

Measurement of Cell Membrane Stability Evaluated by Electrolyte Leakage as a Drought and Heat Tolerance Test in Rice (*Oryza sativa* L.)

Agarie, Sakae

Laboratory of Practical Botany, Faculty of Agriculture, Kyushu University

Hanaoka, Naomi

Laboratory of Practical Botany, Faculty of Agriculture, Kyushu University

Kubota, Fumitake

Laboratory of Practical Botany, Faculty of Agriculture, Kyushu University

Agata, Waichi

Laboratory of Practical Botany, Faculty of Agriculture, Kyushu University

他

<https://doi.org/10.5109/24109>

出版情報：九州大学大学院農学研究院紀要. 40 (1/2), pp.233-240, 1995-12. Kyushu University
バージョン：
権利関係：



Measurement of Cell Membrane Stability Evaluated by Electrolyte Leakage as a Drought and Heat Tolerance Test in Rice (*Oryza sativa* L.)

Sakae Agarie, Naomi Hanaoka, Fumitake Kubota,
Waichi Agata and Peter B. Kaufman*

Laboratory of Practical Botany, Faculty of Agriculture, Kyushu University
Fukuoka 812, Japan.

(Received June 4, 1995)

To explore possibilities of adaptation of cell membrane stability measurement as a evaluation of drought and heat tolerance (or injury) in rice (*Oryza sativa* L.), we have measured electrolyte leakage (EL) from leaf discs exposed to desiccation and temperature stresses. In desiccation test with polyethylene glycol (PEG), EL increased from 10% to 80% with increasing from 20% to 60% of PEG concentration [PEG]. We have used 30% of [PEG] at which 25% of EL was occurred. EL was increased with the time of PEG treatment; accordingly, 34 hours as a period of submerged leaf were selected. We have investigated an optimal temperature and treatment period, and around 42°C is likely to be suitable for testing membrane thermostability. EL was increased almost linearly with the time of leaf tissues submerged in test temperature (42°C) solution; we selected 24 hours for the following experiments. EL of PEG test was greater in senescing leaf (the 4th leaf from the youngest fully expanded leaf) than leaf in which senescence is less advanced (the youngest, fully expanded leaf). EL of leaf tissue was also increased with water deficit, indicating physiological status of plant itself can be reflected by the technique. Therefore, this technique is considered to be useful to assess injury, presumably at the membrane level, resulting from an environmental stress in rice.

INTRODUCTION

Plant survival and production under environmental stress are conditioned by complex mechanisms. Many studies point to cell membrane as an initial site of stress injury, the function and structure of plant cell membranes is drastically damaged by environmental stress (Liebermann *et al.*, 1958; Mckersie and Tomes 1980; Mckersie *et al.*, 1982; Siminovich *et al.*, 1964). Thus, evaluation of cellular membrane integrity as a measure of environmental stress tolerance appears to be relevant criterion (Sullivan, 1972).

Most commonly, changes in the electrical impedance and electrolyte leakage have been measured to detect stress injury of cell membrane. Leakage will vary in relation to the membranes' abilities to take up and retain solutes and, therefore, will reflect stress-induced changes in both membrane potentials and membrane permeability. This technique is a modification of a method developed by Dexter *et al.* (1932) and Dexter (1956) for measuring freezing resistance. It is based on desiccation or heat shock in vitro of leaf discs by a solution including osmotica or adjusted temperature, and subsequent measurement of electrolyte leakage expressed as specific conductance of the aqueous bathing solution in which the discs were immersed, indicating degree of damage of cell membrane resulting from these treatments.

* Department of Biology, University of Michigan, Ann Arbor, Michigan 48109-1048 USA.

