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Development of Serum-free Medium for Clonal Growth of Human-human Hybridomas

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The cloning of human-human hybridomas in serum-free medium may be beneficial to industry and research. Currently, the use of serum is essential when human-human hybridomas are cloned. To develop serum-free medium for cloning human-human hybridomas, we screened a variety of compounds to determine their effects on the clonal growth of NAT-30 cells, which are derived from the human Burkitt lymphoma Namalwa. These effective compounds were added to ERDF medium supplemented with insulin, transferrin, ethanolamine and sodium selenite (ITES-ERDF). Bovine serum albumin (BSA) and α -tocopherol elevated clonal cell growth remarkably. Casein significantly stimulated the growth of NAT-30 cells at relatively low cell densities. Almost all human-human hybridomas, which were not able to proliferate in ITES-ERDF medium alone, could grow in ITES-ERDF medium supplemented with BSA, α -tocopherol, and casein (ITES-BTC-ERDF medium). Cloning efficiencies of human-human hybridomas cultured in the ITES-BTC-ERDF medium were as high as when cultured in medium supplemented with 15% fetal calf serum.

INTRODUCTION

Culturing hybridomas in serum-free medium has many advantages (easy purification of monoclonal antibodies (MAbs), low medium cost, medium quality stability, etc.) for the mass-production of MAbs. ERDF medium supplemented with insulin, transferrin, ethanolamine and sodium selenite (ITES-ERDF) has been widely used as a serum-free culture medium for lymphocytes and hybridomas (Murakami *et al.*, 1982; Murakami, 1989). Establishment of human fusion partners such as NAT-30 (Murakami *et al.*, 1985), HO-323 (Ohashi *et al.*, 1986) and A4H12 (Kawahara *et al.*, 1992) make it possible to produce many human-human hybridomas reactive to various human antigens (Hashizume *et al.*, 1987; Shinmoto *et al.*, 1988; Aihara *et al.*, 1988). However, ITES-ERDF medium not only does not support the growth of some hybridomas, but the cloning of lymphoid cells and hybridomas in this serum-free medium is impossible because of poor cell growth at low cell densities. Even hybridomas which can grow in ITES-ERDF medium must use serum-supplemented medium for cloning, which may result in an unfavorable selection of cells. Serum is also expensive and cloning efficiencies vary from serum lot to lot.

This background prompted us to develop a new serum-free medium that will allow for the cloning of human-human hybridomas. There are several factors which affect cloning efficiencies: (1) damage by oxygen radicals, (2) deficiencies of various autocrine growth factors, and (3) physical cell damages. Among these factors, the most important factor

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negatively influencing clonal growth seems to be the damage caused by oxygen radicals (Darfler & Insel, 1983), particularly in serum-free medium which lacks free radical capturing potency (Mulholland & Strain, 1991). Clonal growth of lymphoid cells is known to be inhibited by H_2O_2 in serum-free medium which produces hydroxyl radical (Darfler & Insel, 1983). It has been reported that reducing agents such as mercaptoethanol (Kan & Yamane, 1982) and anti-oxidant reagents such as α -tocopherol (Gavino *et al.*, 1981) are able to elevate the cloning efficiency. Human hybridomas are generally more susceptible to chemical and physical damage than murine hybridomas. In fact, as far as we know, there has been only one report which briefly describes the development of a serum-free medium in which the cloning of a human hybridoma cell line is possible.

This paper reports that ITES-ERDF medium supplemented with BSA, α -tocopherol, and casein can clone human-human hybridomas as efficiently as serum-supplemented medium, and is able to support serial passages of human-human hybridomas which are not able to proliferate in ITES-ERDF medium alone.

MATERIALS AND METHODS

Reagents

BSA (fraction V, Sigma, U.S.A.) and casein (Wako Pure Chemicals, Japan) were dissolved in phosphate-buffered saline (PBS). Egg yolk lipoprotein (YLP) was prepared by the method described previously (Murakami *et al.*, 1988). α -Tocopherol, ascorbic acid, dithiothreitol and 2-mercaptoethanol were purchased from Wako Pure Chemicals and dissolved in ethanol. MTT reagent was obtained from Dojin Chemicals (Japan).

