# Absorption, Translocation and Assimilation of Ammoniumand Nitrate-Nitrogen in Rice Plants as Affected by the Supply of Potassium, Calcium and Magnesium

Ali, Abmed A. Laboratory of Plant Nutrition, Faculty of Agriculture, Kyushu University

Ikeda, Motoki Laboratory of Plant Nutrition, Faculty of Agriculture, Kyushu University

Yamada, Yoshio Laboratory of Plant Nutrition, Faculty of Agriculture, Kyushu University

https://doi.org/10.5109/23817

出版情報:九州大学大学院農学研究院紀要. 30 (2/3), pp.113-124, 1985-12. Kyushu University バージョン:

権利関係:

### Absorption, Translocation and Assimilation of Ammoniumand Nitrate-Nitrogen in Rice Plants as Affected by the Supply of Potassium, Calcium and Magnesium

#### Abmed A. Ali\*, Motoki Ikeda and Yoshio Yamada

Laboratory of Plant Nutrition, Faculty of Agriculture, Kyushu University 46-02, Fukuoka 812 (Received July 24, 1985)

Rice plants (*Oryza sativa* L., var. Asominori) were treated with complete, minus K (-K), Ca-limited (-Ca) and minus Mg(-Mg) nutrient solutions for 10 days. Absorption, translocation and assimilation of <sup>15</sup>NH<sub>4</sub>-N and <sup>15</sup>NO<sub>3</sub>-N fed for 2 days to the treated plants were investigated, and in *vivo* nitrate reductase activity (NRA) was intermittently determined after the initiation of the treatment. 1) Absorption of <sup>15</sup>NH<sub>4</sub>-N was not significantly different among treatment, while lower absorption of <sup>15</sup>NO<sub>3</sub>-N was observed in -K and -Ca treatment. 2) Translocation of <sup>15</sup>NH<sub>4</sub>-N from root to upper parts was not affected by treatment, but translocation of <sup>15</sup>NO<sub>3</sub>-N was inhibited by -K and -Ca treatment. 3) NRA of rice plants was decreased by -K and -Ca treatment, and repressed when NO,-N was replaced with NH,-N 'alone and in combination with NO,-N. 4) Assimilation of <sup>15</sup>NH<sub>4</sub>-N into amino acids and protein was increased by -K treatment, but assimilation of <sup>15</sup>NO<sub>3</sub>-N was inhibited by -K and -Ca treatment. These results indicate that nitrogen metabolism was disturbed by -K and -Ca treatment when rice plant was fed with NO,-N but not when plant fed with NH,-N.

#### INTRODUCTION

Most plant species are different in their ability to absorb NH<sub>4</sub>-N and NO,-N as nitrogen sources. The high ability of rice plant to absorb and utilize NH,-N has been reported by many investigators (Fried et al., 1965; Oji and Izawa, 1970; Dijkshoorn and Ismunadji, 1972). Absorption of ions from a culture medium may be influenced by other ions through effects of competition at the site of absorption. Ions more similar to NH,+ can compete with it and may decrease its absorption. Bange et al. (1965) found that K+ inhibited the NH,+ absorption by barley root. Recently, Rosen and Carison (1984) found that increasing K+ concentration had no significant effect on the NH,+ uptake by plants. Thus it seems that the influence of K+ on NH<sub>4</sub>+ absorption may be not yet fully understood. Nitrate absorption is usually associated with increased uptake and transport of cations (Minotti et al., 1968; Ben-Zioni et al., 1970; Ben-Zioni et al., 1971; Cox and Reisenauer, 1973; and Blevins et al., 1974). Matsumoto et al. (1980) observed that NO, content of cucumber leaves

<sup>\*</sup> Present Address: Faculty of Agriculture, Menoufia University, Shiben Elkom, Egypt

114 A. A. Ali et al.

was decreased by Ca deficincy. Burstrom (1954) and Shrader and Hageman (1967), however, claimed that Ca did not appear to be essential for absorption of  $NO_3$ -N. The relationship between Mg content and nitrogen metabolism has been reported by Webster (1961) and Haeder and Mengel (1969). The investigators frequently suggested that K deficiency inhibited protein synthesis in plants because amino acids accumulated (Koch and Mengel, 1974). On the other hand, Hsiao et al. (1970) and Besford (1975) did not find any direct effect of K on the protein content of plant. This finding, however, was not always accepted. As nitrate reduction is the first step in nitrate assimilation (Hageman *et al.*, 1961), a lower activity of nitrate reductase leads to declined assimilation of  $NO_3$ -N.

