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Dietary Effect of Yeast Extract or Glutathione on Lipid Metabolism and Immune Function of Sprague–Dawley Rats

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Dietary effect of yeast extract YH (YH) and glutathione on lipid metabolism and immune function of Sprague–Dawley rats was examined. Dietary effect of these diets was small on growth and tissue weights, as well as the effect on serum levels of cholesterol, triglycerides, phospholipids and triarbituric acid reactive substances. On the other hand, LTB₄-releasing activity of peritoneal exudate cells was significantly lower in the rats fed 10% YH diet than the rats fed control or 1% YH diet. In the rats fed 0.01% GSH diet, enough number of peritoneal exudate cells was not recovered. In addition, YH and GSH feeding slightly decreased the levels of serum IgA, IgG and IgM, as well as the productivities of these antibodies in spleen lymphocytes. These results suggest that YH feeding affects rat immune functions such as eicosanoid and immunoglobulin production.

INTRODUCTION

It has been reported that various food components affect diverse biological functions. Among them, the immunoregulatory effect of food components is related to the incidence or prevention of infectious diseases, allergies, and cancers. For instance, some unsaturated fatty acids regulate allergic reactions such as eicosanoid production through the regulation of lipid metabolism (Terano *et al.*, 1984; Thien *et al.*, 1993). In addition, antioxidants such as α -tocopherol (Gu *et al.*, 1994, 1995), tea polyphenols (Matsuo *et al.*, 1996, 1997), and flavonoids (Baumann *et al.*, 1980; Corvazier and Maclouf 1985; Laughton *et al.*, 1991) have been reported to suppress eicosanoid production, probably by inhibiting lipid oxidizing enzymes such as lipoxygenase and cyclooxygenase.

In addition, unsaturated fatty acids and antioxidants have been reported to regulate immunoglobulin (Ig) production of rat lymphocytes *in vitro* in a class-specific manner. For instance, unsaturated fatty acids enhance IgE production of rat and mouse lymphocytes, suppressing the production of IgA, IgG and IgM (Yamada *et al.*, 1996; Hung *et al.*, 1997). Since the enhancement of IgE production by unsaturated fatty acids is cancelled by hydrophobic antioxidants such as α -tocopherol, lipid peroxidation in a hydrophobic circumstance seems to be responsible for the enhancement of IgE production. In addition, tea polyphenols have also been reported to regulate Ig production of rat lymphocytes in a class specific manner (Yamada *et al.*, 1996). These results suggest that lipid metabolism is deeply related to the regulation of Ig production, as well as eicosanoid production.

Glutathione (GSH) is a tripeptide (Glu–Cys–Ala) widely present in micro-organisms,

as well as in animals and plants. It exerts diverse physiological functions such as detoxification in the liver, production and metabolism of various biomaterials, suppression of peroxidation and radiation disorders, protection of reactive thiol group. In the present study, we examined the dietary effect of GSH and GSH-rich yeast extract on lipid metabolism and immune function of Sprague-Dawley rats.

MATERIALS AND METHODS

Animals and diets

Yeast extract YH (YH) is a GSH-rich yeast extract afforded by Kohjin (Tokyo, Japan). Four-week-old male Sprague-Dawley rats were obtained from Seac Yoshitomi (Yoshitomi, Japan) and individually housed in the room of Biotron Institute, Kyushu University, with a controlled temperature of 20–23 °C and light from 08:00 to 20:00. After acclimatizing for 7 days, the rats were divided into 4 groups with 5 animals each, and free access was provided to the experimental diets and to deionized water. The diets were prepared according to the recommendations of the American Institute of Nutrition AIN-93G, and contained 36.75% cornstarch, 20% casein, 13.2% α -cornstarch, 10% sucrose, 10% safflower oil, 5% cellulose, 3.5% mineral mixture, 1% vitamin mixture, 0.3% L-cystine, 0.25% choline bitartrate and 0.0014% *tert*-butylhydroquinone. YH were added to the diet at the 1 or 10% level in place of cornstarch, and GSH at the 0.01% level which is corresponding to the level in 1% YH diet.

After three weeks of feeding, the rats were killed by withdrawing blood from the abdominal aorta under diethyl ether anesthesia. The heart, lung, liver, kidney, brain, spleen and epididymal adipose tissue of each rat were immediately excised and weighed. Lymphocytes were isolated from the spleen and mesenteric lymph node (MLN) to measure their Ig productivity. The levels of serum cholesterol, triglycerides and phospholipids were enzymatically determined with commercial kits, such as Cholesterol Test, TG-G Test and PL-B Test (Wako Pure Chemicals, Osaka, Japan). A thiobarbituric acid test kit was purchased from Wako and the amount of thiobarbituric acid-reactive substance (TBARS) was measured according to the directions of supplier. This experiment was carried out under the guidelines for Animal Experiments in the Faculty of Agriculture and Graduate Course of Kyushu University, and according Law No. 105 and Notification No. 6 of the Japanese government.