Cells and cell culture

NAT-30 cells are a 6-thioguanin-resistant clone derived from human Burkitt lymphoma Namalwa cells (Murakami *et al.*, 1985). HB4C5 and HF10B4 cells are human-human hybridomas obtained by the fusion of NAT-30 cells with human lymph node lymphocytes. HO-323 cells were established as a parent cell line from the human lymphoblast WIL2-NS (Ohashi *et al.*, 1986). EMK-F7 cells are hybridomas produced by the fusion of HO-323 cells with human lymph node lymphocytes (Koyama *et al.*, 1990). BD9-D12 cells are hybridomas produced by the fusion of A4H12 cells, a human parent cell line with human lymph node lymphocytes (Kawahara *et al.*, 1992).

NAT-30 cells and hybridomas derived from NAT-30 cells were cultured in ERDF medium supplemented with ITES (insulin, 5 μ g/ml; transferrin, 10 μ g/ml; ethanolamine, 25 μ M; and sodium selenite, 2 nM) at 37°C in a humidified atmosphere of 95% air/5% CO_2 . Other cells were cultured in ITES-ERDF medium supplemented with 50 μ g/ml of BSA, 1 μ M of α -tocopherol and 50 μ g/ml of casein.

Measurement of cell growth

MTT assay was used to measure the growth stimulating effect of various compounds on cultured cells (Mosmann, 1983).

Screening of compounds promoting clonal cell growth in serum-free medium

Compounds which promote cloning efficiencies were screened using two methods; limiting dilution method and colony formation assay. In the limiting dilution method, cells were plated at 1 cell/well in a 96-well microplate in serum-free medium containing the compounds to be examined. After incubating for 7 days, the number of wells containing growing cells were counted. Simultaneously, a more convenient assay method for detecting colony formation was also developed. Cells were plated at 10 cells/well in a 96-well microplate in serum-free medium containing the compounds to be examined. After statically incubating the plate for 7 days, the number of colonies formed was counted using a microscope and the colony formation ratio (%) was calculated.

RESULTS

Screening of compounds which are able to promote the growth of NAT-30 cells in serum-free medium

Effects of various compounds on the growth of NAT-30 cells at a low cell density were examined in ITES-ERDF medium. As shown in Fig. 1, casein remarkably stimulated the growth of NAT-30 cells at a low cell density (3×10^3 cells/ml) when applied in a wide range of concentrations (1 - 100 μ M). YLP showed a maximum stimulating effect at 10 μ g/ml, but caused a severe cell damage at concentrations higher than 20 μ g/ml. BSA exhibited a weak growth stimulating effect. Antioxidants such as α -tocopherol and

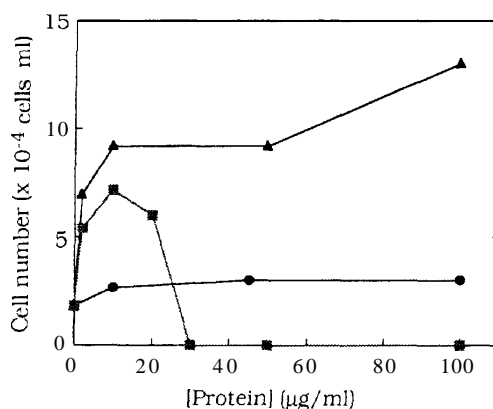


Fig. 1. Effects of BSA, casein and YLP on the growth of NAT-30 cells. NAT-30 cells were seeded in a 96-well microplate at a concentration of 3×10^3 cells/well in 0.1 ml of ITES-ERDF medium containing various concentrations of BSA, casein and YLP. After incubation for 4 days, the relative cell numbers were measured using the MTT method. Average value in triplicate is shown in the figure. Standard deviation (S.D.) is within 10%. ●, BSA; ▲, casein; ■, YLP

ascorbic acid and reducing agents such as 2-mercaptoethanol and dithiothreitol also showed weak growth stimulating effects (data not shown).