In this study we have compared metabolism of  $NH_4$ -N and NO,-N in K-, Ca- and Mg-deficit rice plants to make clear roles of K, Ca and My in nitrogen metabolism.

#### MATERIALS AND METHODS

#### Plant culture and feeding of <sup>15</sup>N labeled nitrogen sources

Rice (*Oryza sativa* L., var. Asominori) seeds were sterilized by 70 % ethanol and then 2 % (w/v) calcium hypochlorite, washed with tap water, and germinated on moist filter paper in an environment controlled room (natural light, 25°C, 70 % RH). Then seedlings were transferred to moist sand in plastic pots and supplied with half strength of the nutrient solution (Table 1) for a few days. The seedlings were then transplanted to a plastic plate with holes which was held over a plastic pot containing tap water for a few days in the greenhouse, and then tap water was replaced with half strength of the nutrient solution for several days. The complete nutrient solution of full strength was supplied at 27 days after germination. The nutrient solution was renewed once a week in the first three weeks, then twice a week, and

<u></u>		
Element	Concentration (mg/l)	Compound
N*	28	NH <sub>4</sub> NO <sub>3</sub> NaH <sub>2</sub> PO <sub>4</sub> . H <sub>2</sub> O
P K	31 78	KC1
Са Мо	80 18	CaCl <sub>2</sub> • 2H <sub>2</sub> O MgSO <sub>4</sub> • 7H <sub>2</sub> O
Mg S** Si	24	$ \widetilde{\text{MgSO}}_{4}^{4} \cdot 7\widetilde{\text{H}}_{2}^{2}\widetilde{\text{O}} $ $ \widetilde{\text{Na}}_{2}\widetilde{\text{SiO}}_{3} $
Fe	50 3	Na <sub>2</sub> 31O <sub>3</sub> C <sub>10</sub> H <sub>12</sub> N <sub>2</sub> O <sub>8</sub> NaFe • 3H <sub>2</sub> O MnSO <sub>4</sub> • 4H <sub>2</sub> O
Mn B	0. 2 0. 2	$H_{2}BO_{2}$
Zn Cu	0. 05 0. 05	ZnSO₄.7H₂O CuSO₄.5H₂O
Mo	0. 05	$Na_2MoO_4 \cdot 2H_2O$

**Table 1.** Composition of the complete nutrient solution used.

<sup>\*</sup> N sources were changed to 4mM <sup>15</sup>NH<sub>4</sub>Cl and Na<sup>15</sup>NO<sub>3</sub> at 52-day age of plant for 2 days (during <sup>15</sup>N-feeding).

<sup>\*\*</sup> In Mg-omitted solution Na<sub>2</sub>SO<sub>4</sub> was used as S source.

every 2 days during treatment. The aeration system was used during solution culture. The pH of nutrient solution was adjusted at 5.6 twice a day with 1N  $\rm H_2SO_4$  or NaOH using a pH meter. The nitrogen source was NH4NO3 before the supply of  $^{15}N$ . In the treatment of minus K (-K) and minus Mg (-Mg), K and Mg were omitted from the complete (control) nutrient solution, respectively, and 4 ppm of Ca was added in Ca-limited (-Ca) treatment. Treatment was continued for 10 days. The experiment was done to determine the absorption, translocation and assimilation of NH4-N and NO,-N under different K-, Ca- and Mg-supply conditions. The duplicated treatment was begun at 42 days after sowing, and then 4mM  $^{15}NH_4Cl$  (31.2 atom  $\%\,^{15}N$ ) and Na $^{15}NO_3$  (30.8 atom  $\%\,^{15}N$ ) were fed to 52-day-old plants for 2 days. During  $^{15}N$  feeding 3 plants for each treatment were held in one plastic bottle containing 1 liter of nutrient solution. The plants were harvested at 54 days after sowing, separated to three parts as blade, stem+sheath, and root. Then they were freeze-dried for chemical analysis.

