain, thyroid <sup>g</sup>	M64299, M69045
65.	human A2 adenosine R.	brain	M97370
66.	human A2b adenosine R.	brain	M97759
67.	rat A3 adenosine R.	testis > kidney, lung, heart > brain	M94152
(M) amine receptors			
(M-1) muscarinic acetylcholine receptors (mAChRs)			
68.	human m1 mAChR	brain <sup>h</sup>	X52068, M35128
69.	human m3 mAChR (hM4)	brain, pancreas	X15266
70.	human m5 mAChR	brain	M80333
71.	human m2 mAChR	brain, heart	M16404, X15264
72.	chicken m2 mAChR	brain, heart	M73217
73.	human m4 mAChR (hM3)	brain	X15265
74.	chicken m4 mAChR	brain, heart	J05218
75.	<i>Xenopus</i> m4 mAChR	ovary	X65865
76.	<i>Drosophila</i> mAChR	head	M23412
(M-2) histamine H1 receptor			
77.	bovine histamine H1 R.	lung, small intestine > adrenal medulla, uterus > brain, spleen	D10197

Table 1 (continued)

No.	Gene	Expression	Accession No.
(M-3) histamine H2 receptor			
78.	human histamine H2 R.	gastric fundus > brain <sup>i</sup>	M64799
(M-4) $\beta$ adrenergic receptors			
79.	human $\beta$ 1 adrenergic R.	brain, adipose tissue, heart, lung <sup>j</sup>	J03019
80.	turkey $\beta$ 1 adrenergic R.	erythrocyte	M14379
81.	human $\beta$ 2 adrenergic R.	lung > brain, heart, adipose tissue <sup>k</sup>	J02960
82.	human $\beta$ 3 adrenergic R.	adipocytes, liver, soleus muscle, ileum	M29932
(M-5) $\alpha$ 1 adrenergic receptors			
83.	human $\alpha$ 1A adrenergic R.	vas deferens, brain, heart, spleen <sup>l</sup>	M76446
84.	rat $\alpha$ 1B adrenergic R.	liver	M60655
85.	bovine novel $\alpha$ 1 adrenergic R.	brain	J05426
(M-6) $\alpha$ 2 adrenergic receptors			
86.	human $\alpha$ 2A (2C10) adrenergic R.	platelet	M18415
87.	human $\alpha$ 2B (2C4) adrenergic R.	brain, kidney	J03853
88.	human $\alpha$ 2C2 adrenergic R.	liver, kidney	M34041
(M-7) tyramine receptor			
89.	<i>Drosophila</i> tyramine R. (= octopamine R.)	head	X54794, M60789

Table 1 (continued)

No.	Gene	Expression	Accession No.
(M-8) serotonin (5HT) receptors (1A, 1B, 1D, 1E, dro, dro2A, dro2B)			
90.	human 5HT1A R. (G21)	brain <sup>m</sup>	X13556
91.	human 5HT1B R.	brain	M75128, M83180, M81590, D10995
92.	human 5HT1D R.	brain	M81589, M89955
93.	human 5HT1E R.	brain	M91467, Z11166
94.	<i>Drosophila</i> 5HT-dro R.	head	M55533
95.	<i>Drosophila</i> 5HT-dro2A R.	head	Z11489
96.	<i>Drosophila</i> 5HT-dro2B R.	head	Z11490
(M-9) serotonin (5HT) receptors (1C, 2, SRL)			
97.	human 5HT1C R.	brain <sup>n</sup>	M81778
98.	human 5HT2 R.	brain	M86841
99.	rat 5HT R. (SRL)	stomach fundus	X66842
(M-10) dopamine receptors (D1, D5)			
100.	human D1 dopamine R.	brain	X55760
101.	human D5 dopamine R.	brain	M67439, X58454
(M-11) dopamine receptors (D2, D3, D4)			
102.	human D2 dopamine R.	brain, retina	M30625, X51362; M29066
103.	<i>Xenopus</i> D2 dopamine R.	brain	X59500
104.	rat D3 dopamine R.	brain	X57764
105.	human D4 dopamine R.	brain	X58497

Table 1 (continued)

No.	Gene	Expression	Accession No.
(N) mas oncogene related receptors			
106.	human mas oncogene (angiotensin R.)	brain <sup>o</sup>	M13150
107.	human mrg (ligand unknown)	not detected	A39485 <sup>a</sup>
108.	rat RTA (ligand unknown)	brain, vas deferens, uterus, intestine, stomach, aorta	M35297
(O) arachidonic acid derivative receptors			
109.	human thromboxane-A2 (TXA2) R.	placenta, lung	S13647 <sup>a</sup>
110.	mouse prostaglandin E (PGE) R. EP3 subtype	kidney, uterus > brain, thymus, lung, heart, stomach, spleen	D10204
(P) chemoattractant peptide receptors			
111.	human FPRL1 (formylpeptide R. like)	granulocyte	M84562, X63819, M88107
112.	human N-formylpeptide R.	myeloid cell	M60627
113.	human RMLP-related R.I	leukocyte	M76673
114.	human C5a anaphylatoxin R.	myeloid cell series	M62505
115.	human vasoactiveintestinal peptide (VIP) R.	brain, colon, heart, kidney, lung, spleen, small intestine	M64749
116.	human angiotensin II-1 R.	heart, placenta, lung, liver, skeletal muscle, adrenal	M87290, M93394, M91464, Z11162
117.	human interleukin-8 (IL-8) R. (high affinity)	neutrophil	M68932
118.	human interleukin-8 (IL-8) R. (low affinity)	neutrophil	M73969
119.	human neuropeptide Y3 R.	spleen	M99293
120.	human platelet activating factor (PAF) R.	leukocyte > spleen, lung, kidney <sup>P</sup>	M76674
121.	human thrombin R.	platelet, vascular endothelial cell	M62424



**Table 1 (continued)**

No.	Gene	Expression	Accession No.
122.	human BK-2 bradykinin R.	kidney, uterus, lung > testis, pancreas, brain, heart	M88714
123.	human somatostatin 1 (SST1) R.	jejunum, stomach, colon, kidney	M81829
124.	human somatostatin 2 (SST2) R.	brain, kidney > jejunum, colon, liver	M81830

Note. - Sequence data are from GenBank release 77.0 and NBRF release 36.0 (data not from the banks are referred). Expression data were taken from the papers listed on the data banks. If not available, the expression data were from the papers of other mammalian receptors. > indicates that tissue expressions of the right are weaker than those of the left.

- a data from NBRF release 36.0
- b data from Montell et. al. (1987).
- c expressions of rat LH-CG receptor (McFarland et al., 1989).
- d expressions of pig LH-CG receptor (Loosfelt et al., 1989).
- e expressions of rat substance P receptor (Hershey et al., 1991).
- f expressions of rat neuromedin B receptor (Wada et al., 1991).
- g expressions of dog A1 adenosine receptor (Libert et al., 1989).
- h expressions of pig m1 mAChR (Kubo et al., 1986).
- i expressions of dog histamine H2 receptor (Gantz et al., 1991).
- j expressions of rat  $\beta$ 1 adrenergic receptor (Mussin et al., 1991).
- k expressions of rat  $\beta$ 2 adrenergic receptor (Mussin et al., 1991).
- l expressions of rat  $\alpha$ 1A adrenergic receptor (Lomasney et al., 1991).
- m expressions of rat 5HT1A receptor (Albert et al., 1990).
- n expressions of rat 5HT1C receptor (Julius et al., 1988).
- o expressions of rat mas oncogene (Young et al., 1988).
- p expressions of guinea-pig platelet activating factor receptor (Honda et al., 1991).

**Table 2**

Corrected nucleotide differences at the nonsynonymous and synonymous positions of rhodopsin family receptors that are compared between human and mouse (or rat)

Gene	$k_A$	$k_S$	Accession No.	
			Human	Mouse (or Rat)
thromboxane-A2 R.	0.14 ± 0.02	0.65 ± 0.06	a	D10849
interleukin-8 R.(low affinity)	0.13 ± 0.02	0.62 ± 0.06	M94582	L13239
odorant R. I15 <sup>b</sup>	0.096 ± 0.014	0.49 ± 0.05	X64994	M64392 <sup>c</sup>
thrombin R.	0.094 ± 0.014	0.85 ± 0.09	M62424	L03529
BK-2 bradykinin R.	0.082 ± 0.012	0.56 ± 0.06	M88714	M59967 <sup>c</sup>
$\beta$ 3 adrenergic R.	0.052 ± 0.010	0.49 ± 0.05	M29932	M74716 <sup>c</sup>
D4 dopamine R.	0.051 ± 0.010	0.35 ± 0.04	X58497	M84009 <sup>c</sup>
thyrotropin R.	0.046 ± 0.009	0.57 ± 0.06	M32215	M34842 <sup>c</sup>
A2b adenosine R.	0.045 ± 0.009	0.56 ± 0.06	M97759	M91466 <sup>c</sup>
histamine H2 R.	0.044 ± 0.009	0.49 ± 0.05	M64799	S57565 <sup>c</sup>
LH-CG R.	0.041 ± 0.009	0.72 ± 0.07	M63108	M81310
substance K R.	0.038 ± 0.008	0.58 ± 0.06	M57414	X62933
antidiuretic hormone R.	0.037 ± 0.008	0.60 ± 0.06	Z11687	Z11932 <sup>c</sup>
A2 adenosine R.	0.036 ± 0.008	0.55 ± 0.06	M97370	M91214 <sup>c</sup>
neuromedin B R.	0.036 ± 0.008	0.62 ± 0.06	M73482	c,d
$\beta$ 2 adrenergic R.	0.036 ± 0.008	0.62 ± 0.06	J02960	X15643
D5 dopamine R.	0.035 ± 0.008	0.65 ± 0.06	X58454	M69118 <sup>c</sup>
serotonin 1D R.	0.032 ± 0.008	0.61 ± 0.06	M89955	M89953 <sup>c</sup>
rhodopsin	0.029 ± 0.007	0.55 ± 0.06	K02281	M36695-9
$\alpha$ 2A adrenergic R.	0.028 ± 0.007	0.51 ± 0.05	M18415	M99377
angiotensin II-1 R.	0.022 ± 0.006	0.84 ± 0.09	M87290	X62295 <sup>c</sup>
D1 dopamine R.	0.021 ± 0.006	0.68 ± 0.07	X55760	M35077 <sup>c</sup>
follicle-stimulating hormone R.	0.020 ± 0.006	0.56 ± 0.06	M65085	c,e
gastrin-releasing peptide R.	0.019 ± 0.006	0.46 ± 0.05	M73481	M57922
serotonin 1B R.	0.018 ± 0.006	0.39 ± 0.04	D10995	M85151
serotonin 1C R.	0.018 ± 0.006	0.55 ± 0.06	M81778	f
endotheline-B R.	0.017 ± 0.006	0.52 ± 0.05	D90402	X57764 <sup>c</sup>
m1 mAChR	0.015 ± 0.005	0.39 ± 0.04	X52068	J04192
somatostatin R. 2	0.015 ± 0.005	0.58 ± 0.06	M81830	M81832
neuromedin K R.	0.015 ± 0.005	0.62 ± 0.06	M89473	J05189 <sup>c</sup>
neuropeptide Y R.	0.013 ± 0.005	0.93 ± 0.10	M84755	Z11504 <sup>c</sup>
m3 mAChR (hM4)	0.013 ± 0.005	0.53 ± 0.06	X15266	M16407 <sup>c</sup>

Table 2 (continued)

Gene	<i>k<sub>A</sub></i>	<i>k<sub>S</sub></i>	Accession No.	
			Human	Mouse (or Rat)
β1 adrenergic R.	0.012 ± 0.005	0.24 ± 0.03	J03019	J05561 <sup>c</sup>
substance P R.	0.012 ± 0.005	0.39 ± 0.04	M84425	X62934
D2 dopamine R.	0.012 ± 0.005	0.42 ± 0.04	X51362	X55674
α1A adrenergic R.	0.012 ± 0.005	0.62 ± 0.06	M76446	M60654 <sup>c</sup>
α2C2 adrenergic R.	0.011 ± 0.005	0.66 ± 0.06	M34041	L00979
angiotensin R. (mas)	0.011 ± 0.005	0.83 ± 0.09	M13150	J03823 <sup>c</sup>
m5 mAChR	0.011 ± 0.004	0.39 ± 0.04	M80333	M22926 <sup>c</sup>
serotonin 1A R.	0.0091 ± 0.0041	0.76 ± 0.08	X13556	J05276 <sup>c</sup>
serotonin 2 R.	0.0085 ± 0.0039	0.52 ± 0.05	M86841	X13971 <sup>c</sup>
α2B adrenergic R.	0.0082 ± 0.0039	0.54 ± 0.05	J03853	M99376
m4 mAChR (hM3)	0.0073 ± 0.0036	0.47 ± 0.05	X15265	M16409 <sup>c</sup>
cannabinoid R.	0.0054 ± 0.0031	0.45 ± 0.05	X54937	X55812 <sup>c</sup>
endotheline-1 R.	0.0033 ± 0.0023	0.93 ± 0.10	D90348	M60786 <sup>c</sup>
m2 mAChR	0.0018 ± 0.0018	0.60 ± 0.06	M16404	J03025 <sup>c</sup>
somatostatin R. 1	0.0	0.59 ± 0.06	M81829	M81831

Note - Corrected nucleotide differences at the nonsynonymous (*k<sub>A</sub>*) and synonymous (*k<sub>S</sub>*) were calculated (Miyata and Yasunaga, 1980) for the homologous regions of the sequences corresponding to the aligned sites of figure 1. The short regions between transmembrane 1 and 2, 2 and 3, 3 and 4, and 6 and 7 were included for the difference calculations. Abbreviations: R. = receptor, LH-CG = luteinizing hormone-choriogonadotropic hormone, mAChR = muscarinic acetylcholine receptor.

a Sequence data from Hirata et al. (1991).

b human HGMP071.

c rat data.

d Sequence data from Wada et al. (1991).

e Sequence data from Sprengel et al. (1990).

f Sequence data from Yu et al. (1991).

Table 3

## Expressions and Accession Numbers of G-protein α subunits

No.	Gene	Expression	Accession No.
(A) G <sub>s</sub>			
1.	human α <sub>s</sub> -1	ubiquitous <sup>a, b</sup>	M21142, X04408
2.	<i>Xenopus</i> α <sub>s</sub>	NI	X56091
3.	rat α <sub>olf</sub>	olfactory neuro-epithelium <sup>a, b</sup>	<sup>c</sup>
4.	<i>Drosophila</i> α <sub>s</sub> -S	head > body brain <sup>b</sup>	M33998, M23233
5.	<i>Lymnaea</i> α <sub>s</sub>	NI	Z15096
6.	<i>Schistosoma</i> α <sub>s</sub> (SG12)	NI	M81085
(B) G <sub>i</sub>			
7.	human α <sub>i</sub> 1	nearly ubiquitous <sup>a</sup> brain <sup>b</sup>	M17219
8.	<i>Xenopus</i> α <sub>i</sub> 1	NI	X56089
9.	human α <sub>i</sub> 3	nearly ubiquitous <sup>a, b</sup>	J03005, M20604, J03198; M27543, J03238
10.	<i>Xenopus laevis</i> α <sub>i</sub> 3	NI	X56090
11.	human α <sub>i</sub> 2	ubiquitous <sup>a, b</sup>	M20593, X04828
12.	<i>Asterina</i> α (SG)	NI	X66378
13.	<i>Lymnaea</i> α <sub>i</sub>	NI	Z15095
14.	<i>Drosophila</i> DGα1	embryo, pupa > adult head	M23094

Table 3 (continued)

No.	Gene	Expression	Accession No.
(C) Go			
15.	human $\alpha$ 2	brain <sup>a</sup> , b, others <sup>a</sup>	M60165
16.	<i>Xenopus laevis</i> $\alpha$ o	NI	X14636
17.	<i>Drosophila</i> $\alpha$ o (DGo2)	head > body	M86660, M29731, M30151, M29602,
18.	<i>Lymnaea</i> $\alpha$ o	NI	Z15094
19.	<i>Caenorhabditis</i> $\alpha$ o ( <i>goa-1</i> )	NI	M38251
(D) Gz			
20.	human $\alpha$ z	brain <sup>a</sup> , b, adrenal, platelets <sup>a</sup>	J03260
(E) Gt			
21.	human $\alpha$ t1	rod cell <sup>a</sup> , b	X15088
22.	human $\alpha$ t2	cone cell <sup>a</sup> , b	D10384
23.	rat $\alpha$ g ( <i>gustducin</i> , <i>gnat-3</i> )	taste bud <sup>a</sup>	X65747
(F) Gq			
24.	mouse $\alpha$ q	nearly ubiquitous <sup>a</sup>	M55412
25.	human $\alpha$ y	nearly ubiquitous <sup>a</sup>	M69013
26.	mouse $\alpha$ 14	bone marrow adherent cell lines (lung, kidney, liver) <sup>a</sup>	M80631
27.	<i>Drosophila</i> <i>dgq</i>	adult retina, ocellus	M58016

Table 3 (continued)

No.	Gene	Expression	Accession No.
(G) G16			
28.	human $\alpha$ 16	T cell, myeloid cell <sup>a</sup>	M63904
29.	mouse $\alpha$ 15	B cell, myeloid cell <sup>a</sup>	M80632
(H) G12			
30.	mouse $\alpha$ 12	ubiquitous <sup>a</sup>	M63659
31.	mouse $\alpha$ 13	ubiquitous <sup>a</sup>	M63660
32.	<i>Drosophila</i> <i>cta</i>	oocyte, nurse cell embryo, larva, adult	M63651, M94285
(I) others			
33.	<i>Dictyostelium</i> $\alpha$ 4	early mound stage >> vegetative growth, early development	A40990 <sup>d</sup>
34.	<i>Dictyostelium</i> $\alpha$ 1	loose aggregate formation >vegetative cell	M25060
35.	<i>Dictyostelium</i> $\alpha$ 2	during aggregation >> vegetative cell	M25061
36.	<i>Caenorhabditis</i> <i>gpa-1</i>	NI	e
37.	<i>Caenorhabditis</i> <i>gpa-2</i>	NI	X53156
38.	<i>Caenorhabditis</i> <i>gpa-3</i>	NI	M38250
39.	<i>Schizosaccharomyces</i> <i>gpa1</i>	NI	M64286
40.	<i>Coprinus</i> <i>CGP1</i>	NI	X68031
41.	<i>Saccharomyces</i> <i>GPA2</i>	NI	J03609
42.	<i>Saccharomyces</i> <i>GPA1</i>	NI	M15867
43.	<i>Candida</i> <i>CAG1</i>	mycelial and yeast cells	M88113
44.	<i>Arabidopsis</i> <i>GPA1</i>	vegetative plant tissue	M32887
45.	tomato <i>TGA1</i>	NI	M74419

**Table 3 (continued)**

Note. - Sequence data are from GenBank release 77.0 and NBRF release 36.0 (data not from the banks are referred). Expression data were taken from the papers listed on the data banks and (or) from Kaziro et al. (1991) and Hepler and Gilman (1992). > and >> indicate that tissue expressions of the right are weaker and much weaker than those of the left, respectively.

NI = not identified.

- a Expression data from Hepler and Gilman (1992).
- b Expression data from Kaziro et al. (1991).
- c Sequence data from Jones and Reed (1989).
- d Data from NBRF release 36.0.
- e Sequence data from Lochrie et al. (1991).

**Table 4**

**Corrected nucleotide differences at the nonsynonymous and synonymous positions of G-protein  $\alpha$  subunit that are compared between human and mouse (or rat)**

Gene	$k_A$	$k_S$	Accession No.	
			Human	Mouse (or Rat)
$\alpha_{16}^a$	$0.065 \pm 0.010$	$0.97 \pm 0.10$	M63904	M80632
$\alpha_{y}^b$	$0.019 \pm 0.005$	$0.67 \pm 0.07$	M69013	M55411
$\alpha_{t1}$	$0.011 \pm 0.004$	$0.81 \pm 0.08$	X15088	M25506-13
$\alpha_z$	$0.0077 \pm 0.0034$	$0.46 \pm 0.05$	J03260	J03773 <sup>c</sup>
$\alpha_{i2}$	$0.0076 \pm 0.0034$	$0.48 \pm 0.05$	X04828	M13963
$\alpha_{o2}$	$0.0061 \pm 0.0031$	$0.41 \pm 0.04$	M60156-62	M36778
$\alpha_{i3}$	$0.0060 \pm 0.0030$	$0.45 \pm 0.05$	M27543	M20713 <sup>c</sup>
$\alpha_{s-1}$	0.0	$0.29 \pm 0.03$	X04408	Y00703
$\alpha_{i1}$	0.0	$0.95 \pm 0.10$	M17219	M17527 <sup>c</sup>

Note - Corrected nucleotide differences at the nonsynonymous ( $k_A$ ) and synonymous ( $k_S$ ) were calculated (Miyata and Yasunaga, 1980) for the homologous regions of the sequences corresponding to the aligned sites of figure 3.