Screening of compounds which are able to promote the clonal growth of NAT-30 cells in serum-free medium

Next, effects of various compounds on the clonal growth of NAT-30 cells were examined using the colony formation assay. As shown in Fig. 2A, BSA stimulated the

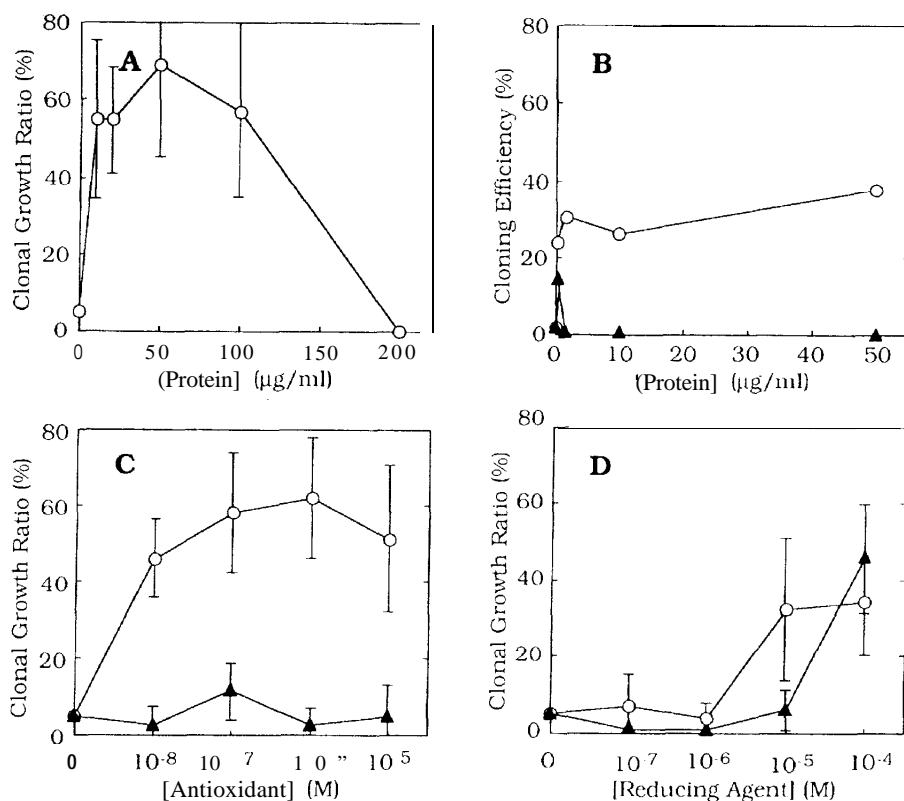


Fig. 2. Effects of BSA, casein, YLP, α -tocopherol, ascorbic acid, dithiothreitol and 2-mercaptoethanol on the clonal growth of NAT-30 cells. (A) NAT-30 cells were plated in a 96-well microplate at a concentration of 10 cells/well in ITES-ERDF medium containing various concentrations of BSA. After 7 day, the colonies formed were counted. Clonal growth ratio (%) was calculated by dividing the number of colonies by 10. Vertical bar represents S.D. (N=6). (B) Cells were plated in a 96-well microplate at a density of 1 cell/well in ITES-ERDF medium containing various concentrations of casein or YLP. After 7 days, the percentage of wells containing growing cells n-w-r determined. Average value in triplicate is shown in the figure. S.D. is within 10%. ○, casein, A, YLP. (C) Effects of α -tocopherol and ascorbic acid on the clonal growth of NAT-30 cells n-err examined under the same conditions 3s in (A). ○, α -tocopherol; ▲, ascorbic acid. Vertical bar represents S.D. (N=6). (D) Effects of reducing agents, dithiothreitol and 2-mercaptoethanol were examined under the same conditions as in (A). ○, dithiothreitol; A, 2-mercaptoethanol. Vertical bar represents S.D. (N=6).