Sullivan and Ross (1979) have conducted many experiments concerned with the relationship between electrolyte leakage following a desiccation treatment and the general ability of the plants to tolerate stress, and they found that membrane integrity and stability to the stress as evaluated by electrolyte leakage correlate well with tolerance of other plant process to the stress. To date, this method has been successfully used to measure membrane integrity in plants subjected to a variety of environmental stress (Liebermann *et al.*, 1958; Mckersie and Tomes, 1980; Mckersie *et al.*, 1982; Siminovich *et al.*, 1964). This method was found to be efficient in estimating stress tolerance of several crop plants, including wheat (*Triticum aestivum* L.) (Blum and Ebercon, 1981; Premachandra and Shimada, 1987a), orchardgrass (*Doctylis glomerata* L.) (Premachandra and Shimada, 1987b), and maize (*Zea mays* L.) (Premachandra *et al.*, 1989). However, no attempt has been made to be established in rice (*Oryza sativa* L.).

Electrolyte leakage measured was markedly influenced by age, sampling part and season, degree of stress hardening, and plant species. Therefore, these factors should be taken into consideration at the measurement. In the present study, our objective was to explore possibilities of adaptation or cell membrane stability measurement as a measure of drought and heat tolerance in rice with a few alterations to Sullivan's methods.

MATERIAL AND METHODS

Plant material

Rice cultivar, Koshihikari was water-cultured using Yoshida's standard nutrient solution (Yoshida *et al.*, 1976) containing 100ppm of SiO₂ in a green house. The culture solution was renewed once a week, and pH of the solution was adjusted to 5.0 to 5.5 with 1N NaOH and 1N HCl. Three samples were subjected to drought stress which consisted of exposing to 0.2 M mannitol (-0.68MPa of water potential) until leaf water potentials reached approximately 70% of control plant on the average. To conduct Si-deficient treatment, four plants were cultured in Yoshida's standard nutrient solution without SiO₂.

Measurement of electrolyte leakage

Polyethylene glycol test

Leaf samples were taken from the uppermost fully expanded leaves and were cut into 1 cm pieces. Thirty or twenty leaf pieces were put into a 100ml flask and washed slowly with three changes of deionized distilled water to remove surface adhered electrolytes. Following the washing, the leaf pieces were submerged in 30ml of polyethylene glycol (PEG) solution for a given period at 10°C in the dark to minimize secondary effects. PEG 600 was used in this study. The concentration of the solution and the duration were adjusted to the severity of desiccation desired. After the treatment period, the leaf pieces were washed quickly for three times with deionized distilled water. Thirty ml of deionized distilled water were then added and kept for 24 hours at 10°C in the dark. Then the flask was warmed to 25°C, shaken well and the electrical conductivity was measured. Following the conductivity measurement, the leaf tissues were killed by autoclaving for 15 minutes to release all ions from the tissue, cooled to 25°C and then the electrical conductivity was measured. Degree of electrical leakage (EL) is evaluated using following

equation:

$$EL = (EC_i / EC_t - C_i / C_t) \times 100$$

where, EC_i is the initial electrical conductivity, EC_t is the total electrical conductivity, C_i is the initial electrical conductivity of non-desiccated control. C_t is the total electrical conductivity of the control.

Heat shock treatment

Procedures and calculations were as above. Instead of incubating samples with an osmotic medium, ten leaf pieces were heated in 30ml of deionized water at 30, 35, 40, 45°C for 24 hours in the dark. After the treatment period, the electrical conductivity was measured according to PEG treatment.

RESULTS AND DISCUSSION

PEG test for estimating cell membrane stability is long-standing method for assessing drought tolerance. This method is based on the measurement of the electroconductivity of aqueous media containing leaf discs that were previously water stressed in vitro by exposure to a solution of PEG. Therefore, the values obtained with PEG test have been considered to be influenced by factors in terms of membrane permeability such as leaf surface wax content, thickness of cuticular layer and cell wall, changes in cytoplasmic lipids, and osmotic adjustment (Premachandra *et al.*, 1989, 1992).