- a mouse  $\alpha_{15}$ .
- b mouse  $\alpha_{11}$ .
- c rat data.

## Figure Legends

**Figure 1.** Alignment of the consensus sequences of 35 groups of the rhodopsin family receptors. The gene numbers shown at the left of the sequences correspond to those of Table 1. In the parentheses, the shortest and longest amino acid intervals of the members of each group between the aligned segments and the shortest and longest amino acid lengths of the unaligned N- and C-terminal regions are shown. Above the alignment, the seven transmembrane regions (TM-1 to TM-7) assumed by the hydrophathy profiles (Kyte and Doolittle 1982) using the average hydrophilicity score of every aligned site are indicated by bars. - indicates not identical sites. Short region just after transmembrane 4 indicated by a double headed arrow under the alignment was not included for the construction of phylogenetic tree (Fig. 2). Complete alignment of the receptors are on Appendix A.

**Figure 2.** Phylogenetic tree of the rhodopsin family receptors. The tree was constructed by NJ method (Saitou and Nei, 1987) on the basis of the alignment of Fig. 1. Alignment length for tree construction was 195 amino acids. (a) Schematic unrooted tree of the receptors. The shaded polygons on the tree indicate the clusters of the receptors. Arrows point the separation of vertebrates and insects (or mollusks). (b) Rooted tree of the receptors. The root of the tree does not indicate the oldest time because there is no information on the tree about the oldest gene duplication or oldest separation of the organisms. The tree is essentially unrooted as (a). Filled circles point the separation of vertebrates and insects (or mollusks). The number at each node is a bootstrap probability that the members diverged from the node make a cluster, which has been estimated by a standard procedure with 1000 times resamplings (Felsenstein, 1985). A - P represent the group names of the receptors shown in Table 1. Accession numbers of the sequences are also listed on Table 1. Abbreviations: LHCG = luteinizing hormone-

choriogonadotropic hormone; FSH = follicle-stimulating hormone; TSH = thyroid stimulating hormone; TXA2 = thromboxane-A2; PGE = prostaglandine E; GRP = gastrin-releasing peptide; ET = endotheline; GNRH = gonadotropin-releasing hormone; VIP = vasoactiveintestinal peptide; IL-8 = interleukin-8; PAF = platelet activating factor; SST = somatostatin; GI = glucocorticoid-induced; CCK = cholecystokinin; TRH = thyrotropin-releasing hormone; mACh = muscarinic acetylcholine; adr. = adrenergic; dopa. = dopamine.

**Figure 3.** Alignment of the consensus sequences of 16 groups of the G-protein  $\alpha$  subunits. The gene numbers shown at the left of the sequences correspond to those of Table 3. In the parentheses, the shortest and longest amino acid intervals of the members of each group between the aligned segments and the shortest and longest amino acid lengths of the unaligned N- and C-terminal regions are shown. - indicates not identical site. Abbreviations: *D.d.* = *Dictyostelium discoideum*; *C.e.* = *Caenorhabditis elegans*; *S.p.* = *Schizosaccharomyces pombe*; *C.c.* = *Coprinus congregatus*; *S.c.* = *Saccharomyces cerevisiae*; *C.a.* = *Candida albicans*. Complete alignment of the G-protein  $\alpha$  subunits are on Appendix B.

**Figure 4.** Phylogenetic tree of the G-protein  $\alpha$  subunits. The tree was constructed by NJ method (Saitou and Nei, 1987) on the basis of the alignment of figure 3. Alignment length for tree construction was 277 amino acids. (a) Schematic unrooted tree of the  $\alpha$  subunits. The shaded polygons on the tree indicate the clusters of the  $\alpha$  subunits. Arrows point the separation of vertebrates and insects (or mollusks). Abbreviations: *D. d.* = *Dictyostelium discoideum*; *C. e.* = *Caenorhabditis elegans*. (b) Rooted tree of the  $\alpha$  subunits. The root of the tree does not indicate the oldest time because there is no information on the tree about the oldest gene duplication or oldest separation of the organisms. The tree is essentially unrooted as (a). Filled circles indicate

the separation of vertebrates and insects (or mollusks). The number at each node is a bootstrap probability that the members diverged from the node make a cluster, which has been estimated by a standard procedure with 1000 times resamplings (Felsenstein, 1985). Accession numbers of the sequences are listed on Table 3.

**Figure 5.** Phylogenetic tree of adenylyl cyclases. The tree was constructed by NJ method (Saitou and Nei, 1987) for the conserved regions (377 amino acid sites). Filled circle indicates the separation of vertebrates and insects. The number at each node is a bootstrap probability that the members diverged from the node make a cluster, which has been estimated by a standard procedure with 1000 times resamplings (Felsenstein, 1985). *Dictyostelium* AC-A was used as the outgroup of the tree (not shown on the tree). Sequence data are from GenBank Release 77.0. Accession numbers of the sequences are as follows: bovine I (M25579); rat II (M80550); rat III (M55075); rat IV (M80633); rat V (M96159); rat VI (M96160); *Drosophila* rutabaga (M81887); *Dictyostelium* AC-A (M87279). Data list and alignment of the adenylyl cyclases are on Appendix C1 and C2, respectively.

**Figure 6.** Phylogenetic tree of the phosphodiesterases. The tree was constructed by NJ method (Saitou and Nei, 1987) for the conserved regions (168 amino acid sites). Filled circle indicates the separation of vertebrates and insects. The number at each node is a bootstrap probability that the members diverged from the node make a cluster, which has been estimated by a standard procedure with 1000 times resamplings (Felsenstein, 1985). Abbreviations: cGS PDE = cGMP-stimulated phosphodiesterase; cGI PDE = cGMP-inhibited cAMP phosphodiesterase; CaM-PDE. = calmodulin-dependent phosphodiesterase. Sequence data are from GenBank Release 77.0 except for bovine CaM-PDE 61kDa which is from NBRF release 36.0.

Accession numbers of the sequences are as follows: mouse cGMP PDE $\alpha$  (X60664); mouse cGMP PDE $\beta$  (X55968); bovine cGMP PDE (M37838); bovine cGS PDE (M73512); rat CaM-PDE (M94537); bovine CaM-PDE 61kDa (NBRF, A40282); human cGI PDE (M91667); rat cAMP PDE1 (M25347); human cAMP PDE2 (M37744); rat cAMP PDE3 (M25349); rat cAMP PDE4 (J04563); *Drosophila dunce* (X55167); *Saccharomyces* cAMP PDE (M14563). Data list and alignment of the phosphodiesterases are on Appendix D1 and D2, respectively.

**Figure 7.** Phylogenetic tree of phospholipase Cs. The tree was constructed by NJ-method (Saitou and Nei, 1987) for the conserved regions (331 amino acid sites). Filled circle indicates the separation of vertebrates and insects. The number at each node is a bootstrap probability that the members diverged from the node make a cluster, which has been estimated by a standard procedure with 1000 times resamplings (Felsenstein, 1985). Sequence data are from GenBank Release 77.0 except for bovine  $\delta$ 2 (NBRF release 36.0). Accession numbers: human  $\gamma$ 1 (M34667); human  $\gamma$ 2 (X14034, M37238); rat  $\delta$ 1 (M20637); bovine  $\delta$ 2 (NBRF, S14113); rat  $\beta$ 1 (M20636); human  $\beta$ 2 (M95678); rat  $\beta$ 3 (M99567); *Drosophila plc-21* (M60452), bovine retina  $\beta$  (L13935); *Drosophila norpA* (J03138); *Saccharomyces PLC1* (D12738) *Dictyostelium* DdPLC (M95783). Data list and alignment of the phospholipase Cs are on Appendix E1 and E2, respectively.

**Figure 8.** Schematic phylogenetic trees of rhodopsin family receptors and G-protein  $\alpha$  subunits. Major couplings of the receptors and the G-proteins are indicated by arrows. In the G protein  $\alpha$  subunit tree, mammalian q subtype and the insect counterpart, dgq, are denoted as a branch of q. Tissue specific subtypes of G protein  $\alpha$  subunits are also represented as the branches of "anisoforms".

**Figure 9.** Signal pathways from receptors to effectors. Family members of rhodopsin family receptors, G protein  $\alpha$  subunits, adenylyl cyclases, phosphodiesterases, phospholipase Cs, cyclic nucleotide gated channels, and calcium channels are surrounded by squares, respectively. Abbreviations: R = receptor; G = G protein; AC = adenylyl cyclase; PDE = phosphodiesterase; PLC = phospholipase C; cAMP = cyclic AMP; cGMP = cyclic GMP; IP<sub>3</sub> = inositol 1,4,5-trisphosphate; DG = diacylglycerol; [+] = activation or increase; [-] = inhibition or decrease.

	Gene No.	TM-1	TM-2	
opsin	1- 8 ( 32- 44)	-----G-P-N-----T-----L (1)	--LNYIL-N----- (1)	
	9- 15 ( 48- 53)	-----M--VV-AS--TN-LV--A--KFKKL (1)	HPLNWIILVNLA--DL-ET--AS TIS--NQ-- (1)	
	16- 22 ( 34- 58)	-----GNG-V--F--KSL (1)	TP-N--NLA--D-----P----- (1)	
odorant R.	23- 35 ( 22- 35)	-----L--Y-----N--I-----L (1)	--PMY-FL--S-----L----- (1)	
pituitary glycoprotein hormone R.	36- 38 (359-414)	--LR--W--LA--GN--VL--L-TS-YK (1)	--VPRFLMCNL--FAD-C-G-YLL LIASVD--T (1)	
	39- 44 ( 32-101)	--W-----GN--V-WI-----M (1)	TVTN-F--NL-----N--N----- (1)	
tachikinin peptide R.	45- 46 ( 71- 90)	-----Y--F-L-GN--VC-----RM (1)	--T--FI--LA--DI-----P----- (1)	
	47 ( 40 )	FTLALAYGAVI ILGVSGLNALI I I I LKQKEM (1)	NVTN I L I VNLSFSDLLVA I MCL PLTFVYTLM (1)	
gastrointestinal hormone R.	48- 50 ( 54- 56)	A----LY--IFL-SV-GN-L-I-VL----R- (1)	TVTN-FLLSLAVSDL-L--CM PF-L-PNL- (1)	
endotheline R.	51- 52 ( 80-101)	YINTV-SC--F--G--GN-TLLRI IY-NKCM (1)	NGPN-LIASLALGDL--VID- P I NV-KLLA (1)	
bombesin-like peptid R.	53- 54 ( 40- 43)	-VIP--Y--I I--GL-GNI-L-KIF-T--M (1)	--VPN-FIS-LA-GDLLL--TC- PVDASRY-- (1)	
neurotensin R.	55 ( 64 )	VLVTA IYLALFVGTGNSVTAFTLARKKSL (4)	STVHYHLGSLALSDLL ILLLAM PVELYNFIW (4)	
posterior pituitary hormone R.	56- 58 ( 37- 51)	--E-A-L-----N--VL-AL----- (1-4)	-----F--HL--ADL-VA-FOV LPQL-W-- (1-4)	
	59 ( 35 )	K I RVTVTFFL FLLSTAFNASFLLKLOK WTQ (8)	SRMKVLLKHL TLANLLET L I VM PLDGMWNI T (8)	
thyrotropin-releasing hormone R.	60 ( 25 )	VVT ILLVVI I CGLGIVGNIMVVLVWRTKHM (1)	TPTNICYLVSLAVADLMVLVAAG LPNI TDSIY (1)	
cannabinoid R. and the related genes	61- 63 ( 44-116)	-----EN--V--I----- (1-2)	--P----IG-LA--DLL----- (1-2)	
adenosine R.	64- 67 ( 3- 14)	--Y--E--I-----GN-LV--V-----L (1)	--T--F-VSLA-AD-AVG--I P-AI----- (1)	
amine R.	muscarinic acetylcholine R.	68- 76 ( 23-104)	-----T--GN--V--S-K----L (1)	T--NY-L-SLA-AD--IG--SM --L----- (1)
	histamine H1 R.	77 ( 28 )	TPLVVVLS T I SLVTVGLNLLVLYAVRSERKL (1)	TVGNLY I VLSVADL I VGVVVM PMN ILYLLM (1)
	histamine H2 R.	78 ( 18 )	I T I TVLAVL I L I TVAGNVVCLAVGLNRRRL (1)	NLTNCF I VSLA I T D L L L G L L V L P F S A I Y Q L S (1)
	$\beta$ adrenergic R.	79- 82 ( 33- 58)	-----L-VL--V-GN-LVI-AI--RL (1)	T-TN-F--SLA-ADLV MGL-VV P--A----- (1)
	$\alpha$ 1 adrenergic R.	83- 85 ( 26- 54)	--G--L--IL--GN-LV ILSVAC-RHL (1)	--T-Y-IVNLA-ADLLL--TVL PFS A-E-L (1)
	$\alpha$ 2 adrenergic R.	86- 88 ( 12- 51)	-----L--T--GN-LV--AV-TSR-L (1)	APQNLFLVSLA-ADILVATL-- PFSLANE-- (1)
	tyramine R.	89 ( 109 )	LLTALVLSVI I VLT I I GNILV ILSVFYKPL (1)	I VQNF I VSLAVADL TVALLV L P F N V A Y S I L (1)
	serotonine R.	90- 96 ( 22-226)	-----L-----N--V-----L (1)	-----NYL--SLA--DL-V--LV- P--Y-- (1)
		97- 99 ( 53- 74)	--W-AL-----I I-TI-GN I LVI-AVS-EK-L (1)	--ATNYFLMSLA-AD-L-G-VM P--L-I-- (1)
	dopamine R.	100-101 ( 23- 40)	--TAC-L-LL I--TLLGN-LVCAA--R-RHL (2)	--TN-F--SLAVSDL-VA-LVM PWKAVAE-A (2)
102-105 ( 28- 34)		-----L-----GN-LVC--V--E-AL (1)	T-TN--VSLA-ADLL-A-LV- P--VY-EV- (1)	
mas oncogene and the related genes	106-108 ( 32- 75)	-----G--NG--W----- (0-1)	--P--Y--HL--AD--L----- (0-1)	
arachidonic acid derivative R.	109-110 ( 24- 25)	--S-F-----V-G--N-LA----- (5-6)	--SFL-----L-LTD--G-L-T--V----- (5-6)	
	111-114 ( 26- 37)	I-----V-F--GVLGN-LV-WV-F-- (1)	T--I--LNLA-ADF--LP----- (1)	
chemoattractant peptide R.	115-119 ( 28- 44)	-----Y--F-----N--V--V----- (1)	-----L-L--ADL--T-P--W----- (1)	
	120-121 ( 11-102)	--P-VY--FV--N--VF----- (1-3)	-----M--L--AD-LF--LP--I-YY-- (1-3)	
	122 ( 57 )	T I Q P P F L W L F V L A T L E N I F V L S V F C L H K S S (1)	T V A E I Y L G N L A A A D L I L A C G L P F W A I T I S N N (1)	
	123-124 ( 43- 58)	---FIY-VVC--GLCGN--VIYVILR YAK (2)	T-TNIYILNLA I ADEL-ML-P FL--L- (2)	
consensus		N		