clonal growth of NAT-30 cells noticeably. The maximum colony formation ratio (60%) was obtained at a BSA concentration of 50 $\mu\text{g/ml}$ and cell damage was observed at concentrations over 200 $\mu\text{g/ml}$. Since casein caused a tendency for cell to move to the edge of the well at concentrations higher than 50 $\mu\text{g/ml}$, cloning efficiencies were examined using the limiting dilution method, instead of the colony formation assay. Casein was shown to elevate the cloning efficiency at concentrations higher than 1 $\mu\text{g/ml}$ (Fig. 2B). However, YLP, which strongly stimulates cell growth, inhibited clonal growth completely at concentrations higher than 1 $\mu\text{g/ml}$. As shown in Fig. 2c, α -tocopherol, an antioxidant, exceptionally elevated the colony formation ratio to 60% at 1 μM . Whereas, ascorbic acid, an another antioxidant, showed no stimulating activity. Reducing agents such as 2-mercaptoethanol and dithiothreitol elevated the colony formation ratio to 40% at a relatively high concentration of 100 μM (Fig. 2D).

Next, the combined effects of BSA and α -tocopherol were examined. As shown in Fig. 3, supplementation of both BSA and α -tocopherol to the ITES-ERDF medium resulted in a high colony formation ratio of about 80%. Further addition of casein to the ITES-ERDF medium supplemented with BSA and α -tocopherol did not lowered the cloning efficiency, but addition of YLP inhibited the clonal growth completely at concentrations higher than 10 $\mu\text{g/ml}$ (data not shown).

From the results obtained, the supplementation of BSA (50 $\mu\text{g/ml}$), α -tocopherol (1 μM) and casein (50 $\mu\text{g/ml}$) to ITES-ERDF medium was very effective for elevating both the cloning efficiency and growth rate.

Growth and cloning efficiencies of various kinds of cell lines in ITES-ERDF medium supplemented with BSA, α -tocopherol and casein

The growth rate of various cell lines were examined in ITES-ERDF medium

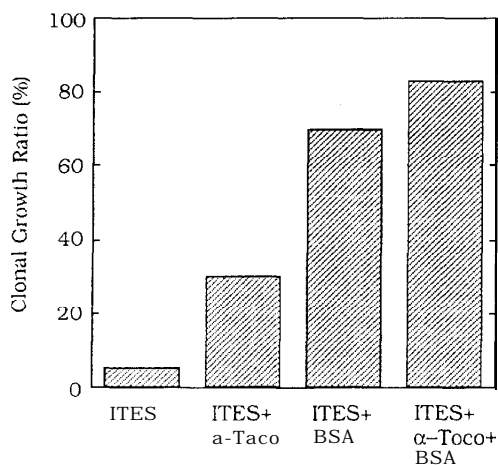


Fig. 3. Clonal growth ratio of NAT-30 cells in ITES-ERDF medium supplemented with α -tocopherol and BSA. The experimental conditions are the same as in Fig. 2 (A). α -Tocopherol, 1 μM ; BSA, 50 $\mu\text{g/ml}$.

supplemented with BSA (50 µg/ml), α -tocopherol (1 µM) and casein (50 µg/ml) (ITES-BTC-ERDF medium). NAT-30 cells and the two hybridomas derived from these cells, all of which were able to grow in ITES-ERDF medium, could also actively proliferate in ITES-BTC-ERDF medium. When plated at a density of 1×10^4 cells/ml, the growth rates of cells grown in ITES-BTC-ERDF medium were lower than cells grown in ERDF medium supplemented with 15% fetal bovine serum (FCS) (Fig. 4A). HO-323 cells and their hybridoma EMK-F7, and A4H12 cell and their hybridoma BD9D-12, could also grow well in ITES-BTC-ERDF medium, but grew poorly in ITES-ERDF medium (Fig. 4B and 4C). Serial passages of all cells mentioned above were possible for a long period of time, although the doubling time in ITES-BTC-ERDF medium was longer than that in serum-containing medium.