#### Determination of mineral content and total nitrogen

Potassium, Ca and Mg content were determined by combination of a dry combustion method and atomic absorption spectrophotometry. Total nitrogen was determined by the modified Kjeldahl method with salicylic acid and the steam distillation method.

#### Determination of <sup>15</sup>N abundance

The samples were prepared for <sup>15</sup>N analysis by the Kjeldahl digestion and distillation method. <sup>15</sup>N abundance of NH, in concentrated distillates was measured by Dumas method and optical spectrometry with a NIA-I <sup>15</sup>N analyzer (Nihonbunko Co. Ltd, Tokyo, Japan) (Fiedler and Proksch, 1975).

#### Determination of free amino acids and protein-N and its 15N abundance

Free amino acids were extracted by addition of 60 ml of 80 % ethanol to 100 mg of powdered materials and boiling for 60 minutes on a water bath with a reflux condenser. After cooling the extract was filtered through Miracloth in a Guchi crucible. The filtrate was transferred into an evaporation flask to concentrate the sample until dryness under reduced pressure at  $40^{\circ}$ C. The remaining was dissolved by addition of 10 ml of  $H_2O$  and about 10 ml of  $CH_2Cl_2$  with shaking, and kept in a separation funnel for several hours. The upper  $H_2O$  layer was centrifuged (1000 rpm for 12 minutes), and an aliquot of the supernatant was used for determination of free amino acids by an automatic amino acid analyzer (JLC 6 AH), according to the method described by Kedenburg (1971). Nitrogen in the residue on Miracloth was regarded as protein-N. Protein-N and its  $^{15}N$  abundance were determined as described above.

#### Determination of nitrate reductase activity

Rice plants were grown in the nutrient solution including  $2~\mathrm{mM\,NaNO_3}$  before treatment was initiated as described above. At 42-day age of rice the plants were treated with 5 kinds of nutrient solution such as  $2~\mathrm{mM}$ 

NaNO $_3$  (NO $_3$ ), 2 mM NH $_4$ Cl (NH $_4$ ), 2 mM NH $_4$ NO $_3$  (NH $_4$ NO $_3$ ), NH $_4$ NO $_3$  minus K (NH $_4$ NO $_3$ -K) and NH $_4$ NO $_3$  limited Ca (NH $_4$ NO $_3$ -Ca) for 10 days. Thereafter, in a series of NH $_4$ NO $_3$  treatment, the nitrogen source was replaced with 4mM NaNO $_3$  (so abbreviated NO $_3$ , NO $_3$ -K, and NO $_4$ -Ca). The latter treatment was continued for 6 days. The experiments were carried out in 2 replications. Nitrate reductase activity was determined by the  $in\ vivo$  assay method (Hageman and Hucklesby, 1971).

#### **Determination of nitrate**

Nitrate content in dry matter was determined according to the method of Cataldo *et al.* (1975).

#### Statistical analysis

The data of experiments were subjected to statistical analysis according to Duncan's multiple range test, and new LSD test (Duncan, 1955; Waller and Duncan, 1969).

#### RESULTS AND DISCUSSION

#### Effect of K, Ca and Mg dificit on plant growth and mineral content

Treatment of -K, -Ca and -Mg for 12 days had no statistically significant effect on the growth of rice plants (data not shown). It can be seen from Table 2 that K, Ca and Mg concentrations of plant tissues were decreased by -K, -Ca and -Mg treatment, respectively, but the concentration remained at the levels which were required for the maintenance of normal plant growth (Loneragan and Snowball, 1969; Draycott and Durrant, 1971; and Fageria, 1976).