**Figure 1**

Gene No.	TM-3	TM-4	TM-5
1- 8 ( 6 )	—C—E—F—T—G—W—S—L—A—E—R—V—C—K ( 10 )	HA—W—PP—G—W—S—R— ( 20 )	—E—F—F—P—I—F—Y—L—
9- 15 ( 6 )	HP—C—E—G—V—C—G—I—L—W—S—L—I—S—W—E—R—W—V—V—C—K ( 10 )	—A—G—I—F—W—W—W—P—P—I—F—G—W—S—R—Y ( 20 )	G—S—M—L—T—C—C—P—L—I—C—Y—V—A—I
16- 22 ( 6 )	—C—G—G—I—D—R—V—I— ( 10-11 )	—I—W—W— ( 18 )	—R—P—I—Y—I—V
23- 35 ( 6-10 )	—C—Q—LL—M—Y—D—R—A—C— ( 11 )	—L— ( 26-27 )	—S—Y—
36- 38 ( 13 )	G—G—C—A—G—F—F—T—V—F—A—S—E—L—S—V—Y—T—L—T—I—T—L—E—R—W— ( 11 )	HA—M—G—W—A—P—G—S—Y ( 13 )	—L—Y—L—L—N—A—F—C—C—Y—I—Y—V
39- 44 ( 6 )	—C—S—A—D—R—Y—A—I— ( 9 )	—I—W—P—Y— ( 17-26 )	—Y—L—P—Y—G—
45- 46 ( 6-7 )	—CH—Q—S—V—S—A—T—L—A—I—D—R—I—M— ( 9 )	—IA—W—A—L—P—I—L— ( 23 )	—Y—L—F—L—P—Y—A—R—
47 ( 6 )	E—A—M—C—K—L—N—P—F—V—Q—C—V—S—I—V—S—I—F—S—L—V—L—I—A—V—E—R—H—Q—L—I—N ( 9 )	H—A—Y—V—G—I—A—V—I—W—L—A—V—A—S—S—L—P—L—I—Y—Q—V—M—T ( 26 )	H—R—L—S—Y—T—L—L—L—V—L—Q—Y—F—G—P—L—C—F—I—F—I—C—Y—F—K—I—Y—I—R—L
48- 50 ( 6 )	—C—K—Y—M—G—S—V—S—V—S—T—L—V—A—I—L—E—R—Y—A—I—C—R ( 11-14 )	—Y— ( 17-18 )	—Q—W—L—L—L—L—F—P—G—V—V—A—Y—G—L—I—S—E—L
51- 52 ( 6-11 )	—C—K—L—P—F—O—K—S—V—G—I—T—V—L—L—C—A—L—S—D—R—Y—R—A—V—A—S ( 11 )	—T—A—E—I—V—I—W—S—L—A—P—E—A—I—G—F— ( 23-25 )	Y—K—D—W—M—L—F—F—Y—F—C—P—L—T—A—F—Y—T—L—M—T—C—E—M
53- 54 ( 6 )	—G—C—K—L—I—P—I—Q—L—T—S—V—G—V—S—V—F—T—L—A—S—A—D—R—Y—A—I—V— ( 11 )	—C—K—A—I—W—S—L—L—A—P—E—A—V—F—S— ( 22-23 )	H—P—K—I—H—S—F—L—V—I—P—L—I—S—Y—Y—Y—I—A—K—L
55 ( 8 )	D—A—G—C—R—G—Y—F—L—R—D—A—C—T—Y—A—T—A—L—N—V—A—S—L—S—V—E—R—Y—A—I—C—H ( 11 )	R—T—K—K—F—I—S—A—I—W—L—A—S—A—L—L—A—I—P—M—L—F—T—M—G—L—Q ( 21 )	T—V—K—V—V—I—Q—V—N—T—F—M—S—F—L—P—M—L—V—I—S—I—L—N—T—V—I—A—N—K—L
56- 58 ( 5-6 )	D—L—C—R—V—K—L—Q—M—A—S—Y—M—D—R—A—C— ( 10-12 )	—S— ( 13-21 )	G—Y—T—W—V—P—C—I—I—
59 ( 6 )	E—F—L—C—K—V—L—S—Y—L—K—L—F—S—M—Y—A—P—A—F—M—M—V—V—I—S—L—D—R—S—L—A—I—T—Q ( 9 )	L—E—O—S—M—I—S—L—A—W—I—L—S—I—V—F—A—G—P—O—L—Y—I—F—R—M—I ( 24 )	H—O—A—F—Y—N—F—F—T—F—G—C—L—F—I—P—L—L—I—M—L—C—N—A—K—I—F—A—L
60 ( 6 )	Y—V—G—L—C—I—T—Y—L—Q—Y—L—G—I—N—A—S—S—C—S—I—T—A—F—T—I—E—R—Y—A—I—C—H ( 11 )	R—A—K—K—I—I—F—V—W—A—F—T—S—I—Y—C—M—L—W—F—L—L—D—L—N ( 19 )	Y—Y—S—P—I—Y—L—M—D—F—G—V—F—Y—V—P—M—L—A—T—V—L—Y—G—F—A—R—I—L
61- 63 ( 2-5 )	—G—A—S—V—S—L—R—Y— ( 10-11 )	—W—L— ( 11-13 )	—L—
64- 67 ( 4 )	—C—L—C—L—T—S—I—L—L—A—A—D—R—Y— ( 11 )	R—W—G—L—T—P—G—W—N— ( 20-30 )	—Y—M—V—F—F—L—P—L—M—Y—F—
68- 76 ( 6 )	—C—D—W—L—A—D—Y—S—N—A—S—V—N—L—L—S—R—F—T— ( 11 )	—A—M—I—A—W—S—L—W—P—I—W— ( 16-17 )	—T—F—G—T—A—A—A—F—Y—P—V—M—L—Y—I—
77 ( 6 )	R—P—L—C—L—F—W—L—S—M—D—Y—V—A—S—T—A—S—I—F—S—V—F—I—L—C—I—D—R—Y—R—S—V—Q—Q ( 11 )	R—A—S—I—T—I—L—A—A—W—F—L—S—F—L—W—I—P—I—L—G—W—R—H—F ( 17 )	N—V—T—W—F—K—V—M—T—A—I—N—F—Y—L—P—T—L—L—M—L—W—F—Y—A—K—I—Y—K—A—V
78 ( 6 )	K—V—F—C—N—I—Y—T—S—L—D—V—M—L—C—T—A—S—I—L—N—L—F—M—I—S—L—D—R—Y—C—A—V—M—D ( 11 )	R—V—A—I—S—L—V—L—I—W—V—I—S—I—T—S—F—L—S—I—H—L—G—W—N—S—R ( 16 )	V—N—E—V—Y—G—L—V—D—G—L—V—T—F—Y—L—P—L—L—M—C—I—T—Y—Y—R—I—F—K—V—A
79- 82 ( 6 )	—C—E—W—T—S—D—V—L—C—V—T—A—S—I—E—T—L—C—A—D—R—Y—A—T— ( 11 )	—A—V—W—S—S—F—P—I—W—R— ( 18-19 )	—N—Y—S—S—S—F—Y—P—L—M—F—V—Y—R—V—A
83- 85 ( 6 )	—F—C—W—A—A—V—D—V—L—C—C—T—A—S—I—L—C—I—S—D—R—Y—G—V— ( 11 )	—L—W—V—S—G—P—L—G—W—P— ( 11 )	E—E—Y—F—S—S—F—Y—P—I—V—M—Y—C—R—V—Y—V—A
86- 88 ( 6 )	—W—C—Y—L—A—L—D—V—L—F—C—T—S—S—I—V—H—L—C—A—I—S—L—D—R—Y—W— ( 11 )	R—K—I—W—I—A—V—I—S—P—L— ( 12-16 )	—W—Y—S—I—G—S—F—F—A—P—C—L—I—M—L—V—Y—R—I—Y—A
89 ( 6 )	I—H—L—C—K—L—W—L—T—C—D—V—L—C—C—T—S—S—I—L—N—L—C—A—I—S—L—D—R—Y—W—A—I—T—D ( 11 )	R—V—L—L—L—I—S—G—V—L—L—S—L—L—I—S—S—P—L—I—G—W—N—D—W ( 13 )	S—O—R—G—Y—V—I—Y—S—S—L—G—S—F—F—I—P—L—A—I—M—T—I—V—Y—E—I—F—V—A—T
90- 96 ( 6 )	—C—D—C—C—T—S—I—L—L—I—D—R—Y—T— ( 10-11 )	—W—S—P— ( 12-14 )	—Y—T—F—Y—P—Y—I—A
97- 99 ( 7 )	—L—C—W—L—D—V—L—F—S—T—A—S—I—M—H—L—C—A—I—S—L—D—R—Y—A—I— ( 11 )	—A—K—I—V—W—I—S—G—P—P—G— ( 11-14 )	—F—L—G—S—F—F—P—L—T—I—M—T—Y—L—T—I—L
100-101 ( 6 )	—F—C—W—V—A—F—D—I—M—C—S—T—A—S—I—L—N—L—C—V—I—S—D—R—Y—W—A—I—S— ( 11 )	—A—A—W—T—L—S—L—S—F—I—P—V—Q—L—W—H— ( 23-37 )	L—R—T—Y—A—I—S—S—S—I—S—F—Y—I—P—V—A—I—M—I—V—T—Y—T—R—I—Y—R—I—A
102-105 ( 6-7 )	—C—D—D—V—M—C—T—A—S—I—N—L—C—A—I—S—D—R—A—V— ( 11-14 )	R—I—W—L—P—L—G—N— ( 7-10 )	—V—Y—S—S—S—F—P—L—Y—L—
106-108 ( 6-7 )	—L—L—A—I—S—E—R—C—V— ( 11 )	—S—V—C—L—W—L— ( 9-10 )	—I—L—
109-110 ( 10 )	—R—L—C—F—G—M—F—G—L—S—L—L—A—M—A—E—R—L—I— ( 10-11 )	R—A—V—W—L—A—L—L—P—L—G—V—G—R—Y ( 14-29 )	A—A—L—L—V—S—
111-114 ( 6 )	—C—N—S—L—I—D—R—V— ( 11 )	L—A—W—L—T—P— ( 18-29 )	—I—G—F—P—Y—I—
115-119 ( 4-6 )	—C—K—N—L—S—D—R—Y—L—I— ( 10-11 )	—W—P— ( 16-21 )	—G—P—Y—
120-121 ( 6 )	—L—C—F—N—Y—S—V—I—R—F—A—V— ( 11 )	R—L—I—W—T— ( 21-22 )	—F—F—I—C—I—R—L
122 ( 6 )	E—T—L—C—R—V—V—N—A—I—S—M—N—L—Y—S—I—C—F—L—M—L—V—S—I—D—R—Y—L—A—L—V—K ( 11 )	W—A—K—Y—S—L—V—I—W—G—C—T—L—L—L—S—S—P—M—L—V—F—R—T—M—K ( 20 )	W—E—V—F—T—N—M—L—L—N—V—V—G—F—L—L—P—L—S—V—I—F—T—C—T—M—Q—I—M—Q—V—L
123-124 ( 5 )	—C—R—V—V—D—N—F—T—S—I—C—L—T—V—S—D—R—Y—A—V—V—H ( 11 )	—A—K—V—W—S—L—L—V—I—L—P—I— ( 19 )	W—G—F—Y—T—F—G—F—L—P—I—C—L—C—Y—I—K—

Figure 1 (Continued)

Gene No.	TM-6	TM-7
1- 8 ( 15 )	A—E—V—M—V—F—P—Y—A— ( 9 )	—P—F—K—S—Y—N—P—Y—N—K— ( 37-44 )
9- 15 ( 15 )	A—E—V—R—M—V—V—M—A—C—W—G—P—Y—F—F—A— ( 9 )	—A—A—P—A—F—A—K—S—A—T—I—Y—N—P—Y—V—F—M—N—R— ( 17-39 )
16- 22 ( 27-29 )	A—E—A—K—W—P—Y—F— ( 8-9 )	—K—A—P—Y—S—H—P— ( 39-135 )
23- 35 ( 6 )	—F—T—C—S—H—L—V—Y— ( 7 )	—P—N—P—Y—L—R—N— ( 13-38 )
36- 38 ( 9 )	—D—T—I—A—K—M—A—L—I—F—T—D—F—C—M—A—P—I—S—F—A—S—A— ( 9 )	—K—L—L—V—L—F—P—N—S—C—A—N—P—F—L—Y—A—I—F—T—K— ( 64-81 )
39- 44 ( 18-19 )	—K—V—V—F—A—I—C—W—L—P—H—F— ( 12 )	—Y—L—W—L—A—M—S—M—N—P—Y—N— ( 86-136 )
45- 46 ( 19 )	—K—V—K—M—V—V—F—W—P—N—L—L—L— ( 8-11 )	—F—A—F—H—W—A—M—S—C—Y—N—P—Y—C—N— ( 73-76 )
47 ( 18 )	E—T—K—R—I—N—I—M—L—L—S—I—V—V—A—F—A—V—C—W—L—P—L—T—I—F—N—T—V—F—D—W ( 12 )	L—F—L—L—C—H—L—T—A—M—I—S—T—C—V—N—P—I—F—Y—G—F—L—N—K— ( 59 )
48- 50 ( 71-86 )	A—K—K—R—V—R—M—L—V—I—V—L—F—F—L—C—W—P—S—N—W—R—A— ( 12 )	P—I—S—F—I—L—L—S—Y—S—C—V—N—P—Y—C—F—M— ( 52-53 )
51- 52 ( 16-17 )	Q—R—R—E—V—A—K—T—V—F—C—L—V—F—A—L—C—W—P—L—H—L—S—R—I—L—K—T— ( 18 )	—D—Y—I—G—I—N—A—N—S—C—I—N—P—I—A—L—Y—V—S—K— ( 51-53 )
53- 54 ( 20 )	—R—K—R—L—A—K—V—L—V—F—V—G—F—F—C—W—P—N—H—Y—Y—R—S— ( 13 )	—A—R—L—F—N—S—C—V—N—P—F—A—L—Y—L—L—S— ( 57-61 )
55 ( 37 )	A—L—R—H—G—V—L—V—L—R—A—V—V—I—A—F—V—V—C—W—L—P—Y—H—V—R—L—M—F—C—Y ( 16 )	F—Y—M—L—T—N—A—L—F—Y—V—S—S—A—I—N—P—I—L—Y—N—L—V—S—A— ( 50 )
56- 58 ( 32-45 )	A—T—V—M—T—I—V—C—W—P—F—V—Q—W—W— ( 9-12 )	—L—L—A—S—L—N—S—C—N—P—W—I—Y—F— ( 36-55 )
59 ( 22 )	A—R—L—R—T—L—K—M—T—V—A—F—A—T—S—F—V—V—C—W—T—P—Y—Y—V—L—G—I—W—Y—W—F ( 11 )	V—N—H—F—F—L—F—A—F—L—N—P—C—F—D—P—L—I—Y—G—Y—F—S—L— ( 0 )
60 ( 41 )	S—R—K—O—V—T—K—M—L—A—V—V—V—I—L—F—A—L—L—W—M—P—Y—R—T—L—V—V—N—S—F ( 9 )	F—L—L—F—C—R—I—C—I—Y—L—N—S—A—I—N—P—V—I—N—L—M—S—Q— ( 68 )
61- 63 ( 16-35 )	—T—L—C—W—P— ( 6-10 )	—N—S—N—P—Y— ( 15-70 )
64- 67 ( 18-22 )	—E—A—K—S—L—F—A—L—W—L—P—N—F— ( 7-11 )	—I—L—H—N—S—N—P—Y—A— ( 30-119 )
68- 76 ( 143-398 )	—E—K—T—A—I—L—L—F—I—T—W—P—Y—N—V—L— ( 8-10 )	—W—Y—L—C—Y—N—S—T—N—P—C—Y—A—L—C—N— ( 21-41 )
77 ( 194 )	R—E—R—K—A—A—K—Q—L—G—F—I—M—A—A—F—I—C—W—I—P—Y—F—I—F—M—V—I—A—F ( 8 )	V—H—M—F—T—I—W—L—G—Y—I—N—S—T—L—N—P—L—I—Y—P—L—C—N—E— ( 14 )
78 ( 18 )	R—E—H—K—A—T—V—T—L—A—A—V—M—G—A—F—I—C—W—F—P—Y—T—A—F—V—Y—R—G—L ( 9 )	L—E—A—I—V—L—W—L—G—Y—A—N—S—A—N—P—I—Y—A—A—L—N—R— ( 66 )
79- 82 ( 40-66 )	—E—A—L—T—L—G—I—M—G—F—T—L—C—W—L—P—F—N— ( 8-9 )	—N—W—G—Y—N—S—F—N—P—Y—C—R—S—P— ( 52-137 )
83- 85 ( 54-60 )	R—E—K—K—A—A—K—T—L—V—V—G—F—L—C—W—P—F—P—G—S— ( 9 )	V—F—K—F—W—L—G—Y—N—S—C—N—P—Y—P—C—S—S— ( 135-162 )
86- 88 ( 137-165 )	—E—K—R—F—T—F—V—L—A—V—V—G—V—F—V—C—W—F—P—F—F—Y—L— ( 7-10 )	L—F—F—F—W—G—Y—C—N—S—L—N—P—V—I—Y—T—F—N— ( 19-20 )
89 ( 226-226 )	K—E—R—R—A—R—T—L—G—I—M—G—V—F—V—I—C—W—L—P—F—L—M—Y—V—I—L—P—F ( 9 )	F—K—N—F—I—T—W—L—G—Y—I—N—S—G—L—N—P—V—I—Y—T—F—N—L— ( 13 )
90- 96 ( 70-331 )	—E—K—L—I—F—W—L—P—F—L— ( 8-10 )	—W—L—G—Y—N—S—N—P—Y— ( 16-52 )
97- 99 ( 55-73 )	N—E—A—K—V—L—G—I—V—F—M—M—C—P—F—I—T—N— ( 11-12 )	L—L—F—V—W—G—Y—S—N—P—L—Y—T—L—F—N—K— ( 85-95 )
100-101 ( 37-44 )	—E—T—K—V—L—K—T—L—S—V—I—M—G—V—F—V—C—W—L—P—F—I—L—N—C—P—F ( 14-18 )	T—F—D—V—F—V—W—F—G—W—A—N—S—S—L—N—P—I—Y—A—F—N—A— ( 111-114 )
102-105 ( 88-155 )	—E—K—A—V—G—F—C—W—P—F—H— ( 8-9 )	L—A—T—W—L—G—Y—N—S—A—N—P—Y—T—F—N— ( 12-14 )
106-108 ( 0-6 )	—L— ( 6-7 )	—L—I—N—S—S—A—P—Y—F—G— ( 40-58 )
109-110 ( 15-18 )	—E—Q—L—G—I—M—V—S—V—C—W—P—L—L— ( 18-19 )	—L—I—R—A—N—Q—I—L—D—P—W—Y—L—R— ( 30-39 )
111-114 ( 6 )	—S—L—V—V—A—F—W—P— ( 13-16 )	—A—N—C—N—P—Y—V—G— ( 44-46 )
115-119 ( 6-11 )	—F—W—P—D— ( 17-18 )	—T—C—N—P—Y—F— ( 40-52 )
120-121 ( 9-12 )	—K—R—A—L—V—F—I—I—C—F—P—V— ( 13-16 )	A—C—S—D—P—Y— ( 44-49 )
122 ( 12 )	T—E—R—R—A—T—V—L—V—L—L—L—F—I—C—W—L—P—F—Q—I—S—T—F—L—D—T—L ( 17 )	I—T—Q—I—A—S—F—M—A—Y—S—N—S—C—L—N—P—L—V—Y—V—I—V—G—K— ( 54 )
123-124 ( 14 )	S—E—K—T—M—V—V—V—F—C—W—P—F—Y—V— ( 7-11 )	—V—L—Y—A—N—S—C—A—N—P—I—Y—F—L—S—D— ( 52-63 )

Figure 1 (Continued)



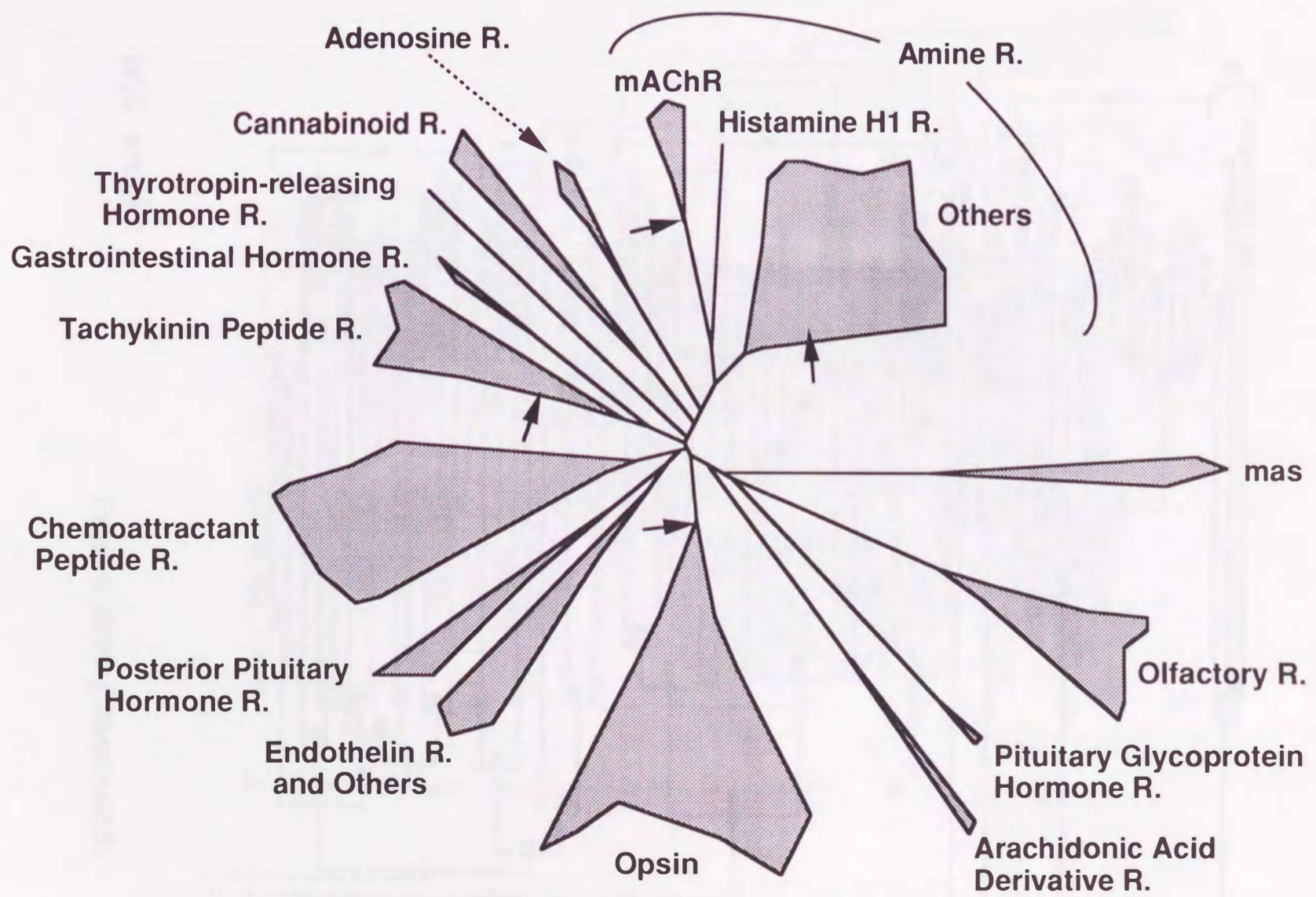


Figure 2(a)