Cells and cell culture

Spleen and MLN lymphocytes were isolated from the rats that had been fed the above diets for 3 weeks, using Lympholyte-rat (Cedarlane, Hornby Canada), and cultured for 24 hr in the RPMI 1640 medium (Nissui Pharmaceutical, Tokyo, Japan) supplemented with 10% fetal bovine serum (FBS; Intergen, Purchase, NY) as described previously (Lim *et al.*, 1994). The IgA, IgG and IgM contents in the serum and culture supernatant were determined by the enzyme-linked immunosorbent assay (ELISA) as described previously (Lim *et al.*, 1994).

Statistics

Data were analyzed by Duncan's new multiple-range test (Duncan, 1955) to deter-

mine the exact nature of the differences among groups.

RESULTS AND DISCUSSION

Effect on growth and tissue weight

Table 1 shows the effect of YH and GSH feeding on food intake and weight gain. Weight gain of the rats fed 1% YH diet was higher than that of the rats fed control diet, and food intake of the former was lower than that of the latter. However, there was no significant difference in these indices and food efficiency.

Table 1. Effect of yeast extract or glutathione feeding on the growth of Sprague–Dawley rats.

	Initial body weight (g)	Weight gain (g)	Food intake (g/day)	Food efficiency (Weight gain/food intake)
Control	144±5	123±10	19.0±0.5	0.31±0.02
1% YH	144±2	132±3	18.4±0.2	0.34±0.01
10% YH	144±5	126±5	18.9±0.5	0.32±0.01
0.1% GSH	144±5	128±7	18.5±0.3	0.33±0.02

Data are means±SE (n=5).

On the other hand, significant changes of tissue weight were observed in some tissues (Table 2). Heart weight of the 1% YH group was significantly lower than that of the control rats, and 10% YH and 0.1%GSH groups gave intermediate values. In the case of kidney, 10%YH group gave a significantly higher value than control group, and the other groups gave intermediate values. Adipose tissue weights of 1% YH, 10% YH and 0.1% GSH groups were significantly higher than that of control rats. There was no significant effect in lung, liver, brain and spleen weights. These results suggest that the effect of YH or GSH intake was small on growth and tissue weight of Sprague–Dawley rats, except epididymal adipose tissue weight. The increase of adipose tissue weight of the 0.1% GSH group was smaller than those 1% and 10% YH groups, though there was no significant difference among them.

Table 2. Effect of yeast extract or glutathione feeding on the tissue weights of Sprague–Dawley rats.

	Tissue weight (g/100 g body weight)						
	Heart	Lung	Liver	Kidney	Brain	Spleen	Adipose
Control	0.37±0.01 ^a	0.47±0.01	4.10±0.19	0.81±0.03 ^{ab}	0.70±0.01	0.22±0.01	1.22±0.05 ^a
1% YH	0.32±0.01 ^b	0.46±0.01	4.18±0.12	0.81±0.02 ^{ab}	0.67±0.01	0.20±0.01	1.86±0.04 ^b
10% YH	0.33±0.01 ^{ab}	0.47±0.02	4.37±0.14	0.86±0.01 ^a	0.70±0.03	0.22±0.01	1.88±0.02 ^b
0.1% GSH	0.35±0.02 ^{ab}	0.51±0.05	3.91±0.16	0.78±0.02 ^a	0.65±0.02	0.22±0.01	1.77±0.01 ^b

Data are means±SE (n=5) and values without a common superscript letters are significantly different at $p<0.05$.

Effect on serum lipid levels

Table 3 shows the effect of YH and GSH feeding on serum lipid levels. Cholesterol levels of YH and GSH groups were lower than that of control group, but there was no significant difference among them. Triglyceride level of the 10% YH group was higher than that of control group and the level of 0.1% GSH group was lower than that of control rats. However, there was no significant difference among them. In addition, there was no significant difference in serum phospholipid level, as well as the level of TBARS, an index of lipid peroxidation. These results suggest that the effect of YH and GSH feeding was not so significant.

Table 3. Effect of yeast extract or glutathione feeding on serum lipid and TBARS levels of Sprague–Dawley rats.

	Cholesterol (mg/dl)	Triglyceride (mg/dl)	Phospholipids (mg/dl)	TBARS (ng/dl)
Control	65 ± 7	53 ± 13	148 ± 11	0.15 ± 0.02
1% YH	62 ± 3	57 ± 12	154 ± 6	0.20 ± 0.00
10% YH	59 ± 4	63 ± 16	156 ± 9	0.17 ± 0.02
0.1% GSH	56 ± 8	38 ± 7	141 ± 15	0.18 ± 0.01

Data are means ± SE (n=5).