In this study, first, we have investigated the relationship between PEG concentration ([PEG]) and electrolyte leakage (EL) of the leaf tissues (Fig. 1). EL increased from 10% to 80% with increasing from 20% to 60% of [PEG]. For testing drought tolerance in

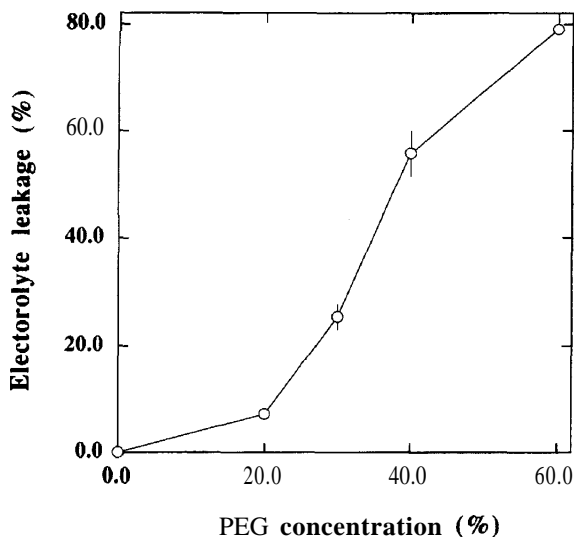


Fig. 1. Relationship between PEG concentration and electrolyte leakage.

orchardgrass, Premachandra and Shimada (1987b) used 50% of [PEG] at which the difference between cultivars was maximum. In sorghum, 30% of [PEG] was used in the same reason (Premachandra *et al.*, 1992). The similar degree of leakage was occurred at 30% of [PEG] in rice leaves, therefore, this [PEG] was used as a standard osmotic medium for further work in rice.

With the time of submerging leaves in PEG solution, EL was increased (Fig. 2). Accordingly, 24 hours were selected for the following experiments. An experiment was also conducted to find out the suitable number of the leaf pieces. The differences of injury among the two samples (10 and 20 leaf pieces) were small and not statistically significant (data not shown).

Although the mechanisms probably differ, drought resistance are often positively correlated with heat resistance in plants. Sullivan (1971, 1972) has described a technique employing leaf discs for evaluating tolerance to high temperature in terms of membrane thermostability. This method is based on heat shock in vitro of leaf discs by a solution adjusted temperature and subsequent measurement of EL from plant tissue into an aqueous medium.

Since the amount of the EL is a function of membrane permeability, the membrane thermostability (the degree of damage resulting from high temperature) can be assessed in terms of electrolytic conductance. The results obtained with this technique correlate with degree of the heat stability of photosynthetic activity in isolated chloroplasts (Sullivan *et al.*, 1977) and with degree of heat tolerance of whole plants (Sullivan, 1972). Therefore, this technique considered to be useful in investigating the membrane competence as affected by temperature stress.

In this study, we have investigated a optimal temperature and treatment period to applicate this technique on rice. Martineau *et al.* (1979) have tested membrane

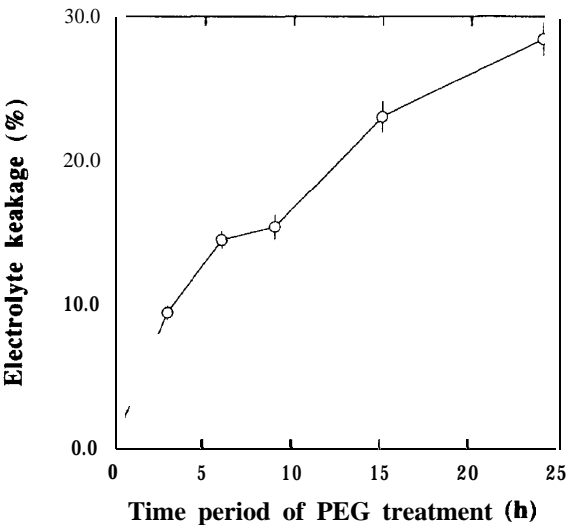


Fig.2. Effect of time period of 30% PEG treatment on electrolyte leakage.

thermostability of soybean in the range of 30°C to 60°C, and they have rated around 48°C at which 50% of EL was occurred a suitable temperature in soybean leaves. We have tested the leakage in the range of temperature 30°C to 45°C to test the thermostability (Fig. 3). EL increased from 8.1% to 71.6% with temperature increase from 40°C to 45°C. Therefore, around 42°C is likely to be suitable for testing membrane thermostability in rice leaves.