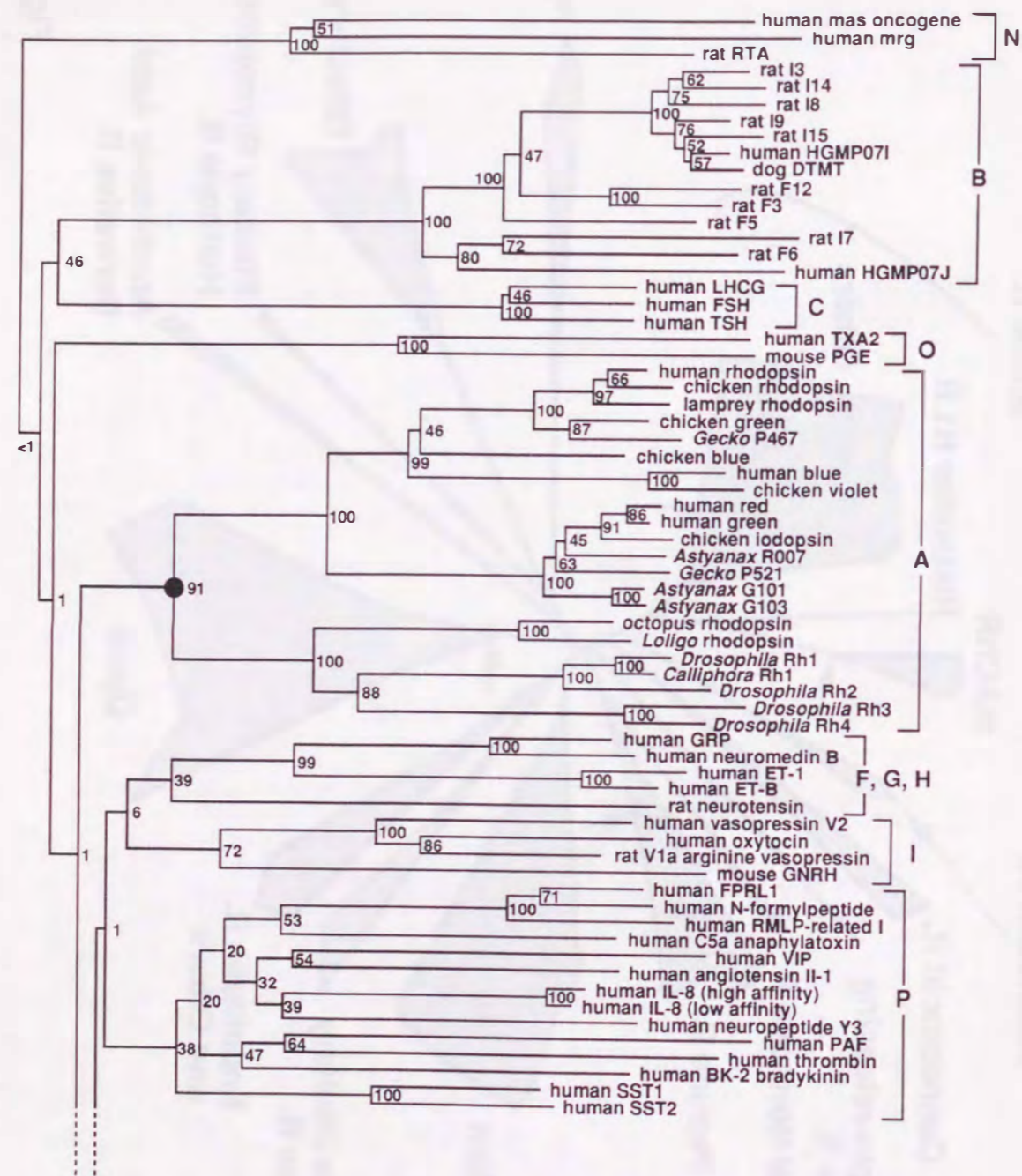


Figure 2(b)

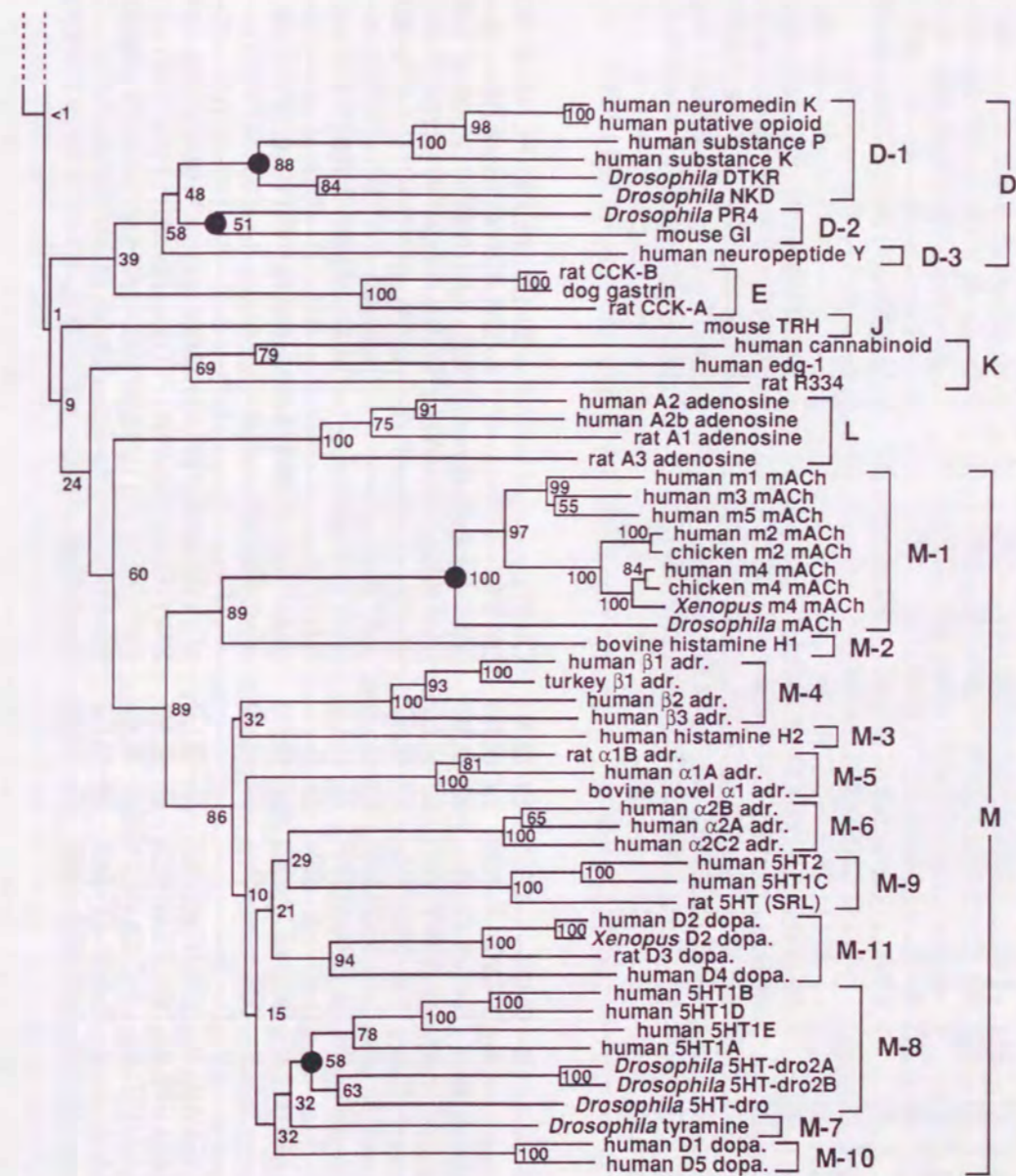


Figure 2(b) (Continued)

Gene No.

Gs	1- 6	( 35- 41)	-THRLLLGAGESGKS-IVKQMR LH-	-GF---	(0-15)	E---K---I---N---A ---I---AM---
Gi	7-14	( 22- 32)	-EVKLLLLGAGESGKST VKQMK IH-	-GYS--	( 0 )	-C---Y---VV-SN--QS---I---AMG-L
Go	15-19	( 31 )	KD-KLLLLGAGESGKST VKQMK IHE	-GF-E	( 0 )	D-KQY-PVVYSNT-QSL-AI-RAM--L
Gz	20	( 31 )	REIKLLLLGTSNSGKST VKQMK IHS	GGFNLE	( 0 )	ACKEYKPLI YNAIDSLTRI RALAAL
Gt	21-23	( 27- 31)	-TVKLLLLGAGESGKST VKQMK IH-	-GYS--	( 0 )	EC-EF-A---Y-N-LQSLA I---AMTTL
Gq	24-27	( 31- 37)	RELKLLLLGTGESGKSTF KQMR IHG	-GYS-E	( 0 )	D-G---KLV-QNIF-AMQ-MI-AM--L
G16	28-29	( 40 )	-ELKLLLLGPGESGKSTF KQMR IHG	-GYSEE	( 0 )	-R--FR-L-YQNFVSM-AMI-AM-RL
G12	30-32	( 46-130)	R-VK-LLL GAGESGKSTFLKQMR IHG	--FD--	( 0 )	---E---I---N---G---VL-DAR-KL
<i>D.d.</i> $\alpha$ 4	33	( 29 )	KDVKLLLLGPGESGKST FKQMK IQEDGGYSVE	( 0 )	( 0 )	ELLEYRAFVYSNC SOMEALLTASAKL
<i>D.d.</i> $\alpha$ 1,2	34-35	( 29- 34)	-E-KLLLLGAGESGKST KQMK IH-	-G---E	( 0 )	E---K-I-I---N---MRVL-----L
<i>C.e.</i> <i>gpa</i>	36-38	( 31 )	---KLLLLGAGE-GKSTVLKQMK-----	-----E	( 0 )	E-----V-N---M-----M---
<i>S.p.</i> <i>gpa1</i>	39	( 72 )	NDIKVLLLGAGDSGKTT MKQMRLLYS	PGFSQV	( 0 )	VRKQYRVM FENI SSLCLEAMDNS
<i>C.c.</i> <i>CGP1</i>	40	( 32 )	NEIKMLLLGAGESGKSTVLKQMK IHH	GGYSDQ	( 0 )	EKDSYKE IFSNTVQSMRA LDALPAL
<i>S.c.</i> <i>GPA2</i>	41	( 121 )	KELKVLLLGAGESGKSTVLQQLK ILHQ	NGFSEQ	( 0 )	E KEYIPLIYQNLLE GRNL QARTRF
<i>S.c.</i> <i>GPA1</i> , <i>C.a.</i> <i>CAG1</i>	42-43	( 39 )	---KLLLLGAGESGKSTVLKQLLLH-	GGF--Q	( 0 )	ER-QY---VIW-D-IQSMK-LI QARKL
Plants	44-45	( 36- 37)	HI-KLLLLGAG-SGKST FKQ KLLFQ	TGFDE-	( 0 )	ELK-Y-PVIHANVYQT-K-LHDG-KE-
consensus			LLL G GK Q			

Gene No.

1- 6	( 32 )	F-----LW- D-G-----ERSN	EY QLID	CA-YFL-----Y-P--QD-LRCRVLTS	GIFET-F-V	DKVN
7-14	( 31- 32)	-----LW- D-GVQ--F-RSR	EY QLND	SA-YYL----R-----YIPT-QDVLTRVKT	TGI-ETHF--	K-L-
15-19	( 32 )	LL--M-RLW- D-G-Q-C--R--	EY QLND	SAKY-LD-L-R-G---Y-PTEQ-ILRTRVKT	TGIVE-HF-F	KNL-
20	( 32 )	LLGVMRRLWA DPGAQACFSRSS	EY HLED	NAAYYLNDLER AAADY IPTVED ILRSRDM	TTGIVENKFTF	KELT
21-23	( 31 )	---I-RLW- D-G-QACFERA-	EY QLND	SA-YYL--L-R---P-Y-P-EQDVL-SRVK	TGI IET-FS-	KDLN
24-27	( 30 )	---AIK-LW- D-GIQECYDRRR	EY QL-D	S-KYYL-D-R-----PT-QD-LR-RVPTT	GI-EYPFDL	---
28-29	( 30 )	YA-AMQ-LWR DAGIRACYERRR	EF HLLD	SAVYYLSHLERI-E--Y-PTAODVLRSRM	PTTGINEYCFSV	-KT-
30-32	( 31- 38)	Y-P---LW- D-GI--A--RR-	EF Q---	SV-YFLD-----Y-P---DIL--R--TKG--	E-----	---P
33	( 27 )	LAADIKHLWE DKGIKETYAQKD	KHF QLND	SAAYFFDN DRYMRED FVPEQDVLRCR	VRTTGIQESEFTF	DKIR
34-35	( 30- 36)	---IKALW- DPG-----R--	EF--L-D	SA-Y-FDSIDR---P-Y-P---D-L--R--T-G--	ET-FEI	----
36-38	( 31- 32)	-----LW- D-V-----	---	F-----R---Y-P---D-L--R--T-G--	EV-F--	K---
39	( 30 )	IYEAHALTL DTKLRTVQSCGT	NL SLLD	NFYQQDH DRIFDPQY IPSDOD LHCRIK	TTGISEETFL	NRHH
40	( 30 )	IADAIQLWA DPGLKEAVRRSR	EF QLND	SAVYYFNS DRMSAPGYLPTDQD ILRSR	VKTG TETTFKV	GELT
41	( 34 )	IAGVISTLWA LPSTQDLVNGPNASKF	YLMD	STPYFMENFTRI TSPNYRPTQD ILRSRQ	MTSG FDTV DMGSDIK	
42-43	( 98-141)	IA-AI--LW--D-GIK-CF-RSN	EF QLEG	SA-YYFDN--FA--NY--TD-DILKGR	IKTTG TET-F-I	-S-
44-45	( 38 )	---IE-LWK DPAIQET--RGN	EL QVPD	C--Y-MENL-R-SD--YIPTKEDVL-AR-RT	TGVVEIQFSPVGENK	
consensus		L		L R T G		

Figure 3

Gene No.

1- 6	( 0 )	FHMFVGGQR-ERRKWIQCFN	DVTA I-V-A-S-YNMV-RED--NR-E-L-L--SIWNNRWLR-IS-ILFLNKQD-L-EK--A
7-14	( 0 )	FK-FDVGGQSRERKKWIHCFE	GVTAI IF-VA-S-YDL-LAED-EMNRM-ES-KLFDS CN-KWF--TSI LFLNKKDLFEKI
15-19	( 0 )	F-LFDVGGQSRERKKWIHCFE	DVTA IFCVA--YDQ-LHEDETTNRM-ESLKLFDSD CNNKWF-TSI LFLNKKD-F-EKI
20	( 0 )	FKMVDVGGQSRERKKWIHCFE	GVTAI FCVELSGYDLKLYEDNQTSRMAESLRLFDS CNNNWF INTSL LFLNKKD LLAEKI
21-23	( 0 )	FRMFVGGQSRERKKWIHCFE	GVT C IF-AAL-AYDMVLVED-EVNRMHESLHLFNS CNH--FA-TS VFLNKKD-F-EK-
24-27	( 0 )	FRMVDVGGQSRERKKWIHCFE	-VTSI-FLVALSEYDQ-L-E-DNENRMEESKALFRTI ITYPWF-NSSVI LFLNKKD LLE-KI
28-29	( 0 )	LRIVDVGGQ-SER-KWIHCFE	NVIALIYLASLSEYDQCLEEN-QENRM-ESLALF-TILELPWFKSTSV LFLNKD ILE-KI
30-32	( 0 )	F--VDVGGQ--R--W--CFD--	TSI-F-VSSSE-DQVL-EDR-TNRL-ES-NIF-TIVNN--F--SI LFLNK-DLL--KV--
33	( 0 )	LK VDVGGQSRERKKWIHCFD	CVTAV FVAAMSDYDQVLREDES VNRTRTRESLALFKE VNCDFYK ETP VFLNKKD LFEKEL
34-35	( 0 )	FR--VDVGGQSRERKKW--CF-	-VTAV-FCVALSEYDL-LYED--TNRM-ESL--F--CN--WF-NT-- LFLNK-D-F-EKI
36-38	( 0 )	FRV-DVGGQSR-RKKWIHCFE	D--A-I--A--SEY-VL-ED-TNRM-ES-LF--N--F-NT-- LFLNK-DLF-EKI--
39	( 0 )	YRFFDVGGQSRERKKWIHCFE	NVTALLFLVSLAGYDQCLVEDNSGNQMQEALLLWDS CNSSWFSESAMI LFLNKD LDFKRG
40	( 0 )	YKLFVGGQSRERKKWIHCFE	NVTALVFLVSLSEYDQMLYEDES VNRMQEALTLFDS CNSRWFVKT SI LFLNK DLFAEKL
41	( 0 )	MHIYDVGGQSRERKKWIHCFD	NVTLV FCVSLSEYDQTLMEDKNONRFQESLVLFDN VNSRWFARTSVV LFLNK DLFAEKL
42-43	( 0 )	FKVLDAGGQSR-RKKWIHCFE	-ITAVLFLVA-SEYDQ-LFEDERVNRMHESI-LFD-L-NSKWF--TPFI LFLNK D-FE-K-
44-45	( 5 )	YRLFVGGQSRNERRKWIHLFE	GVTAV FCAA SEYDQTLFEDE-KNRMETKELF-WVLKQPCFEKTSFML LFLNKFD FE-KV
consensus		D GGQ W F	E E LFLNK D K

Gene No.

1- 6	GKSK--	-YF-----Y--P (18-21)	F-RD-FL-	( 0 )	-----G-HYCYPH-TCAVDTENI-RVF-DCRD IORMHL (6)
7-14	--SPLT	IC-PEY-G-N-	( 5 )	YI---FE-	( 0 ) LN--KD-KEIY- H-TCATDT-N--FVFDVAVTDV IK-NL (6)
15-19	--SPLT	ICFPEY-G----	( 5 )	--Q-Q-E-	( 0 ) -NKS-- KEIY- H-TCATDT-N QFVFDVAVTDV IA-NL (6)
20	RR PLT	ICFPEYKQNTY	( 5 )	YIQRQFED	( 0 ) LNRNKETKEIYS HFTCATDTSN QFVFDVAVTDV IQNNL (6)
21-23	-K-HLS	ICFP-Y-G-N--	( 5 )	YIK-QFL-	( 0 ) LN-----KEIYS HMTCATDTQNVKF-FDAVTD I KENL (6)
24-27	-YSHL-	-YFPE--GP----	( 6 )	F-LK----	( 0 ) -NPD-- --YS HFT-ATDTENI--VF-AVKDT I-Q--L (6)
28-29	-TSHLA	TYFPSFQGP--D	( 6 )	FILDMY-R	( 12 ) -KG-R- RR-F- H-TCATDTQ--R-VFKDVRDSVLARYL (6)
30-32	----I-	----F-G-PH-	( 6 )	-----F-	( 0 ) -RR-----H HFTTAI-T-NI--VF--VKDT I-L--NL (6)
33	KRVPLQ	SCFSDYTGPNKY	( 5 )	FIQSOYLA	( 0 ) QGPSP RTIYT HATCAVDTEN KFVFRAVRQT ILSQAL (3)
34-35	-----	-F-EY-G---Y	( 5 )	-IK--F--	( 0 ) -N---- K-IY- H-TCATDTNNI--VVF-AVKDI----- (7)
36-38	K-----	-AF----G----	( 5- 6)	----K---	( 0 ) ---N-- K-Y- H-TCATDT-QVQ--LD-V---I----L (6)
39	SHFPIQ	KHFDPYQEVGST	( 20 )	YFYLFKES	( 0 ) LNRIAS RSCYC HFTTATDTSLLQRMVSVQDT MSNNL (5)
40	PARRS	TYFPDFTGGDNY	( 5 )	YLLHRFVS	( 0 ) LNQSAATKQIYA HYTCATDTQI KFVLSA QD ILLQLHL (6)
41	RKVPME	NYFPDYTGSDI	( 5 )	YILWRFVQ	( 0 ) LNRRAN LSIYP HVTOATDTSN IRLVFAA KET ILENTL (7)
42-43	K--P--	-YFPDY-G---D	( 6 )	-FE--FL-	( 0 ) -N-TN KPIYV -RTCATD---MKFVLSAVTD-I-QQNL (6)
44-45	--VPLN-CEWF-DYQ-VS-G	( 13 )	KFEE-Y-Q	( 0 )	-TAPDRVDRVFKI YRTTALDQKLVKKTFLVDETLRRRNL (6)
consensus					T A

Figure 3 (Continued)