### Effect of K, Ca and Mg deficit on absorption of NH<sub>4</sub>-N and NO<sub>3</sub>-N in rice plants

In order to decrease the effect of pH on the absorption of NH,+ and NO<sub>3</sub>-,

	K(%	<i></i>		Ca(%)			Mg(%)	)
Treatment	Root Stem	+ Blade	Root	Stem+ sheath	Blade	Root	Stem+ sheath	Blade
Ammonium-N								
Complete -K -Ca -Mg	4.85 a 3.25 3.80 b 1.68 4.85 a 2.30 5.10 a 3.60	b 1.60 b a 2.35 a	a 0. 10 a 0. 08 a 0. 09 a 0. 08 a	0.13 a 0.10 a 0.08 b 0.11 a	0.36 a 0.32 a 0.21 b 0.33 a	0. 12 a 0. 13 a 0. 12 a 0. 09 b	0.13 a 0.17 b 0.11 a 0.07 c	0. 15 a 0. 26 b 0. 16 a 0. 07 c
Nitrate-N								
Complete - K -Ca -Mg	5.00 a 3.10 4.25 a 1.55 5.00 a 2.25 4.85 a 3.75	b 1.30 b a 2.55 a	0. 16ac 0. 14 a 0. 10 b 0. 18 c	0. 14 a 0. 12 a 0. 09 b 0. 15 a	0. 36 a 0. 32 a 0. 21 b 0. 36 a	0. 13 a 0. 14 a 0. 13 a 0. 09 a	0. 14 a 0. 18 b 0. 11 a 0. 07 c	0. 14 a 0. 28 b 0. 17 a 0. 08 c

Table 2. Effect of mineral supply on mineral content of rice plants.

Within a column, means followed by the same letter are not significantly different according to Waller and Duncan, using new LSD test for k=100.

the pH of nutrient solution was adjusted at 5.6 twice a day and recorded before the pH was corrected. The deviations from the desired pH value (5.6) were not so great among treatment (data not shown). The data in Table 3 indicate that absorption of NH,-N was not affected by -Ca and -Mg treatment. The absorption of NH,-N in -K treatment was slightly larger than the other treatment, although the difference was statistically insignificant. This difference is probably due to lack of the effect of a monovalent cation (K+) competing with NH<sub>4</sub>+ for selective binding sites in the absorption process. There was a paper that a relative high K+ concentration in nutrient solution resulted in somewhat reduced uptake of NH,+ by rice roots (Fried et al., 1965). The absorption of NO,-N was decreased by 38 % and 30 % by -K and -Ca treatment, respectively, as compared to control treatment (Table 3).

Table 3. Effect of mineral supply on the absorption of  $^{15}{\rm NH_4}$  and  $^{15}{\rm NO_3}$  by rice plants.

T.,	<sup>15</sup> N(mg/g dry weight)						
Treatment	Root	Root Stem-tsheath		Whole plant			
Ammonium-N							
C⊢K∋lete -Ca	4.72 a 6.01 4.59 a a	5.37 a 5.67 3.60 aa	4.22 a 2.496a5 aa	4.79 a 5.47 3.50 aa			
-Mg	4.99 a	4.77 a	3.23 a	4.19 a			
Nitrate-N							
Complete -K -Mg	5954.95aa 5.19 a 4.94 a	2.55 4.25 ba 2.56 b 3.72 a	3.07 1.52 ab 1. 92 b c 2.40 a c	4.14a257b 2.89 b c 3.43 a c			

Within a column, means followed by the same letter are not significantly different at 5% level according to Duncan's multiple range test.

These results are in good agreement with those obtained by Koch and Mengel (1972, 1974) and by Matsumoto *et al.* (1980). The influence of K and Ca was stronger on the absorption of NO,-N than on that of NH,-N. Mg deficit did not affect absorption of  $NO_3$ .

## Effect of K, Ca and Mg deficit on translocation of NH<sub>4</sub>-N and NO<sub>3</sub>-N from root to upper parts of plants

It was supposed that inhibition of absorption of NO,-N in the plant may be caused through preventing the translocation of NO<sub>3</sub>-N from root to upper parts which may occur by the function of K<sup>+</sup> and Ca++. In the data of Table 4 the translocation of  $^{15}N$  supplied as NH<sub>4</sub>-N was not significantly different among treatment, but the translocation of NO,-N was inhibited by -K and -Ca treatment. These results strongly suggest that translocation of NO<sub>3</sub>-N was stimulated by the supply of K and Ca. Similar results have been reported by Frost et al. (1978).