Next, the cloning efficiencies of these cells cultured in ITES-BTC-ERDF medium or 15% FCS-ERDF medium were examined. The results are shown in Table I. Hybridomas derived from NAT-30 and HO-323 cell lines in ITES-BTC-ERDF medium showed a similar cloning efficiency as in serum-containing medium. HF10B4 cells showed a 1.6-fold higher efficiency rate in ITES-BTC-ERDF medium than in serum-containing medium. A4H12 cells and their hybridomas derived from these cells showed lower cloning efficiencies in ITES-BTC-ERDF medium than in serum-containing medium. When O_2 concentration was lowered to 5%, the cloning efficiency of NAT-30 cells in ITES-BTC-ERDF medium elevated 1.6-fold, as compared to cells cultured in an atmosphere of 5% $CO_2/95\%$ air.

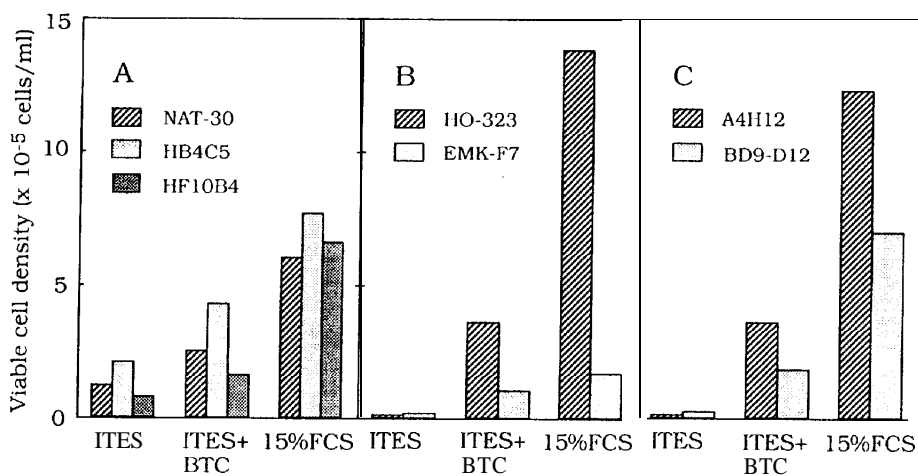


Fig. 4. Comparison of growth of lymphoid cells and their hybridomas in ITES-ERDF, ITES-BTC-ERDF, and 15% FCS-ERDF medium. Cells were plated in a 96-well microplate at a concentration of 1×10^4 cells/ml per well. After 4 days, the cell number was counted with a cell counter. Viability was measured using the trypan blue exclusion method. (A) Growth of NAT-30 cell line and its hybridomas, HB4C5 and HF10B4. (B) Growth of HO-323 cell line and its hybridoma EMK-F7. (C) Growth of A4H12 cell line and its hybridoma BD9-D12.

Table 1. Comparison of the cloning efficiencies of various lymphoid cells and hybridomas grown in ITES-BTC-ERDF medium and 15% FCS-ERDF medium.

	Cloning efficiencies (%) ^a		ITES-BTC/15% FCS
	ITES-BTC ^b	15% FCS	
Namalwa	88	31	2.80
NAT-30	40	40	1.00
NAT-30	63	58	1.07
HB4C5	67	50	0.94
HFIOB4	48	29	1.64
HO-323	54	54	1.08
EMK-F7	46	44	1.05
A4H12	58	67	0.88
BD9-D12	25	50	0.50

^a Cells subcultured in ITES-BTC-ERDF medium were plated on a 96-well microplate at a density of 1 cell/well in ITES-BTC-ERDF medium or in 15% FCS-ERDF medium under an atmosphere of 5% CO₂/95% air. After 7 days, the percentage of wells containing growing cells were determined.

^b ITES-BSA (50 µg/ml) + α -tocopherol (1 µM) + casein (50 µg/ml)

^c O₂ concentration decreased to 5%.