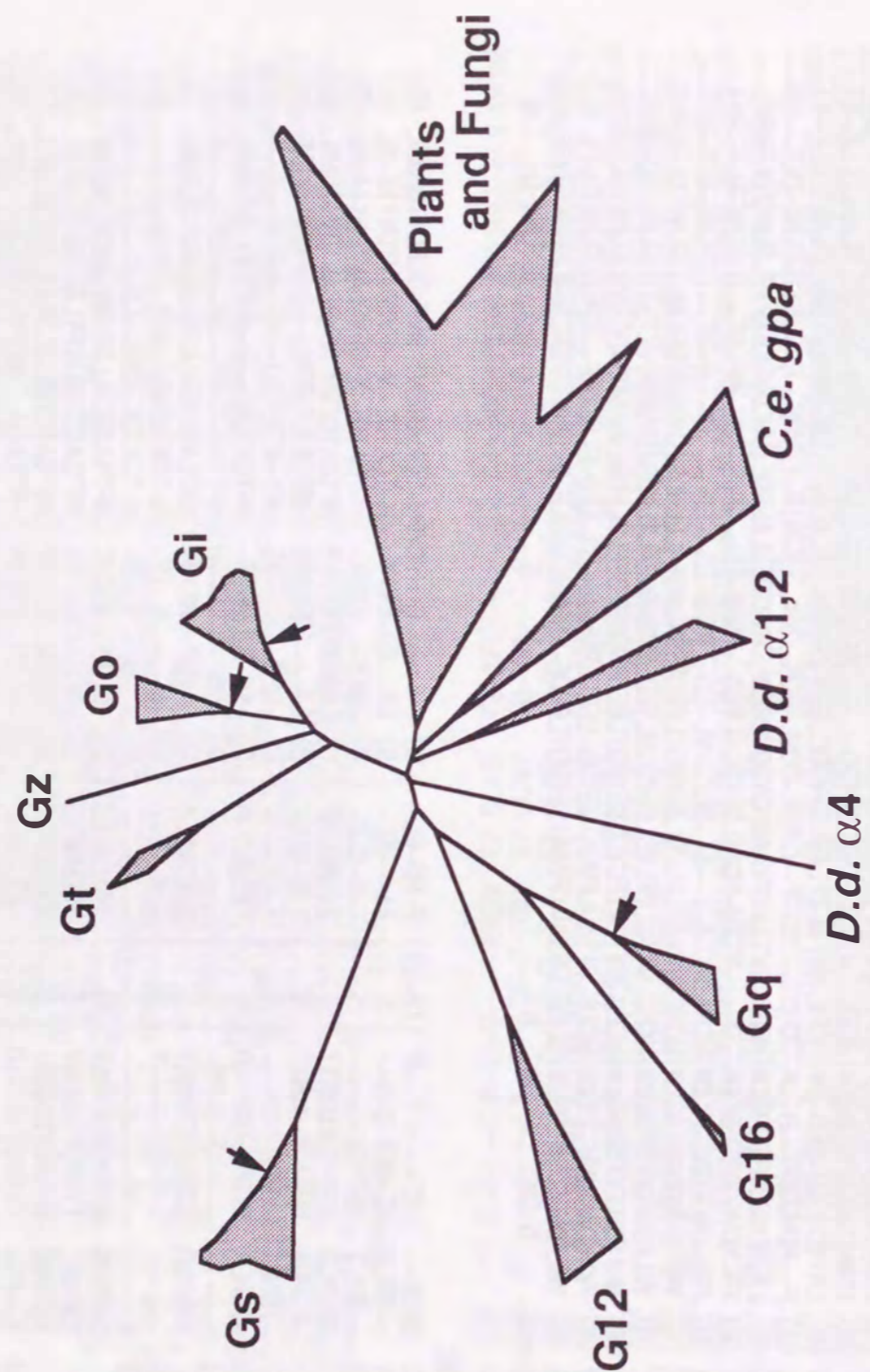


Figure 4(a)

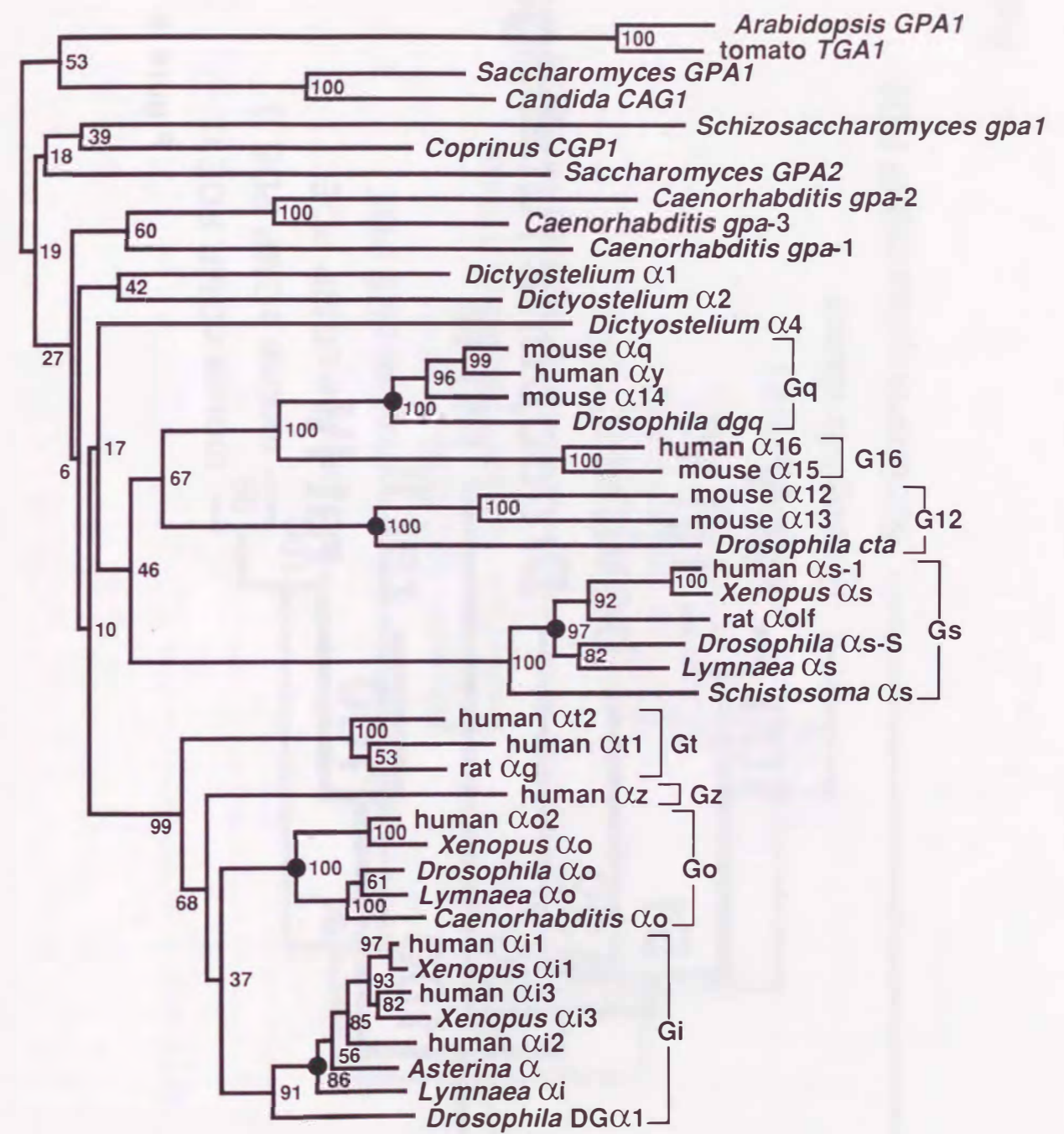


Figure 4(b)