Effect on immune functions

Table 4 shows the effect of YH or GSH feeding on immune indices. LTB₄ releasing activity of PEC isolated from the 10% YH group was significantly lower than those obtained from the rats of control and 1% YH groups. In 0.1% GSH group, enough number of PEC was not isolated. Such decrease of LTB₄ releasing activity has been reported in the rats fed n-3 unsaturated fatty acids (Sugano *et al.*, 1998; Hung *et al.*, 1999, 2000) or antioxidants (Gu *et al.*, 1995; Matsuo *et al.*, 2000). These fatty acids suppress the productivity of 4-series leukotrienes through the decrease of the level of arachidonic acids in membrane phospholipids. On the other hand, antioxidants such as tea polyphenols suppress the activity irrespective of arachidonic acid level, probably by inhibiting the activity of lipoxygenase (Matsuo *et al.*, 2000). Since the inhibitory activity was observed only in lipophilic antioxidants, permeation through cell membrane seems to be essential. Though the effect of GSH feeding was not clarified because of the lack of PEC in this experiment, hydrophilic GSH may not be effective to the suppression of LTB₄ production. Thus, it is possible that the other suppressive factors in YH is responsible for the reduction of LTB₄ releasing activity.

In the case of serum Ig levels, 10% YH group gave a significantly lower IgA level than other groups. On the other hand, 1% YH group gave a significantly lower IgG level than other groups. Serum IgM levels of YH and GSH groups were significantly lower than that of control rats. When spleen or MLN lymphocytes isolated from the rats fed these diets were cultured in the medium without dietary components, some significant differences were observed. In spleen, 10% YH and 0.1% GSH groups exerted IgA productivities sig-

Table 4. Effect of yeast extract or glutathione feeding on immune indices of Sprague–Dawley rats.

	Control	1% YH	10% YH	0.1% GSH
LTB ₄ releasing activity PEC (ng/10 ⁶ cells)	19.4±1.7 ^a	19.3±1.0 ^a	14.1±0.2 ^b	Not tested
Serum Ig level				
IgA (μg/ml)	28.7±0.5 ^a	28.2±0.3 ^a	25.0±0.5 ^b	27.5±0.8 ^a
IgG (mg/ml)	1.2±0.1 ^a	0.6±0.1 ^b	1.4±0.2 ^a	1.0±0.2 ^{ab}
IgM (μg/ml)	263±20 ^a	217±12 ^b	193±10 ^b	222±8 ^b
Ig productivity of spleen lymphocytes				
IgA (ng/ml)	20.5±0.5 ^a	18.5±1.0 ^{ab}	16.1±1.3 ^b	16.2±1.2 ^b
IgG (ng/ml)	48.6±2.6 ^a	44.3±3.0 ^{ab}	41.8±2.8 ^{ab}	37.9±2.6 ^b
IgM (ng/ml)	51.1±2.2 ^a	44.4±2.9 ^{ab}	40.2±4.6 ^b	45.8±1.0 ^{ab}
Ig productivity of MLN lymphocytes.				
IgA (ng/ml)	19.0±1.1	20.7±1.0	20.3±0.7	19.5±0.5
IgG (ng/ml)	40.2±3.0 ^{ab}	45.3±1.1 ^a	44.0±2.2 ^a	37.4±1.3 ^b
IgM (ng/ml)	2.5±1.2 ^a	3.4±1.1 ^b	2.6±0.8 ^a	0.7±0.3 ^c

Data are means±SE (n=5) and values without a common superscript letters are significantly different at $p<0.05$.

nificantly lower than control rats, 1% YH group gave an intermediate value. IgG productivity of the 0.1% GSH group was significantly lower than that of control rats, and YH groups gave intermediate values. On the other hand, IgM productivity of the 10% YH group was significantly lower than that of control rats, and 1% YH and 0.1% GSH groups gave intermediate values. In the case of MLN lymphocytes, there was no significant difference in IgA productivity. IgG productivity of the 0.1% GSH group was significantly lower than those of YH groups and control rats gave an intermediate value. In the case of IgM productivity, 1% YH group gave a value significantly higher than control rats and 0.1% GSH group gave a value significantly lower than control rats. These results suggest that YH and GSH feeding affect to Ig productivity of rat lymphocytes with different manners.

Similar modulation of Ig productivity has been reported in the rat fed water-soluble dietary fibers (Lim *et al.*, 1997; Yamada *et al.*, 1999). In this case, significant increase of serum IgA level was induced in the rats fed pectin, glucomannan or galactomannan, as well as the significant enhancement of IgA and IgG productivity in MLN lymphocytes. Such increase of IgA and IgG levels may be useful for the activation of self defensive system. On the contrary, YH or GSH feeding exerted a tendency to inhibit the production of these antibodies, though the effect was much weaker than the effect of dietary fibers. To estimate physiological meaning of the Ig production-regulating activity of YH and GSH more correctly, further experiment should be necessary.

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