With the time of leaf tissues submerged in test temperature (42°C) solution, EL was

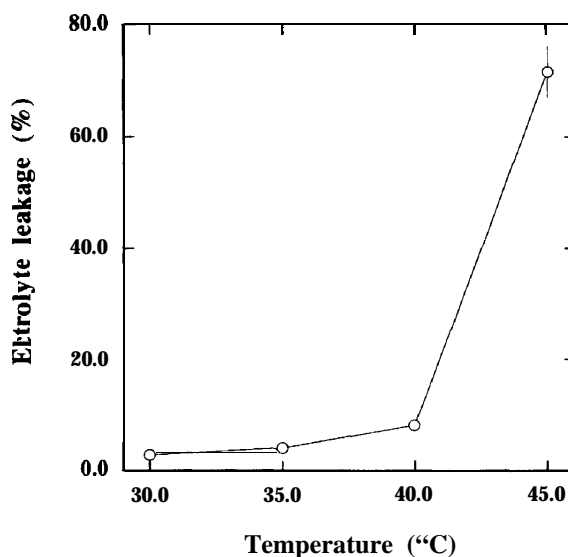


Fig. 3. Relationship between temperature of test solution and electrolyte leakage.

increased almost linearly (Fig. 4), we selected 24 hours for the following experiments.

To test the ability of this technique to correctly describe membrane leakage expressed physiological status of plant itself, we selected model systems representing examples of physiological processes with well-known effects on membrane permeability (Table 1). These phenomena were water stress and senescence that have been shown to make membrane more leaky. In addition, we compared the leakage of silicated leaves with leaves grown under silica deficient condition, because silica is considered to increase the tolerance to environmental stress of plants, such as salt (Ahadam *et al.*, 1992), drought (Emadian and Newton, 1989), and temperature (Larcher *et al.*, 1991).

EL of PEG test was greater in senescing leaf (the 4th leaf from the youngest fully expanded leaf) than leaf in which senescence is less advanced (the youngest fully expanded leaf). Blum and Ebercon (1976, 1981) and Sullivan (1972) have reported that the younger leaf tissues are more tolerant to drought than older tissues in wheat and sorghum, respectively. Our results were well comparable with their results. The concept that membrane failure may be involved in leaf senescence was first suggested by Sacher

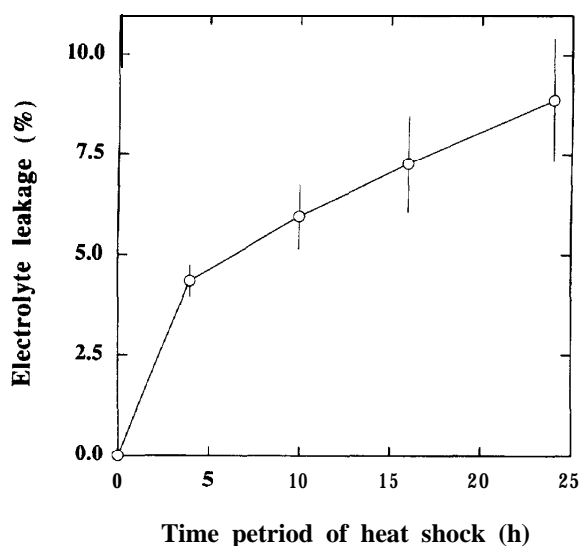


Fig. 4. Effect of time period of heat shock with 42°C test solution on electrolyte leakage.