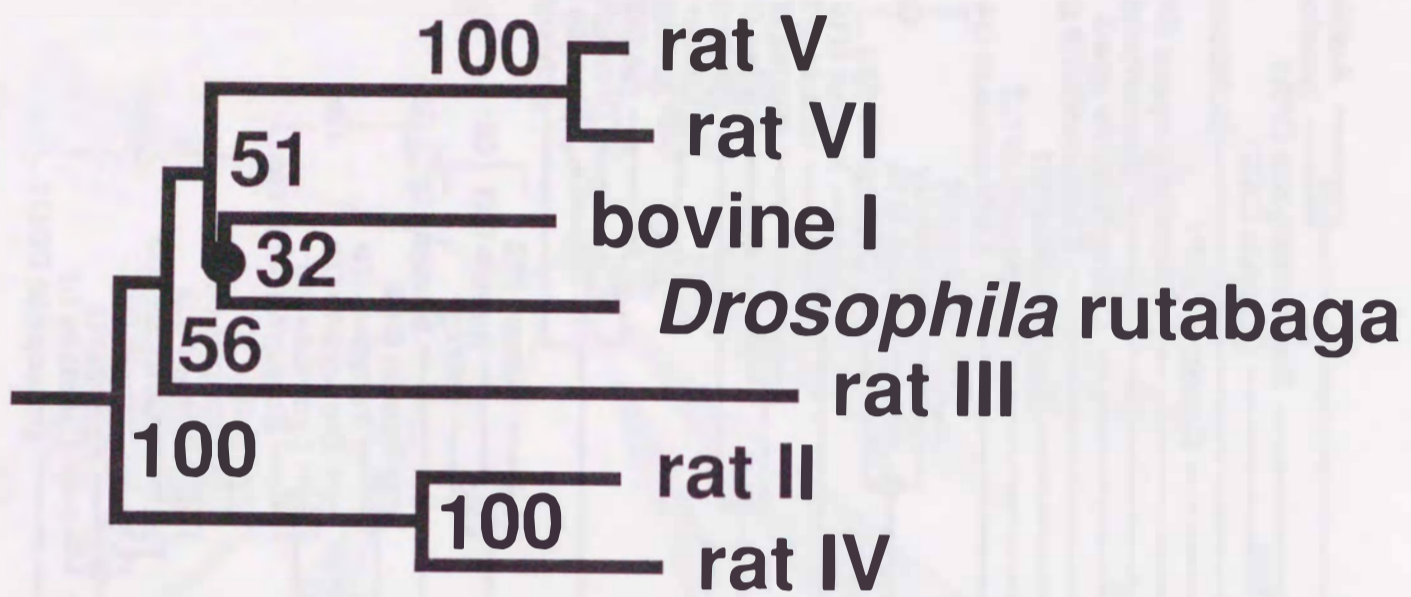


Figure 5

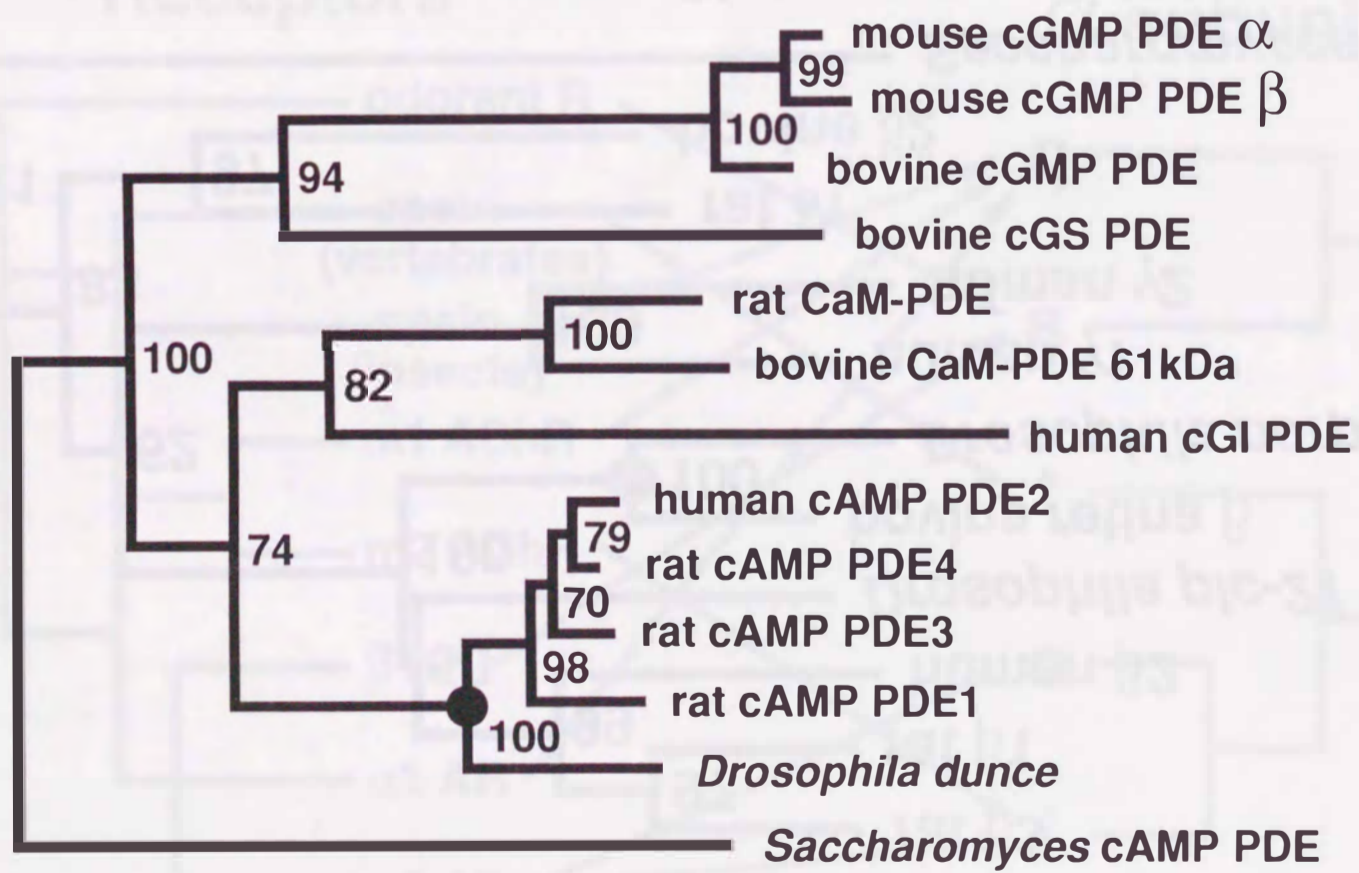


Figure 6

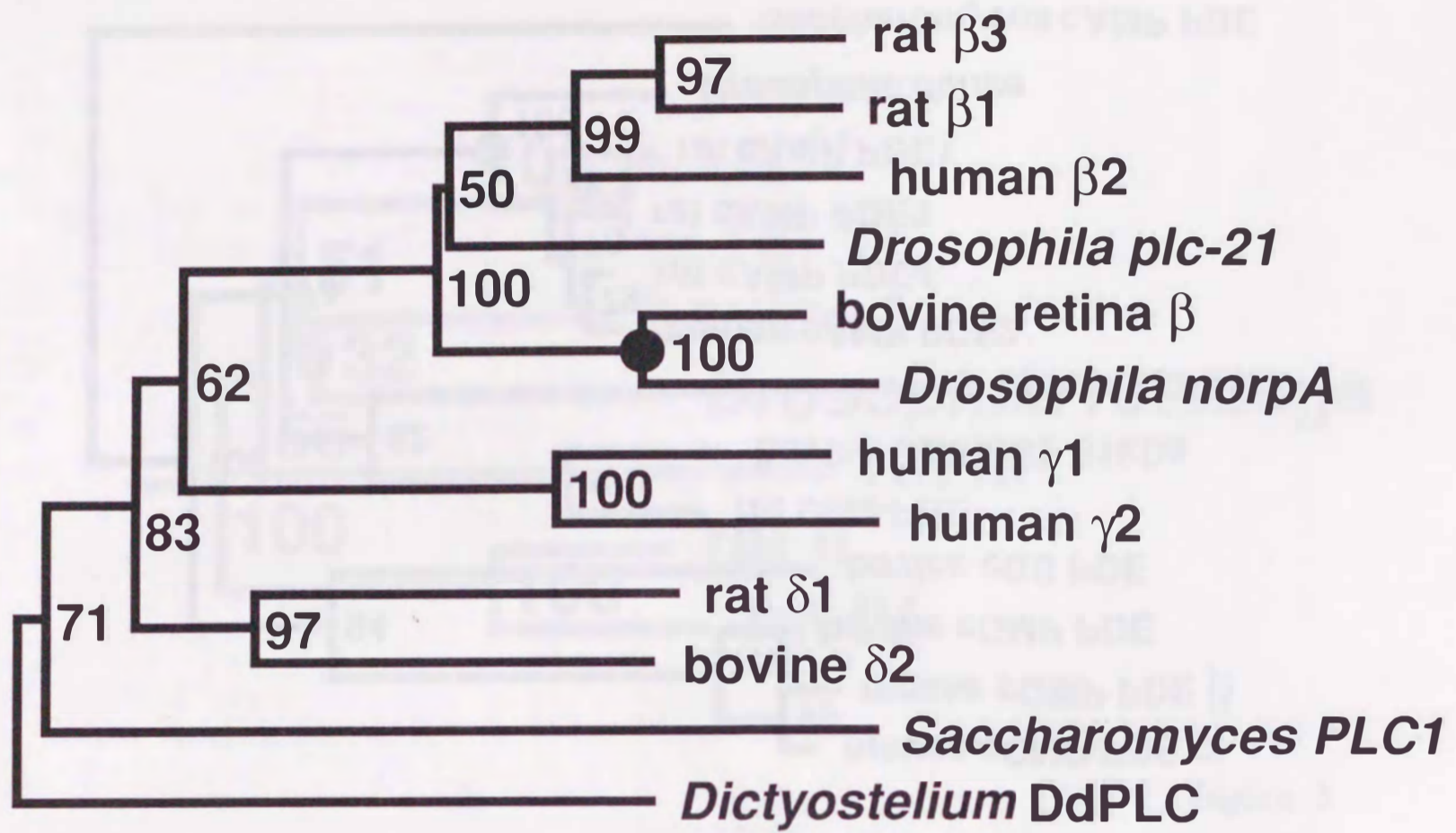


Figure 7

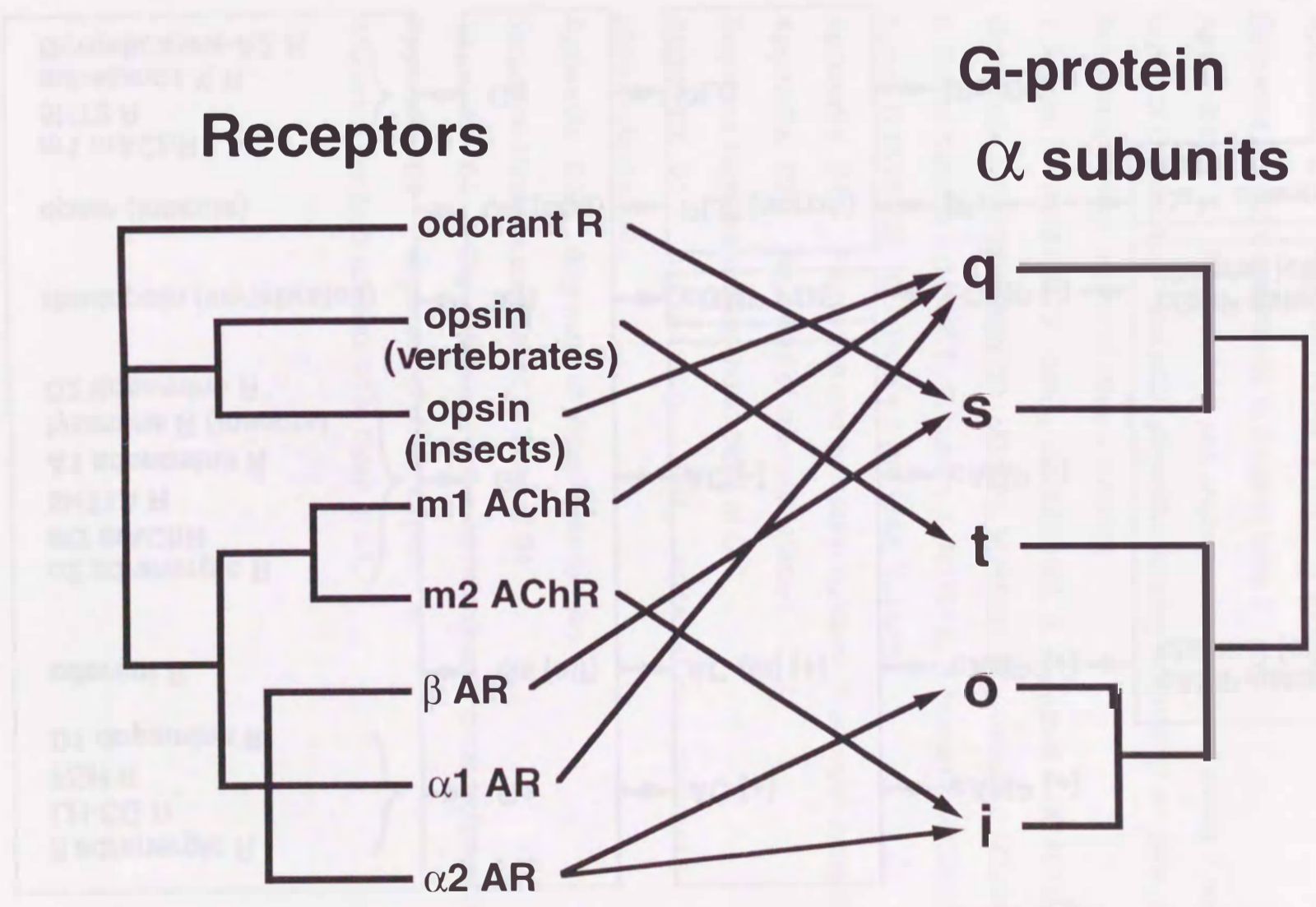


Figure 8

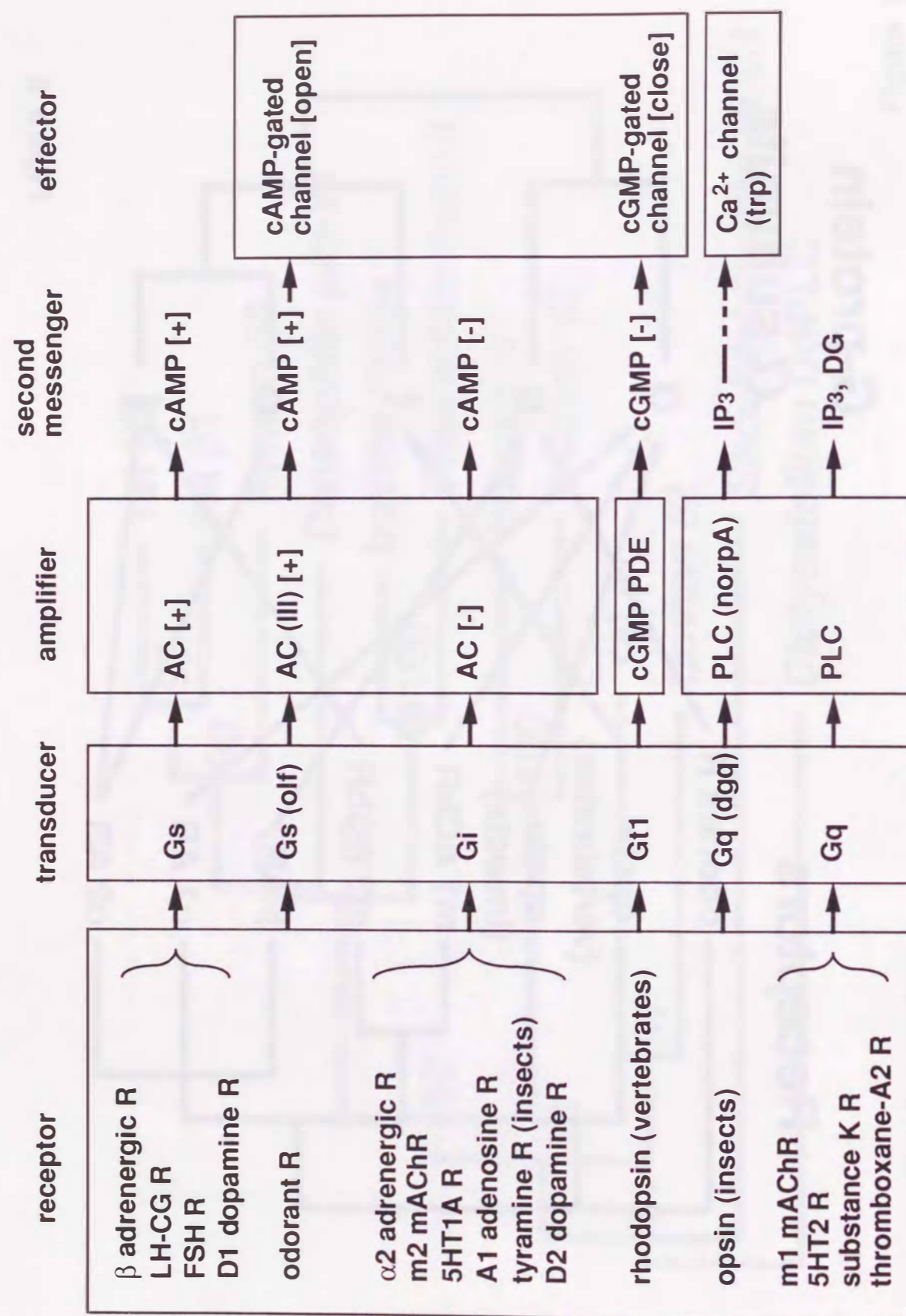


Figure 9

## Appendix

**Appendix A** Alignment of the rhodopsin family receptors.

Sequence numbers correspond to those of table 1.

**Appendix B** Alignment of the G protein  $\alpha$  subunits.

Sequence numbers correspond to those of table 3.

**Appendix B'** Alignment (a) and phylogenetic tree (b) of exons 7 and 8 region of G protein  $\alpha$ 1 and  $\alpha$ 2, alternative splicing products of  $\alpha$ 0 gene, and the corresponding regions of other  $\alpha$  subunits .

1. human  $\alpha$ 1 (M60165), 2. human  $\alpha$ 2 (M60165), 3. *Xenopus*  $\alpha$  (X14636), 4. *Drosophila*  $\alpha$  (M86660, M29731, M30151, M29602), 5. *Lymnaea*  $\alpha$  (Z15094), 6. *Caenorhabditis*  $\alpha$  (M38251), 7. human  $\alpha$ i1 (M17219), 8. human  $\alpha$ z (J03260), 9. human  $\alpha$ t1 (X15088), 10. human  $\alpha$ s-1 (M21142, X04408).

**Appendix C1** Expression and accession numbers of adenylyl cyclases.

**Appendix C2** Alignment of adenylyl cyclases.

Sequence numbers correspond to those of C1.

**Appendix D1** Expression and accession numbers of phosphodiesterases.

**Appendix D2** Alignment of phosphodiesterases.

Sequence numbers correspond to those of D1.

**Appendix E1** Expression and accession numbers of phospholipase Cs.

**Appendix E2** Alignment of phospholipase Cs.

Sequence numbers correspond to those of E1.

rhodopsin family receptor (1)

1. (37) SMLAAYMFLLLVLFVFPINFLTYVTVHKKL	(1) TPLNYILLNLAVADLFLMVGFG	TSTLYTSLH	(6) PTGCNLEGGFATLGGELALWLSVLVAERYVWCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
2. (37) SALAAYMFLLLVLFVFPINFLTYVTVHKKL	(1) TPLNYILLNLAVADLFLMVGFG	TTTMYTSMN	(6) VTGCGYEGGFATLGGELALWLSVLVAERYVWCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
3. (37) SALAAYMFLLLVLFVFPINFLTYVTVHKKL	(1) TPLNYILLNLAVADLFLMVGFG	TVMYTSMN	(6) PTMCSYEGGFATLGGELALWLSVLVAERYVWCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
4. (37) RLVCCYIFFLIIGLSTPLINLLTLVTFKHKKL	(1) QPLNYILLNLAVADLFLMVGFG	TVTFYASWY	(6) PVGCAYEGGFATLGGELALWLSVLVAERYVWCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
5. (37) KVLDFYMFLLIAGMPLNGLTLVTFVHKKL	(1) QPLNYILLNLAVADLFLMVGFG	TVTFYASWY	(6) PIGCAIEGGFATLGGELALWLSVLVAERYVWCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
6. (44) RAMAAMFLLIALGVPIINLTIFFCTARFKL	(1) SHLNYILLNLAVADLFLMVGFG	TTACYSFSO	(6) PTACKIEGGFATLGGELALWLSVLVAERYVWCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
7. (34) YLOAAMFVFLIGFPLNAMVLAATRYKLL	(1) QPLNYILLNLAVADLFLMVGFG	FVVFVASCN	(6) RHVCALEGLGLTVAAGLTVGSLAFLAFERYIVICK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
8. (32) YLOAFNGVIFVAVGTPNAVLAATRYKLL	(1) QPLNYILLNLAVADLFLMVGFG	FVVFVASCN	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
9. (53) HLTSMVMI FVYASVFNGLVLAATMKFKKL	(1) HPLNII LNLAVADLAEVVIAS	TISIVNOVS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
10. (53) HLTSMVMI FVYASVFNGLVLAATMKFKKL	(1) HPLNII LNLAVADLAEVVIAS	TISIVNOVS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
11. (50) NLTSMHMI FVVAASVFNGLVLAATMKFKKL	(1) HPLNII LNLAVADLAEVVIAS	TISIVNOVS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
12. (50) NLTSMHMI FVVAASVFNGLVLAATMKFKKL	(1) HPLNII LNLAVADLAEVVIAS	TISIVNOVS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
13. (48) NVSSLVMI FVYASVFNGLVLAATMKFKKL	(1) HPLNII LNLAVADLAEVVIAS	TISIVNOVS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
14. (48) NVSSLVMI FVYASVFNGLVLAATMKFKKL	(1) HPLNII LNLAVADLAEVVIAS	TISIVNOVS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
15. (52) NVSFFMI FVYASVFNGLVLAATMKFKKL	(1) HPLNII LNLAVADLAEVVIAS	TISIVNOVS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
16. (35) YSVGIFIGVGIIGLGNVVIYLFSTKTKSL	(1) TPANMFI LNLAFSDFSLVNGFPLMTISCFM	NAACKYGLG	(6) NAACKYGLGIGIFGLMSIMTMTMSIDRYNIGR	(11) KAFIMI FVWIMSTIWAIGPFI	GWGAY
17. (34) YSLGIFIAICGIGCVGNVVIYLFSTKTKSL	(1) TPANMFI LNLAFSDFSLVNGFPLMTISCFM	NAACKYGLG	(6) NAACKYGLGIGIFGLMSIMTMTMSIDRYNIGR	(11) KAFIMI FVWIMSTIWAIGPFI	GWGAY
18. (50) KILTAYMI IGMISWCGNGVVIYFATKTKSL	(1) TPANLLVNLAIADLGETVLS	TISVNOVF	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
19. (48) KFLAAYMVLIAIISWCGNGVVIYFSTTKSL	(1) TPANLLVNLAIADLGETVLS	TISVNOVF	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
20. (57) KILGLFTLAIMIISWCGNGVVIYFSTTKSL	(1) TPANLLVNLAIADLGETVLS	TISVNOVF	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
21. (58) YLLGLTYIFFTLMSLGNGLVIVVFAAKSL	(1) TPSNFI LNLAFSDFSLVNGFPLMTISCFM	PIFIYNSFH	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
22. (54) YLLGLTYIFFTLMSLGNGLVIVVFAAKSL	(1) TPSNFI LNLAFSDFSLVNGFPLMTISCFM	PIFIYNSFH	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
23. (22) QLFALFLIMYTLTFLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
24. (22) QLFALFLIMYTLTFLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
25. (24) HLFYALFLAMYLTIIGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
26. (24) HLFYALFLAMYLTIIGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
27. (24) HLFYALFLAMYLTIIGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
28. (24) QVLLFLFLIMYTLTFLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
29. (25) FLIFALFLSMYTLVTLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
30. (24) PLIFGLFLSMYTLVTLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
31. (25) YLLGLFLSMYTLVTLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
32. (27) IGLFLLFLSMYTLVTLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
33. (24) DLFYALFLAMYLTIIGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
34. (35) ITLFGVFLALYITLGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
35. (24) DLFYALFLAMYLTIIGNLLIVLVOLDLH	(1) TPMLYFLSNLFSDFLCSFVSTM	PKLLONMOS	(6) HPMCVLEGTVSLCGITGLWLSLAIISWERWVCK	(10) HAIMGVAFWMAIAACAAPPLA	GWSRY
36. (359) DFLRVL IWLINLAI MGNMVLVFLVLSRYK	(1) TVPFLMNLAFADLCEIIGL	LIASVDST	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
37. (362) NLRVL IWFISILAITGNI IVLVITTSOYK	(1) TVPFLMNLAFADLCEIIGL	LIASVDST	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
38. (414) KFLRI VVVFSLALLGNVFLVLLTSHYK	(1) TVPFLMNLAFADLCEIIGL	LIASVDST	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
39. (32) WLWAAAYTVI VTSVGNVVMIM ILAHKFM	(1) TVTNYFLVNLAFADLCEIIGL	VNFVYAVH	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
40. (33) ALWAPYALVLAIVAGNIAI IWI ILAHKFM	(1) TVTNYFLVNLAFADLCEIIGL	VNFVYAVH	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
41. (85) ALWSLAYGVAVAVLGNL IWI ILAHKFM	(1) TVTNYFLVNLAFADLCEIIGL	VNFVYAVH	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
42. (60) ALWSLAYGVAVAVLGNL IWI ILAHKFM	(1) TVTNYFLVNLAFADLCEIIGL	VNFVYAVH	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
43. (101) VLMSIFLGMV I VAGGNL I VMI VMTTKFM	(1) TVTNYFLVNLAFADLCEIIGL	VNFVYAVH	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
44. (83) TIWAI I FGLMFAVA I AGNI I VLV I VGTGRSM	(1) TVTNYFLVNLAFADLCEIIGL	VNFVYAVH	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
45. (71) ALLI VAYSFTI VFSLFGNLSVCH I VIKNRFM	(1) SATSLFI VNLAVADI I TLLNT	PFTLVRFN	(6) KMGCHVSRFAOYCSLHVSAL TAI AVORHVI	(9) KGV IYAVI VMAVTFSLPHAI	COKLF
46. (90) IYVYMLYPI I FAF I IAGN I VY I VYTPFM	(1) TVTNYFLVNLAFADLCEIIGL	VNFVYAVH	(13) GAGDAAAGFTVFASELSVYTLTIFLERWHTI	(11) HAASVVMWMI FAFAAALPFI	GISSY
47. (40) FTLALAYGAV I LGVSGNALL I I I LKOKEM	(1) NVTNII I VNLFSDDLVAI I MCL	PLTFVYTLM	(7) LALCHVYVSOAVSVLVSATLVAI I SDRYAI	(9) YATFI I AGVFI I ALATLPI	VSGLD
48. (56) ALO LLYSII I FLLSVLGNL I V I I I RKNFM	(1) TVTNI I FLLSVLSDLL I CLFCM	PFLNIPNL	(6) SAVCKTITTYHAGTSVSVSTFNVA I SLERYA	(14) KVIAATWLSFTI I MTPY I YSN	LVPTF
49. (54) A I R I T Y A V I F L M S V G A V L I I V L G L S P R L	(1) TVTNA I F L L S V S D L L A V A C M	P F T L P N L M	(6) TV I C K A I S Y L M G V S V S T L N L V A I A L E R Y S A I C R	(11) H A A R V I A T W M L S G L M P Y P V	Y T A V O
50. (54) A I R I T Y A V I F L M S V G A V L I I V L G L S P R L	(1) TVTNA I F L L S V S D L L A V A C M	P F T L P N L M	(6) TV I C K A I S Y L M G V S V S T L N L V A I A L E R Y S A I C R	(11) H A A R V I A T W M L S G L M P Y P V	Y T A V O
51. (80) Y I N T V I S C T F I V G M V G N A T L R I I Y O N Q M	(1) N G P N I I A S L A L G D L I Y V D I	P I N V F K L L A	(6) A E M C K L V P F I O K A S V G I T V L S L C A L S I D R Y R A V A S	(11) W T A V E I V L I V W V S V L A V P E A I	G F D I I
52. (101) Y I N T V S S L V F V L G I I G N S T L L R I I Y K N Q M	(1) N G P N I I A S L A L G D L I Y V D I	P I N V F K L L A	(6) A E M C K L V P F I O K A S V G I T V L S L C A L S I D R Y R A V A S	(11) W T A V E I V L I V W V S V L A V P E A I	G F D I I
53. (40) Y V I P A V G V I I G L I G N I T L I C F C T V K S M	(1) M P N L F I S S L A D L L I T C A	P V D A S R Y F L A	(6) R I G C K L I P F I O L T S V G S V F T A L S A D R Y A I V N	(11) R A G V M I G L A W I S F I L W A P A I L	FWOYF
54. (43) Y V I P A V G V I I G L I G N I T L I C F C T V K S M	(1) M P N L F I S S L A D L L I T C A	P V D A S R Y F L A	(6) R I G C K L I P F I O L T S V G S V F T A L S A D R Y A I V N	(11) R A G V M I G L A W I S F I L W A P A I L	FWOYF
55. (64) V L T A I Y A L F V G T V G S V T A F T A R K K S L	(1) S V P N I F I S N L A G D L L L L T C V	P V D A S R Y F L A	(6) R I G C K L I P F I O L T S V G S V F T A L S A D R Y A I V N	(11) R A G V M I G L A W I S F I L W A P A I L	FWOYF
56. (37) R A E L A L L S I V E V A V L S N G L V L A A L A R R G R	(1) S T V H Y L G S L A D L L I L L A M	P L P Y N F I W	(6) D A G O R G Y Y F L D Q A C T Y A T A L N V A S L S V E R Y L A I C H	(11) R T K K F I S A I I W A S A L L A I P M L F	W S E V A
57. (51) K L E I A V L A V I F V A V L G N S V L L A L H R T P R K	(1) S R L F F M K H S I A D V I A V A F O V	L P O L W A W K T	(6) D A L C R A V K Y L Q M G M Y A S S Y M I A M T L D R H A I C R	(11) H M N F P V L V A W A F S L L S L P O L F	I F A O R
58. (39) R E V E A V L C L I L L A L S G N A C V L L A R T T R O K	(1) S R L F F M K H S I A D V I A V A F O V	L P O L W A W K T	(6) D A L C R A V K Y L Q M G M Y A S S Y M I A M T L D R H A I C R	(11) H M N F P V L V A W A F S L L S L P O L F	I F A O R
59. (35) K I R V T Y F F L L S T A F N A S F L L K L O K W T O	(1) S P M K V L L K H L T A N L E T L I V M	P L D G M M N I T	(6) E F L C K V L S Y L K F S M Y A P A F M M V I S L D R S L A I T O	(9) L E O S M G L W I L S I V F A G P O Y F	I F S V I
60. (25) W I T S L L V I I C G L G I V G N I M V L V M R T K H M	(1) T P T N C Y L V S L A V A D L M L V A A G	L P N I S T S I Y	(6) E F L C K V L S Y L K F S M Y A P A F M M V I S L D R S L A I T O	(9) L E O S M G L W I L S I V F A G P O Y F	I F S V I
61. (116) L A I A V L S L T G T F T V L E N L L L C V I L H S P S L	(2) R P S Y H F I G S L A V A D L L G S V I F Y	Y S F I D F H F	(5) R N W F L K L G G V T A S F T A S V G S L F L T A I D R Y I S I H R	(11) K A V V A F C L M W T I A I V I A V L P L L	G W N C E
62. (45) K L T S W F L I C C F I L E N I F V L L T I W K T K F	(1) R P M Y Y F I G N L A D L L A G A V Y T	A N L L S G A T	(5) R N W F L K L G G V T A S F T A S V G S L F L T A I D R Y I S I H R	(11) K A V V A F C L M W T I A I V I A V L P L L	G W N C E

Appendix A

rhodopsin family receptor (2)