DISCUSSION

ITES-ERDF medium supports the cell growth of human lymphoid cells and human-human hybridomas at high cell densities (e. g. 1×10^6 cells/ml), however these same cells grow poorly at lower cell densities (e. g. 1×10^4 cells/ml). This stimulating effect of serum on cell growth when plated at a low cell density is not dependent upon differing serum lots. However cloning efficiencies of hybridomas in serum-supplemented medium varies dependent upon serum lot to lot. These facts suggest serum contains components which support both the cell growth at low cell densities and clonal cell growth. For example, human plasma and the conditioned medium of human-human hybridoma cells contain lymphocytic clonal growth factors (Miyata et al., 1988a; Miyata et al., 1988b).

For the development of a new serum-free medium which supports the cloning of human lymphoid cells and human-human hybridomas, we used three kinds of human fusion partners and their hybridomas. NAT-30 cells and hybridomas derived from this cell line can grow well in ITES-ERDF medium as well as in serum-supplemented medium. HO-323 cells can grow in ITES-ERDF medium, but hybridomas derived from this cell line need further supplementation, in particular YLP, to grow in the serum-free medium. A4H12 cells and hybridomas derived from these cells can not grow in serum-free medium. Among these cells, NAT-30 cells were used to screen for effective compounds that, when added to serum-free medium, may support the cloning of lymphoid cells and human-human hybridomas.

As a result of screening, casein and YLP were found to be effective to stimulate the cell growth of NAT-30 cells grown at a density of 3×10^3 cells/ml. Also, the clonal growth of NAT-30 cells was accelerated by BSA and α -tocopherol. Since YLP strongly inhibited the clonal growth of NAT-30 cells, casein, along with BSA and α -tocopherol, was chosen as supplements for our serum-free medium.

ITES-BTC-ERDF medium supported the cloning of almost all lymphoid cell lines and hybridomas. Although cell growth was slower than in 15% FCS-supplemented medium, all colonies grew actively. In addition, all lymphoid cell lines and hybridomas tested could also grow in ITES-BTC-ERDF medium. When cells were plated at density of 1×10^3 cells/ml, cell growth was slower in ITES-BTC-ERDF medium than in 15% FCS-supplemented medium. However, for cells plated at a density of 1×10^5 cells/ml in ITES-BTC-ERDF medium, the growth rate was comparable to cells growing in serum-containing medium. The fact that cloning cells under a 5% O_2 atmosphere resulted in an elevation of the cloning efficiency suggests that clonal growth is inhibited by oxygen radicals.

YLP stimulates the cell growth of lymphoid cells via the supply of fatty acids (Murakami *et al.*, 1988). However, the oxidation of fatty acids in medium will produce lipid peroxides, which generate oxygen radicals, in particular hydroxyl radicals, resulting in cell damage (Spieker-Polet & Polet, 1981; Wang *et al.*, 1991). High concentrations of YLP was toxic to NAT-30 cells, perhaps due to lipid peroxides. α -Tocopherol, an antioxidant, traps radicals in medium (Dean *et al.*, 1991). Ascorbic acid, another antioxidant, did not stimulate the clonal growth of NAT-30 cells. The instability of ascorbic acid may be related to its ineffectiveness (Border *et al.*, 1983).

Although trace amounts of fatty acids are indispensable for normal cell growth, the prevention of fatty acids oxidation plus the detoxification of oxygen radicals formed by fatty acids oxidation are important considerations to take into account when developing serum-free medium for cloning (Darfler, 1990). Darfler and Insel (1983) developed a serum-free medium supplemented with catalase, testosterone, and dilinoleoyl phosphatidylcholine to clone a human hybridoma cell line. However, catalase, testosterone and dilinoleoyl phosphatidylcholine are expensive making it inappropriate for use in industry.

Since the serum-free medium developed here supports both the cloning and the cell growth of almost all human-human hybridomas at low cell densities, analysis of MAb will become easier. The supply of stable and inexpensive serum-free medium will contribute to experimental reproducibility and facilitate mass and high density culture of human-human hybridomas, which before could not be cultured in serum-free medium.

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