Table 1. Effects of senescence, water stress and silicon on electrolyte leakage.

| Leaf | Electrolyte leakage (%) |
|-----------|-------------------------|
| control | 29.1±2.9 |
| senescing | 33.7±1.8 |
| stress | 38.4±1.8 |
| -Si | 38.6±2.1 |

Control, the youngest fully expanded leaf; senescing, the fourth leaf from the control leaf; stress, water-stressed leaf of the control leaf; -Si, the youngest fully expanded leaf grown in the nutrient solution without SiO_2 .

(1957). Electron microscopy revealed a decrease in plastid volume and disruption of the internal mambrane system from senescing leaves (Butler and Simon, 1970). Biochemical studies showed that along with these morphological changes, there is change in the lipid composition of the membrane during senescence (Barton, 1966). The senescence-induced injury is likely to be assessed by the measurement of EL.

EL of leaf tissue was also increased with water deficit of leaf. Water deficits profoundly alter both the structure and the functions of membranes, leading to destructive events such as transition from liquid-crystalline to gel phase, fusion and increased permeability (Crowe *et al.*, 1987; Ferrari-Ilion *et al.*, 1984; Maroti *et al.*, 1984;

Senaratna *et al.*, 1984). This results indicate that EL reflect stress-induced changes in the condition of cellular membranes.

It has been well documented that dry matter production and grain yield of rice is significantly increased by silica application to rice paddies. The growth-promoting effects by silicon application was significant when the plants were grown under stress-inducing conditions, especially drought. This indicate that silica involved in the stress tolerance, assumably with increasing cell membrane stability to water stress condition, In this study, EL was increased in Si deficit leaf to the same levels as water-stressed leaf, indicating silicon has an effect on increasing cell rigidity related to a building drought tolerance.

Collectively, these experiments indicate that the EL from tissues can evaluate the damage to membrane incurred by stress experiences, and thus, this technique would provide useful information to describe the behavior of plant tissues against environmental stresses in rice.

ACKNOWLEDGEMENTS

The authors wish to thank Dr. H. Saneoka for his valuable suggestion.

REFERENCE

- Ahadam, R., S. H. Zaheer and S. Ismail 1992 Role of silicon in salt tolerance of wheat (*Triticum aestivum* L.). *Plant sci.* 85: 43-50
- Barton, R. 1966 Fine structure of mesophyll cells in senescing leaves of *Phaseolus*. *Planta*, 71: 314-325
- Blum, A. and A. Ebercon 1976 Genotypic responses in sorghum to drought stress III. Free proline accumulation and drought resistance. *Crop Sci.*, 16: 428-431
- Blum, A. and A. Ebercon 1981 Cell membrane stability as a measure of drought and heat tolerance in wheat, *Crop Sci.*, 21: 43-47
- Butler, R. D. and E. W. Simon 1970 Ultrastructure aspects of senescence in plants. *Adv. Gerontol. Res.*, 3: 73-129
- Crowe, J. H., L. M. Crowe, J. F. Carpenter and C. Aurell Wistrom 1987 Stabilization of dry phospholipid bilayers and proteins by sugars. *Biochem. J.*, 242: 1-10
- Dexter, S. T., W. E. Tottingham and L. H. Graber 1932 Investigations of the hardiness of the plants by measurement of electrical conductivity. *Plant Physiol.*, 7: 63-78
- Dexter, S. T., 1956 Evaluation of crop plants for winter hardiness. *Adv. Agron.*, 8: 203-209
- Emadian S. F. and R. J. Newton 1989 Growth enhancement of loblolly pine (*Pinus taeda* L.) seedlings by silicon. *J. Plant Physiol.*, 134: 98-103
- Ferrari-Illion, R., A. T. Pham-Thi and J. V. Da Silva 1984 Effect of water stress on the lipid and fatty acid composition of cotton (*Gossypium hirsutum*) chloroplasts. *Physiol. Plant.*, 62: 219-224
- Larcher W., U. Meindl, E. Ralsler and M. Ishikawa 1991 Persistent supercooling and silica deposition in cell walls of palm leaves. *J. Plant Physiol.* 139: 146-154
- Liebermann M., C. C. Craft, W. V. Audia and MS. Wilcox 1958 Biochemical studies of chilling injury in sweet potatoes. *Plant Physiol.*, 33: 307-311
- Maroti, I., Z. Tuba, and M. Csik 1984 Changes in chloroplast ultrastructure and carbohydrate level in *Festuca*, *Achillea*, and *Sedum* during drought and recovery. *J. Plant Physiol.*, 116: 1-10
- Martineau, J. R., J. E. Specht, J. H. Williams and C. Y. Sullivan 1979 Temperature tolerance in soybeans. I. Evaluation of a technique for assessing cellular membrane thermostability. *Crop Sci.*, 19: 75-78
- Mckersie B. D. and D. T. Tomes 1980 Effect of dehydration treatment on germination, vigor, and