63. (44) NFMDDI VLCSGSLTCCENAVVLI I FHSPLS	(1) APMFLLI GSLADLGLGLI	INFFYAYLL	(2) EATKLVTI IGLIVASFSASVCSLLAI TVDRYLSLY	(11) LYLQMLVMLWGTSTCLGLLGY	LLELPE
64. (9) AAYIGIEVLIALVSYPGNVLVWAKVNOAL	(1) DATFCI VSLAVADVAVGLAI	PLAIIINIG	(4) FHTCLMIAQCPVLI LTOSSILALLAI AVDRYLRKI	(11) RAAYA I AGCW ILSLVGLTPMF	GWNKC
65. (3) SYYITVELAI IAVLAI I LGRVYVAVVNLNSML	(1) NYTNYFVSLAAADI AVGLVAI	PFAIITISTG	(4) CHGCLF IACFVVLTOSSIFSLAI IADRYIAIRI	(11) RAKGI IAI CNVLSFAI IGLTPML	GWNKC
66. (7) ALYVLEMAI IGLAVGVMVLCVAAVGTANTL	(1) TPTNYFLVSLAAADVAVGLVAI	PFAIITISTG	(4) FYGCLFACFVVLTOSSIFSLAVADRYLAICY	(11) RARGV IAVLHVLAFGLI IGLTPFL	GWNKC
67. (14) IYVYLEMAI IGLAVGVMVLCVAAVGTANTL	(1) TPTNYFLVSLAAADVAVGLVAI	PFAIITISTG	(4) FYGCLFACFVVLTOSSIFSLAVADRYLAICY	(11) RARGV IAVLHVLAFGLI IGLTPFL	GWNKC
68. (25) AFIGITGLLSLATVTVGNLVLISFKVNTL	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
69. (68) VFI AFLTGI LALVTI I GNI LV I VSKVKNOL	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
70. (30) ITIAAATVAVSLTI I VGNLVM I SFKVNSOL	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
71. (23) VFI VLVAGSLSVTI I GNI LVMS I KVNRFH	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
72. (26) VFI VLVAGSLSVTI I GNI LVMS I KVNRFH	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
73. (42) VFI ATVTGSLSLTVVGNII I VMLSI KVNROL	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
74. (42) VFI ATVTGSLSLTVVGNII I VMLSI KVNROL	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
75. (33) IFI ATVTGSLSLTVVGNII I VMLSI KVNROL	(1) TVNNYFLLSLACADI IIGTFSM	NLYTYYILM	(6) TLACDLWALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
76. (104) VVMGVAII I S V T V A G N V M V M I S F K I D K O L	(1) TVGNYI VLSVADL IGVGVMM	PMNIIYLLM	(6) PIVQDTWLALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
77. (28) TVGNYI VLSVADL IGVGVMM	(1) TVGNYI VLSVADL IGVGVMM	PMNIIYLLM	(6) PIVQDTWLALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
78. (18) ITI TVLAVL I I TVAGNVVCLAVLGNRL	(1) NLNCF I VLSA I D L L L G L L V L	P F S A I Y Q L S	(6) K V F C N I T S L D V M L C T A S I L N C A I S D R Y A I T S	(11) R A R G L C T W A A I S A L V S F L P I L M W N R A	
79. (58) AGMLLMAI I V L I V A G N V I V A I A K T P R L	(1) TLNFI I VLSVADL IGVGVMM	PMNIIYLLM	(6) PIVQDTWLALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
80. (41) AGMLLMAI I V L I V A G N V I V A I A K T P R L	(1) TLNFI I VLSVADL IGVGVMM	PMNIIYLLM	(6) PIVQDTWLALDYVASNASVNNLL IISFDRYFSVTR	(11) RAALMI GLAWLVSFLWAPAI I	FWOYF
81. (33) VGMISVMSL I V L I V F G N V L V I T A I A K F E R L	(1) TVTNYF I T S L A C A D L V M G L A V V	P F G A A H I Y L	(6) N F C W C F T S I D V L C V T A S I E T L C V I A D R Y A I T S	(11) K A R V I I L M W I V S G L T S F P I Q M W N R A	
82. (37) ALGALLALAVLATVGNLVI I VAI IAWTPRL	(1) TMTNVFVSLAAADVAVGLVAI	PFAIITISTG	(6) ATGCELWTSVDVLCCTAS IETLCAI IADRYAVTN	(11) C A R T A V V L V W V S A A V S F A P I M S O W N F	
83. (54) VGVGFLAAFI I VAVAGNLSVLSVACNRL	(1) TVTNYF I V N L A V A D L L S A T V L	P F S A T M E V L	(6) R A F C D V W A A D V L C C T A S I L S L C A I S D R Y V G R H	(11) K A A A I L A L L V A L V S V G P L L G W K P E	
84. (45) ISVGLVGFALFAI I VGNLVI I VLSVACNRL	(1) TPTNYF I V N L A I A D L L S F T V L	P F S A T M E V L	(6) S F F C D I W A A D V L C C T A S I L S L C A I S D R Y I G V R Y	(11) K A I L A L L S V W L S T V I S I G P L L	G W K P E
85. (26) ILLGVI I L G L I F G V L G N I L V I L S V A C N R L	(1) SVTHYI I V N L A V A D L L T S T V L	P F S A I F E I L	(6) R V F C N V W A A D V L C C T A S I M G L C I I S I D R Y I G V S Y	(11) R G L M A L L C V W A L S V I S I G P L F	G W R O P
86. (33) ILLVGLAGLMLLTVFQVNLVI IAVTSRAL	(1) APONFLVLSASADI I LVATLVI	P F S L A N E Y M	(6) K T W C E I L L D L V L F C T S S I V H L C A I S D R Y W S I T O	(11) R I K A I I I T T G S I S A V I S F P P L I	S I E K K
87. (51) AGLAAVGLI I V F T V G N V L V I A V L T S R A L	(1) APONFLVLSASADI I LVATLVI	P F S L A N E Y M	(6) K T W C E I L L D L V L F C T S S I V H L C A I S D R Y W S I T O	(11) R I K A I I I T T G S I S A V I S F P P L I	S I E K K
88. (12) AAIAVMI I T F L I F T I G N A L V I L A V L T S R A L	(1) APONFLVLSASADI I LVATLVI	P F S L A N E Y M	(6) K T W C E I L L D L V L F C T S S I V H L C A I S D R Y W S I T O	(11) R I K A I I I T T G S I S A V I S F P P L I	S I E K K
89. (109) LLTALVLSVI I V L T I I G N I L V I L S V F T Y K P L	(1) IVONFF I V S L A V A D L T V A L L V I	P F N V A Y L S	(6) I H L C K L W L T D V L C C T S S I L H L C A I A L D R Y A I T D	(11) R V L L L I S G V W L L S L I S S P P L I	G W N D
90. (36) VITSLLLGTL					





G protein α subunit (1)

Table with 45 rows of protein sequence alignments for G protein α subunit (1). Each row contains sequence identifiers, amino acid residues, and alignment markers. A consensus sequence 'LLLG GK Q' is shown at the bottom.

Appendix B

G protein α subunit (2)

Table with 45 rows of protein sequence alignments for G protein α subunit (2). Each row contains sequence identifiers, amino acid residues, and alignment markers. A consensus sequence 'D GGO W F E E LFLNK D K' is shown at the bottom.

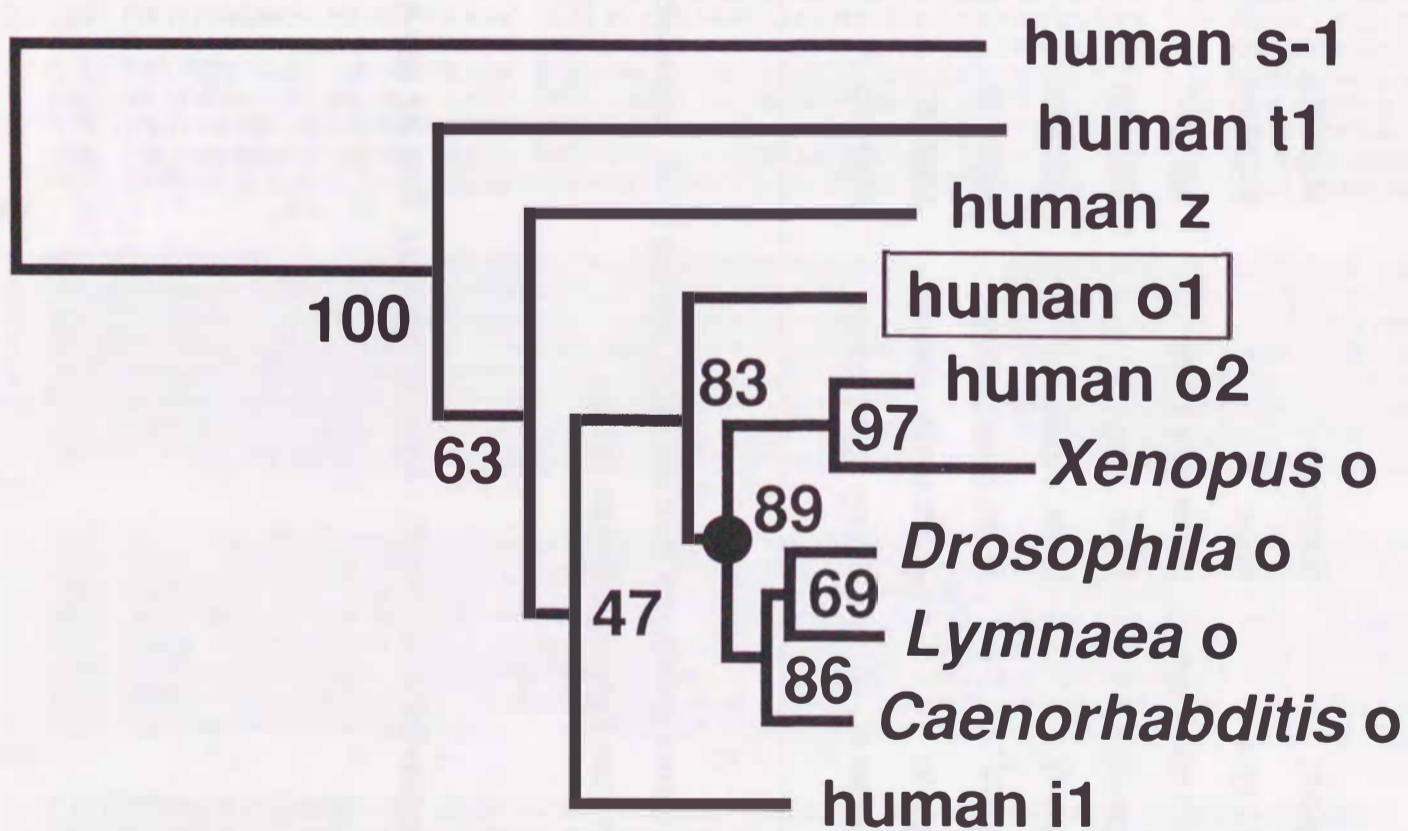
Appendix B (Continued)

G protein  $\alpha$  subunits ( $\alpha$ o subunit exon 7 and 8 region)

1.	(240)	TNRMHESLMLFDSICNNKFFIDTSIILFLNKKDLFGEKI	KKSPLTICFPEYTGPNITYED
2.	(240)	TNRMHESLKLFDSCNNKWFTDTSIILFLNKKDIFEEKI	KKSPLTICFPEYTGPSAFTE
3.	(240)	TNRMHESLKLFDSCNNKWFTDTSIILFLNKKDIFQEKI	KSSPLTICFPEYTGPNSTFE
4.	(240)	TNRMQESLKLFDSCNNKWFTDTSIILFLNKKDLFEEKI	RKSPLTICFPEYTGGOEYGE
5.	(240)	TNRMQESLKLFDSCNNKWFTETSIIILFLNKKDLFEEKI	KKSPLTICFPEYTGKOMYQE
6.	(240)	TNRMHESLKLFDSCNNKWFTDTSIILFLNKKDLFEEKI	KKSPLTICFPEYSGRODYHE
7.	(234)	MNRMHESMKLFDSCNNKWFTDTSIILFLNKKDLFEEKI	KKSPLTICYPEYAGSNTYEE
8.	(240)	TSRMAESLRLFDSCNNWFINTSLIILFLNKKDLLAEKI	RRIPLTICFPEYKGONTYEE
9.	(235)	VNRMHESLHLFNSICNHRYFATTSIVLFLNKKDVFEEKV	KKAHLSICFPDYDGPNTYED
10.	(262)	TNRLQALNLFKSIWNNRWLRTISVILFLNKODLLAEKVLAKGSKIEDYFPEFARYTTPED	
consensus		R E L F S I N S L F L N K D E K	P

1.	AAA	(0)	YIQAQFESKNRSPN	KEIYC	HMTCATDTNNIQVVFDAVTDII	IANNLRGCGLY	(0)
2.	AVA	(0)	YIQAQYESKNKSAH	KEIYS	HVTCATDTNNIQVVFDAVTDVI	IAKNLRGCGLY	(0)
3.	AVA	(0)	HTQHQYESRNKSEN	KEIYT	HITCATDTQNIQVVFDAVTDVI	IAYNLRGCGLY	(0)
4.	AAA	(0)	YIQAQFEAKNKSTS	KEIYC	HMTCATDTNNIQVVFDAVTDVI	IANNLRGCGLY	(0)
5.	ASA	(0)	YIQAQFEAKNKSSA	KEIYC	HQTCATDTNNIQVVFDAVTDVI	IANNLRGCGLY	(0)
6.	ASA	(0)	YIQAQFEAKNKSAN	KEIYC	HMTCATDTTNIQVVFDAVTDVI	IANNLRGCGLY	(0)
7.	AAA	(0)	YIQCFEDLNKRKDTKEIYT		HFTCATDTKNVQVVFDAVTDVI	IKNNLKDCGLF	(0)
8.	AAV	(0)	YIQRFEDLNRNKETKEIYS		HFTCATDTSNIQVVFDAVTDVI	IQNNLKYIGLC	(0)
9.	AGN	(0)	YIKVQFLELNMRDVKEIYS		HMTCATDTQNVKFCFVAVTDII	IKENLKDCGLF	(0)
10.	ATP	(13)	FIRDEFRLISTASGDGRHYCYPHFTCAVDTENIRRVFNDCRDI		IQRMHLROYELL		(0)
consensus		A		Y	H T C A D T N F D I L L		

Appendix B'(a)



Appendix B'(b)

Appendix C1

Expressions and Accession Numbers of Adenylyl Cyclases

No.	Gene	Expression	Accession No.
1.	rat AC type-III	olfactory	M55075
2.	bovine AC type-I	brain	M25579
3.	<i>Drosophila</i> rutabaga	mushroom body	M81887
4.	rat AC type-II	brain, lung	M80550
5.	rat AC type-IV	brain, others	M80633
6.	rat AC type-V	heart, brain, others	M96159
7.	rat AC type-VI	heart, brain, others	M96160
8.	<i>Dictyostelium</i> AC-A	during aggregation	M87279

Note. - Sequence data are from GenBank release 77.0. Expression data were taken from Tang and Gilman (1992).

References

Tang, W.-J. and Gilman, A. G. (1992). Adenylyl cyclases. *Cell* 70, 869-872.

adenylyl cyclase

1.	(306)	FNTMYMYRHENVSILFADIVGFTQLSSAC	SAQELVKLLNELFARFDKLAAYHQLRIKILGDCYYCICGLPDYR	(0)	EDHAVCSILMGLAMVEAISY
2.	(292)	FHKIYIQKHDNVSILFADIVGFTGLASQC	TAQELVKLLNELFGKFDLANTENHCRRIKILGDCYYCVSGLTOPK	(0)	TDHAHCCVEMGLDMIDTITS
3.	(262)	FHR IYIQKHENVSILFADIVGFTVLSQC	SAQELVRLNELFGRFDQLAHDNHLRIKILGDCYYCVSGLPEPR	(0)	KDHAKCAVEMGLDMIDA IAT
4.	(276)	FHNLVYKRHTNVSILYADIVGFTRLASDC	SPGELVHMLNELFGKFDQIAKENECMR I KILGDCYYCVSGLPISL	(0)	PNHAKNCVKMGLDMCEAIKK
5.	(260)	FHSLYVKRHOGVSVLYADIVGFTRLASEC	SPKELVLMNELFGKFDQIAKEHECMR I KILGDCYYCVSGLPLSL	(0)	PDHA I NCVRMGLDMCRA I RK
6.	(293)	FHK IYIQKHDNVSILFADIEGFTSLASQC	TAQELVMTLNELFARFDKLAENHCLRIKILGDCYYCVSGLPEAR	(0)	ADHAHCCVEMGMDMIEA I SS
7.	(378)	FHK IYIQKHDNVSILFADIEGFTSLASQC	TAQELVMTLNELFARFDKLAENHCLRIKILGDCYYCVSGLPEAR	(0)	ADHAHCCVEMGMDMIEA I SL
8.	(425)	IVIPEPEEYKSCSILCFD I VQFTNMSAKLDSPSRLVDLLTQVFRFEDTVVLRNGCOKIKTDGDAY I CACGLKSKK	(95)	HFEKLIDVAIEIMNLDLVLKE	
consensus		S L D I F T	L V L F F D		I K G D Y C G L
1.		VREKTKGVDMRVGVHTGTVLGGVLGQKRWQYDVWSTDVTVANKMEAGGIPGRVHIISQSTMDCLK	(0)	GEFDVEPGDGGSRCDYLDEKGIETYLIIASKPEVKK	
2.		VAEATEVDLNMVGLHTGRVLCGVLGLRKWQYDVWSDVTLANVMEAGLPGKVHIITKTLACLN	(0)	GDYEVEPGHGHENRSLFKTHNIEFTFIVPSHRRKI	
3.		VVEATDVILNMRVGIHTGRVLCGVLGLRKWQYDVWSDVTLANHMEAGGEPGRVHVTRATLDSLS	(0)	GEYEVEAGHGDERSYLRDHGVDTFIVPPHRRKP	
4.		VRDATGVDINMRVGVHSGNVLGCVIGLQKWOYDVWSDVTLANHMEAGGVPGRVHISVTLLEHLN	(0)	GAYKVEEGDGEIRDPLYLQHLVKTYFV I NPKGERRS	
5.		LRVATGVDINMRVGVHSGVLCGVLGKQWQYDVWSDVTLANHMEAGGVPGRVHITGATLALLA	(0)	GAYAVERADMEHRDYPYRELGEPTYLVIDPWAEED	
6.		VREVTGVNVMRVGIHSGRVHCGVLGLRKWQYDVWSDVTLANHMEAGGKAGRIHITKATLNYLN	(0)	GDYEVEPGCGGERNAYLKEHSIETFLILRCTQKRKE	
7.		VREVTGVNVMRVGIHSGRVHCGVLGLRKWQYDVWSDVTLANHMEAGGRAGRIHITRATLOYLN	(0)	GDYEVEPGRGGERNAYLKEOC I ETFLILGASQKRKE	
8.		TGNTG I QVQFRCG I AAGSVYGGV I GSQKYQFD I WGD I ARSHTLEQLGQPGKVVHGET I MTHKN	(21)	HDYEFHKAHGEC I TSYFVDWKDDYREKKKDLSCDF	
consensus		R G G V G V G	Q D W		E G G H
1.	(407)	KRDEELYSQSYDEIGVMFASLPNFADFYTEESINNGGIECLRFLNEIISDFDSDLNPNKFRVITKIKTIGSTYMAASGVTP	(22)	HLADLADFALAMKD	
2.	(370)	PRNMDLYYQSYQVGMFASIPNFNDFYIELDGNMNGVECLRLLEI IADFDLMDKDFYKDLKIKTIGSTYMAAVGLAP	(12)	HLSTLADFALIEIMFD	
3.	(491)	RNNMELYHOSYAKVGVIFASVPNFNEFYTEMDSQGLECLRLLEI IADFDLMDKDFYKDLKIKTIGSTYMAAVGLIP	(14)	HMTALIEYVKAMRH	
4.	(400)	LKNEELYHOSYDCVCMFASIPDFKEFYTESDVNKEGLECLRLLEI IADFDLMDKDFYKDLKIKTIGSTYMAATGLSA	(15)	HIGTMVEFAYALVG	
5.	(392)	RRNDELYHOSYECVCLFASIPDFKEFYSESNINHEGLECLRLLEI IADFDLMDKDFYKDLKIKTIGSTYMAATGLNA	(15)	HLGTMVEFAVALGS	
6.	(405)	RRNDELYYQSCCECVAMFASIANFSEFYVELEANNEGVECLRLLEI IADFDEI I SEDFRQLEKIKTIGSTYMAASGLND	(9)	H I KALADFAMKLM	
7.	(403)	RRNDELYYQSCCECVAMFASIANFSEFYVELEANNEGVECLRLLEI IADFDEI I SEERFRQLEKIKTIGSTYMAASGLNA	(9)	H I TALADYAMRLME	
8.	(437)	ETKGIYVYQPHQDVSIMFIQIAGFOEY	DEPKDLIKKLNDFISFFDGLLNQKYGGTVEKIKTIGNTYMAVSGLDG	(0)	SPSFLEKMSDFALD
consensus		Y Q	F F		LN I F D K I K T G T Y M A G
1.		TLTNINNSQFNFMRLRIGMNGGVLGAGVIGARKPHYD I WGNVNVASRMESTGVMGN I QVVEETQV I LREYGFRRVRRGPIFVKGKGELLTFFLKGR	(22)		
2.		VLDEINYSYNDVFLRVGINVGPVAVG I GARRPOYD I WGNVNVASRMESTGVQGR I QVTEEVHRLLRGSRFVCRGKVSVKGKGEMLTFFLEGR	(74)		
3.		SLQEINSHSYNNFMLRVGINIGPVVAVG I GARKPOYD I WGNVNVASRMESTGVPGYSQVTOEVVDSL VGSHFEFRGRT I KVKGKGMVTYFLCDS	(1094)		
4.		KLDAINKHSFNDFKLRVGINHGPV I AGV I GAQKPOYD I WGNVNVASRMESTGVLDK I QVTEETSL I LQTLGYTCTCRG I INVKGKGD LKTYFVNT	(12)		
5.		KLGVINKHSFNDFKLRVGLNHGPVAVG I GAQKPOYD I WGNVNVASRMESTGVLGK I QVTEETARALQSLGYTCYSRGVIKVKGKGQLCTYFLNTD	(10)		
6.		QMKY I NEHSFNDFOMK I GLNIGPVVAVG I GARKPOYD I WGNVNVASRMESTGVPDR I QVTTDMYQVLAANTYQLECRGVVVKGKGEMMTYFLNGG	(4)		
7.		QMKH I NEHSFNDFOMK I GLNMGPVAVG I GARKPOYD I WGNVNVASRMESTGVPDR I QVTTDL YQVLAAGYQLECRGVVVKGKGEMMTYFLNGG	(3)		
8.		VKAYTNSVAI SRVVRIG I SHGPLVAGCIG I SRAKFDVWGDANTASRMQSNADNE I MVTHSVYERLNKLFYFDDEKE I L VKGKGKMTVHLKKG	(50)		
consensus		N	G G A G I G		D W G T N S R M S V L V K G K G T