- cytoplasmic leakage in wild oats and birdsfoot trefoil. *Can.J. Bot.*, 58: 471-476
- Mckersie, B. D., P. Hucl and W. D. Beversdorf 1982 Solute leakage from susceptible and tolerant cultivars of *Phaseolus vulgaris* following ozone exposure. *Can.J. Bot.*, 60: 73-78
- Premachandra, G.S., H. Saneoka, K. Fujita and S. Ogata 1992 Leaf water relations, osmotic adjustment, cell membrane stability, epicuticular wax load and growth as affected by increasing water deficits in sorghum. *J. Exp. Bot.*, 43: 1569-1576
- Premachandara, G. S., H. Saneoka and S. Ogata 1989 Nutrio-physiological evaluation of polyethylene glycol test of cell membrane stability in maize. *Crop Sci.*, 29: 1287-1292
- Premachandra, G. S. and T. Shimada 1987a The measurement of cell membrane stability using polyethylene glycol as a drought tolerance test in wheat. *Japan. Jour. Crop Sci.*, 56: 92-98
- Premachandra, G. S. and T. Shimada 1987b Measurement of drought tolerance in orchardgrass (*Dactylis glomerata* L.). I. Polyethylene glycol test of measuring cell membrane stability. *J. Japan Grassl. Sci.*, 33: 140-148
- Sacher, J. A. 1957 Relationship between auxin and membrane-integrity in tissue senescence and abscission. *Science*, 125: 1199-1200
- Senaratna, T., B. Mckersie and R. H. Stinson 1984 Association between membrane phase properties and dehydration in soybean axes. *Plant Physiol.*, 76: 756-762
- Siminovich, D., H. Therrien, F. Gfeller, and B. Rheume 1964 The quantitative estimation of frost injury and resistance. *Can. J. Bot.* 42: 637-640
- Sullivan, C. Y. 1971 Techniques for measuring plant drought stress. In "Drought injury and resistance in crops", ed. by K. L. Larson and J. D. Eastin, Crop Sci. Soc. Ame., Madison, Wis.
- Sullivan, C.Y. 1972 Mechanisms of heat and drought resistance in grain sorghum and methods of measurement. In "Sorghum in the Seventies" Ed. by N. G. P. Rao and L. R. House, Oxford and IBH publishing Co., New Delhi, India.
- Sullivan, C. Y., N. V. Norcio, and J. D. Eastin 1977 Plant response to high temperature. In "Genetic diversity in plants", eds by A. Muhammed, R. Aksel, and R. C. Von Borstel, Plenum Publishing Corp. pp. 301-317
- Sullivan, C. Y. and W.M. Ross 1979 Selection for drought and heat resistance in grain sorghum. In "Stress physiology in crop plants", ed. by H. Hüssel and R. Staples, John Wiley and sons, New York pp. 263-281
- Yoshida, S., A. F. Douglas, H.C. James and A. G. Kwanchai 1976 *Laboratory Manual for Physiological Studies of Rice*. IRRI press, Manila, Philippine.