Appendix C2

Appendix D1  
Expressions and Accession Numbers of Phosphodiesterases

No.	Gene	Expression	Accession No.
1.	human cAMP PDE (PDE2)	brain, heart, kidney testis <sup>a</sup>	M37744
2.	rat cAMP PDE (PDE4)	brain, heart, liver, kidney, testis	J04563
3.	rat cAMP PDE (PDE3)	brain, heart, liver, kidney, testis	M25349
4.	rat cAMP PDE (PDE1)	testis, kidney	M25347
5.	<i>Drosophila</i> cAMP PDE ( <i>dunce</i> )	NI	X55167
6.	rat calmodulin-dependent PDE (CaM-PDE)	NI	M94537
7.	bovine calmodulin-dependent PDE 61-kDa (CaM-PDE) (partial)	NI	A40282b
8.	human cGMP-inhibited cAMP PDE (cG1 PDE)	heart	M91667
9.	mouse cGMP PDE $\alpha$	rod cell	X60664
10.	mouse cGMP PDE $\beta$	rod cell	X55968
11.	bovine cGMP PDE	cone cell	M37838
12.	bovine cGMP-stimulated cNMP PDE (cGS PDE)	ubiquitous	M73512
13.	<i>Saccharomyces</i> cAMP PDE (PDE2)	NI	M14563

Note: - Sequence data are from GenBank release 77.0 and NBRF release 36.0.  
NI = not identified.

a Rat expression data from Swinnen et. al. (1989).  
b Sequence data from NBRF release 36.0.

References

Swinnen, J. V., Joseph, D. R., and Conti, M. (1989). Molecular cloning of rat homologues of the *Drosophila melanogaster dunce* cAMP phosphodiesterase: evidence for a family of genes. Proc. Natl. Acad. Sci. USA 86, 5325-5329.

PDE

1. (228) DVAYHNSLHAADVLOSTHVLLATPALDAVFTDLEILAALFAAAIHVDVHPGVSNQFLINTNSELALMY NDESVLEN HHLAVG FKLLQEYNC  
2. (227) DVAYHNSLHAADVAQSTHVLLSTPALDAVFTDLEILAAIFAAAIHVDVHPGVSNQFLINTNSELALMY NDESVLEN HHLAVG FKLLQEEHC  
3. (233) DVAYHNNIHAADVQSTHVLLSTPALEAVFTDLEILAAIFASAIHVDVHPGVSNQFLINTNSELALMY NDSSVLEN HHLAVG FKLLQEEHC  
4. ( 97) NVAYHNSIHAADVQSAHVLLGTPALEAVFTDLEVLAAIFACAIHVDVHPGVSNQFLINTNSELALMY NDSSVLEN HHLAVG FKLLQGENC  
5. (242) DNPFFHNSLHAADVTQSTNVLLNTPALEGVFTPLEVGGALFAACIHVDVHPGLTNOFLVNSSSELALMY NDESVLEN HHLAVA FKLLQOQGC  
6. (217) KNPYHNQIHAADVTQTVHCFLLRTGMVHCLSEIEVLAIFAAAIDHYEHTGTTNSFHIQTKSECAILY NDRSVLEN HHISSV FRMMQDDDEM  
7. (213) KNPYHNL IHAADVTQTVHYIMLHTGIMHWLTELEILAMVFAAAIDHYEHTGTTNMFHIQTRSDVAI LY NDRSVLEN HHVSAA YRLMQEEEM  
8. (791) SGFTGHMGYVFSKTYNVTDDKYGCLSGNIPALELMALYVAAAMHDYDHPGRTNAFLVATSAPOAVLY NDRSVLEN HHA AAAWNLFMSRPEY  
9. (554) RITYHNWRHGFNVGQTMFSLLVGTGKLRKRYFTDLEALAMVTAFAFCHIDHRGTNNLYQMKSONPLAKL HGSSILER HHLEFG KTLRDESL  
10. (552) RITYHNWRHGFNVGQTMFLLMTGKLSYYTDLEAFAMVTAAGLCHIDHRGTNNLYQMKSONPLAKL HGSSILER HHLEFG KFLLAEEESL  
11. (552) AVTYHNWRHGFNVGQTMFLLMTGRLKYYTDLEAFAMLAFAFCHIDHRGTNNLYQMKSTSPARL HGSSILER HHLEYS KTLLODESL  
12. (631) DPPYHNWMMHAFVSHFCYLLYKNELELTYLEDMEIFALFISCMCHLDHRGTNNSFQVASKSVLAALYSSEGSVMER HHFAQA IA I LNTHGC  
13. (260) VNKFNFRHAIDVMOATWRLCTYLLKDN PVQTL L L C M A A I G H D V G H P G T N N Q L L C N C E S E V A Q N F K N V S I L E N F H R E L F Q Q L L S E H W  
consensus H HD H G N A S E H

1. DIFQNLTKRQSLRKMVIDMVLATDMSKHMTLLADL (25) VLRNMVHCADLSNPTKPLELYROWTDRIMAEFFQOQDRERE RGMEISPMC (255)  
2. DIFQNLTKRQSLRKMVIDMVLATDMSKHMSLLADL (25) VLRNMVHCADLSNPTKPLELYROWTDRIMAEFFQOQDKERE RGMEISPMC (132)  
3. DIFQNLTKRQSLRKMVIDVLATDMSKHMTLLADL (25) VLQNMVHCADLSNPTKPLQLYROWTDRIMAEFFQOQDRERE RGMEISPMC (148)  
4. DIFQNLSTKQKLSLRKMVIDMVLATDMSKHMSLLADL (25) VLQSLVHCADLSNPAKPLPLYROWTERIMAEFFQOQDRERE SGLDISPMC (59)  
5. DIFCNMQKQKQSLRKMVIDVLSTDMSKHMSLLADL (25) VLENLVHCADLSNPTKPLPLYKRWVALLMEEFFLOQDKERE SGMDISPMC (139)  
6. NIFINLTKDEFVELRALVIEMLATDMSCHFQOVKTM (14) ALSLLLHAADISHPKQWSVHSRWTKALMEEFFROQDKAE LGLPFSPLC (126)  
7. NVLINLSKDDWRDLRNLVIEMLSTDMSGHFQQIKNI (14) TMSLILHAADISHPAKSWKLHHRWTMALMEEFFLOQDKAE LGLPFSPLC (124)  
8. NFLINLDHVEFKHFRFLVIEAIIATDLKKHFDVAKF (20) VCOMCKLADINGPACKEHLQWTDGIVNEFYEQDDEAS LGLPISPFM (151)  
9. NIFQNLNRRQHEHAIHMDIAIIATDLALYFKKRTMF (29) VMAMMTACDLSAITKPWEVQSKVALLVAAEFWEQDLERTVLQOQPIPM (98)  
10. NIFQNLNRRQHEHVIHLMDIAIIATDLALYFKKRTMF (29) VMAMMTACDLSAITKPWEVQSKVALLVAAEFWEQDLERTVLQOQPIPM (97)  
11. NIFQNLNRRQYETVHILFEVAIIATDLALYFKKRTMF (29) VMAMMTACDLSAITKPWEVQSKVALLVNEFYEQDDEAS LGLPISPFM (96)  
12. NIFDHFSRKDYQRMLDLMDRIILATDLAHLRIFKDL (18) LLCLLMTSCDLSQDTKGWKTTRKIAELIYKEFFSQGDLEKA MGNRPMEM (93)  
13. PLKLSISKKKFD FISEAIIATDMALHSQYEDRL (10) LISLIIKAADISNVTRTLISARWAYLITLEFNDCALLETFFHKAHRPEQDC (85)  
consensus TD D EF E

Appendix D2

Appendix E1

Expressions and Accession Numbers of Phospholipase Cs

No.	Gene	Expression	Accession No.
(A) Phospholipase C $\gamma$ (Activated by Protein Tyrosine Kinases)			
1.	human PLC- $\gamma$ 1	NI	M34667
2.	human PLC- $\gamma$ 2	NI	X14034;M37238
(B) Phospholipase C $\delta$ (Activated by Unknown Factors)			
3.	rat PLC- $\delta$ 1	adrenal > brain	M20637
4.	bovine PLC- $\delta$ 2	brain	S14113a
(C) Phospholipase C $\beta$ (Activated by G-proteins)			
5.	rat PLC $\beta$ 3	NI	M99567
6.	rat PLC- $\beta$ 1	NI	M20636
7.	human PLC- $\beta$ 2	NI	M95678
8.	<i>Drosophila plc-21</i>	adult and larval brain	M60452
9.	bovine retina PLC- $\beta$	retina	L13935
10.	<i>Drosophila norpA</i>	eye	J03138
(D) <i>Dictyostelium</i> and <i>Saccharomyces</i> Phospholipase C			
11.	<i>Saccharomyces PLC1</i>	NI	D12738
12.	<i>Dictyostelium DdPLC</i>	all stages of development	M95783

Note. - Sequence data are from GenBank release 77.0 and NBRF release 36.0. > indicates that tissue expressions of the right are weaker than those of the left. NI = not identified.

a Sequence data from NBRF release 36.0.

PLC	
1. (322) NNPLSHYWISSSHNTYL TGDQFSSSESSLEAYARCLRMGCRCLIELDCWDG	4. (322) PVIVHGYTLTKIKFSDVLTIKKEHAFVASEYPPVILSIEDHCSIA QQRN
2. (314) NNPLSHYWISSSHNTYL TGDQLRSESSPEAYIRCLRMGCRCLIELDCWDG	4. (314) PVIVHGYTRTKIKFDDVVOA IKDHAFVTSSFPVILSIEEHCSVE QQRH
3. (298) DQPLSHYLVSSSHNTYLLEDQLTGPSSSTEAYIRALCKGCRCLIELDCWDG	4. (298) PIIYHGYTFTSKILFCDVLRRAIRDYAFKASPPVILSLENHCSLE QQRV
4. (292) TOPLNHYYINSSHNTYL VGDQLCGOSSVEGYIRALKRGCRCEVDIWDG	4. (292) PIVYHGHTLSRIPFKDVAIAIGYAFOTSDYPPVILSLENHCSWE QQEI
5. (259) TOPLSAYFINSSHNTYL TAGQLAGTSSVEMYROALLWGCRCELDVWKG	6. (259) PFI THGFTMTTEVPLRDVLEIAI AETAFTKSPYPVILSLENHCSAKQQA
6. (314) TOPLNHYYINSSHNTYL TAGQFSGLSAEMYROVLLSGCRCELDVWKG	6. (314) PII THGFTMTTEIFFKEAIEAIAESAFKTSPPVILSLENHCSVSPKQA
7. (318) SÖPLSHYFINSSHNTYL TAGQLAGNSSVEMYROVLLSGCRCELDVWKG	6. (318) PVI THGFTMTTEISFKEVIEAIAESAFKTSPPVILSLENHCSVSPKQA
8. (320) DÖPMSHYFINSSHNTYL TGHÖLTKGSSVEIYRÖCLLAGCRCELDVWKG	4. (320) PVI THGFTMTTEIFFKDVLEIAIAESAFKTSPPVILSLENHCSNPR QQA
9. (151) DHPLAHYFINSSHNTYL TGROFGGKSSVEMYROVLLSGCRCELDVWKG	6. (151) PII THGKAMCTDILFKDVOIAIKETAFTVSEYPPVILSLENHCSKY QQYK
10. (321) DÖPLAHYYINSSHNTYL SGRÖIGGKSSVEMYRÖTLLAGCRCELDVWKG	6. (321) PIVTHGHAYCTEILFKDIOAIA DCAFVSSEYPPVILSLENHCSNRA QQYK
11. (324) SKPLSYFFINSSHNTYLSGHÖLKGSTSEMYTNTLROGCKCELDVWKG	4. (324) PII FHGNTLSÖIKFSHVCETIKARGFETSPPVILSLENHCSVSP QQIM
12. (382) SKPLNHYYINSSHNTYL LGGÖIAETPSVEGYIOVLÖGCRCELDVWKG	3. (382) PVVCHG FLTSAIPLKTVIRVIKYYAFITSPYPLIISLEINCNKD NOKL
consensus P Y SSSHNTYL Q E Y L G C C E D W G P H G I F S P S E Q	
1. MAOYFKKVLGDTL (8) ADGLPSPNQLKRLIKH (544) LSRIPYKGORLSSNYDPLPMWICGSQVALNFOTDPKPMOMNOALF (5) CGYVLOPSTM (22)	
2. MAKAFKEVFGDLL (8) ADOLPSPSÖLREKIIKH (526) LTRVYYPKÖRVSSNYDPPFRWL CGSOMVALNFOTADKYMOMNHAF (5) TGYVLOPESM (21)	
3. MARHLRAILGPIIL (8) TSLPSPEQLKGIKLLKG (107) LSRIPYAGWRDSSNYSPVEMWNGGCOI VALNFOTPGPEMDVYLGCF (5) CGYVLPKPAFL (24)	
4. IVRHLTEILGDÖL (9) PTÖLPSPEDLRGIKLVKG (109) LSRVYPSGLRDTSSNYMPOEFWVWAGCOMVAMNÖTAGLEMDL CDGLF (5) CGYVLPKPDFL (27)	
5. MAEYCRSIFGEAL (12) GTPLPSPÖDLMGRILVKNI (177) LSRIPYKGRVDSNYMPÖLFWNVGCOLVALNFÖTLDPMÖLNAGVF (5) SGYLLKPEFM (24)	
6. MAEYCRIFGDML (12) GVPLPSPEDLRGIKLIKNI (133) MSRIPYKGRTRDSSNYMPOEFWVWAGCOMVALNFÖTMDLPMÖQNMVAV (5) SGYLLKHEFM (24)	
7. MAEYCRIFGDAL (12) GVPLPSPMDLRYKILVKNI (127) LSRIPYKGRVDSNYMPÖLFWNVGCOLVALNFÖTDLAMÖINMGMY (5) SGYRLKPEFM (24)	
8. IANYCREIFGDML (12) NMDLPPAMLRRIKIKNI (187) LSRVYYPAGTRFDSSNYMPOEFWVWAGCOMVALNFÖTDLAMÖNLGIF (5) SGYLLKPEFM (24)	
9. MSKYCEDLFGDLL (12) GRPLPSPNDLKRKILIKKI (156) MSRIPYKGRVDSNYMPOEFWVWAGCOMVSLNYÖTDLAMÖLNÖGKF (5) CGYLLKPDFM (24)	
10. LAKYCDFFGDLL (12) GLPLPPCKLKRKILIKKI (135) MSRIPYKGRTRDSSNYMPOEFWVWAGCOMVSLNFÖSSDLPMÖLNÖGKF (5) CGYLLKPDFM (24)	
11. MANHMKEIFGEML (7) TKELPTLDSLKYIKLLKG (126) LLRVYYPGRTRFDSSNYMPOEFWVWAGCOMVSLNFÖSSDLPMÖLNÖGKF (5) CGYVLPKPDFL (25)	
12. ASLIMREVLAEÖL (6) TDKLPSPRELKHKILLLKS (121) LMRVYYPVHLRYKSSNFNPIPFWKAGVOMVATNÖTNDIGÖÖLNAMF (12) SGYVLPKPKL (27)	
consensus L LP L I K R Y P R S S N P W G Ö N Ö G Y L	
1. IEVLGARHLP (7) CPFVEIEV AGAEDYSTKO KTEFV VDNGLNPVWPAK (5) ISNPEFAFLRFV (11) LAÖATFPVKGLKTGYRAVPL (98)	
2. VKVLGARHLP (7) CPFVEVEI CGAEGNNKF KTTVV NDNGLSPIWAPT (7) IYDPNLALFRFV (11) LAHATYPIKAVKSGFRSVPL (85)	
3. VRIISGÖQLP (10) DPKVIVEI GVGÖRDTGSR ÖTAVI TNGNFNPRWDE (4) VTPDÖLALVRFMV (11) IGÖSTIPWNSLKGÖYRHVHL (21)	
4. LÖVISGÖQLP (11) DPLVRIEIV FGVRPDTTRÖ ETSYV ENNGFNPYWÖT (4) ILVPELALLRFV (11) IGÖYTLPWSCMÖÖGYRH IHL (28)	
5. VKVISGÖFLS (4) GIYVEVDM FGLPVDTRRK YRTRTS ÖGNFNPVWDEE (6) VVLPTLÄSLRIÄÄ (7) VGHRIIPVSAIRSGYHYVCL (410)	
6. ITVISGÖFLS (4) RTYVEVEL FGLPGDPKRR YRTKLSPTSNINPVWKEE (6) ILMPELÄSLRVAV (7) LGHRIIPINÄLNSGYHHLCL (403)	
7. VKIISGÖFLS (4) GTYVEVDM FGLPVDTRRKAFKTKTS ÖGNÄVNPVWEE (6) VVLPÄLÄCLRÄÄ (7) IGHRIIPVÖAIRPGYHYICL (440)	
8. ITVLISGÖFLT (4) NTFVEVDM YGLPADTVRKKFRKTV RDNÖMNPYDEE (6) VVLPÄLÄSLRIÄÄ (7) IGHRIIPVIGLCPGYRHVNL (470)	
9. VÖVISGÖFLS (4) GTYVEVDM YGLPTDTRIRKEFRTRMV MNNGLNPVYNEE (6) VILPDLÄVLRÄÄV (7) IGÖRIIPLDGLÖAGYRHISL (374)	
10. VKVIÄGÖFLS (4) GTYVEVDM FGLPDSÖTVKKEFRTRLV ANNGLNPVYNE (6) VVLPDLÄVLRFGV (7) LGÖRIIPLDGLÖAGYRHVSL (309)	
11. VNVISÄRÖLP (11) DPYVTLISV GTHFDÖKVE KTKVI DNNGFNPHWÖEE (4) LYSNÖLSMLLIRV (10) IGHÖCIRVENIRPGYRIKLL (21)	
12. IRILSTÖLLP (24) EPTMPSIDKGRISÄTEÄ STKSS ÖGNÖFNPIWÄE (4) LKÖTDLTFIKFMV (6) IÄSVCLKLNLYLRMGYRH IPL (20)	
consensus L G T N P G L	

Appendix E2