T		<sup>15</sup> N distribution* (%)					
Treatment	Root	Stem+sheath	Blade	Shoot			
Ammonium-N							
Complete - K - Ca - Mg	20.4 a 22.1 a 25.7 a 23.2 a	45.8 a 44.6 a 45.1 a 45.6 a	33. 8 a 33.3 a 29.2 a 31.2 a	79.6 a 77.9 a 74.3 a <b>76.8</b> a			
Nitrate-N							
Complete -K -Ca -Mg	28. 8 a 35. 4 b 38.6 b 27.6 a	42.9 a <b>40.0</b> a b 36.5 b 44.2 a	28. 3 a <b>24.6</b> b 24.9 b 28.2 a	<b>71.2</b> a <b>64.6</b> b 61.4 b 72.4 a			

Table 4. Effect of mineral supply on 15N distribution in rice plant tissues.

Within a column, means followed by the same letter are not significantly different according to Waller and Duncan, using new LSD test for k=100.

### Effect of nitrogen source and mineral supply on nitrate reductase activity of rice plants

Higher activity of nitrate reductase was found in NO, treatment while lower activity was found in NH<sub>4</sub> treatment. In NH<sub>4</sub>NO<sub>3</sub> treatment the activity was slightly decreased as compared to NO, treatment as shown in Fig. 1. The decreases of levels of blade nitrate reductase activity in NH, and NH<sub>4</sub>NO<sub>3</sub> treatment were 86% and 32 % at 10 days of treatment as compared to NO<sub>3</sub> treatment, respectively. This effect may be caused by ammonium inactivation of nitrate reductase (Orebamjo and Stewart, 1975 a, b) or by decreases in NO, content of rice plant (Fig. 2). A positive relationship was frequently observed between NO, content and nitrate reductase activity of plant tissues (Woodson *et al.*, 1984). Nitrate reductase is dependent upon the presence of nitrate in the tissue (Beevers *et al.*, 1965), which may be related to nitrate flux from roots to shoots (Heimer and Filner, 1971) where the reduction takes place (Muhammad and Kumazawa, 1974).

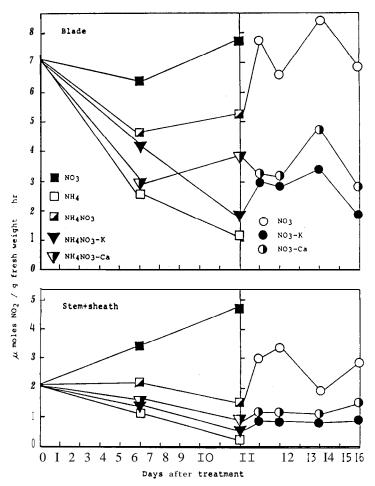
Nitrate reductase activity in response to supply of K and Ca is shown in Fig. 1. The results indicate that nitrate reductase activity in the leaves and stem of rice plants which were supplied with K and Ca was higher than in those of the plants which were deficient in K or Ca. Decreases of nitrate reductase activity in K-deficit leaves were 60, 58, 59 and 71% at 11, 12, 14 and 16 days of treatment, respectively, as compared to control plant. Nitrate reductase activity was also decreased by -Ca treatment. The decreases were 57, 50, 44 and 60 % as compared to control treatment (NO3) at 11, 12, 14 and 16 days of treatment, respectively. These results are in good agreement with those obtained by Paulsen and Harper (1968), Sasakawa and Yamamoto (1977) and Matsumoto *et al.* (1980). As discussed above the results provided evidence for an important physiological role of K and Ca which influence the absorption and translocation of NO,-N. The results presented here also suggest

<sup>\* 15</sup>N distribution in each part was expressed as % of the total amount of 15N absorbed

that K and Ca played a significant role in the nitrate reductase activity through their effect on NO,-N accumulation in the plant leaves, which arises from higher absorption and translocation by the presence of K and Ca. The data in Fig. 2 indicate that NO, content of rice plants was increased by increasing of K and Ca supply. A similar effect was obtained by Blevins et al. (1978).

### Effect of K, Ca and Mg on assimilation of NH,-N and NO<sub>3</sub>-N into amino acids and protein

The concentrations of asparagine, serine, glycine and total amino acids were higher in leaves of rice plants supplied with NH,-N than in those of plants supplied with NO,-N (Table 5). Likewise, protein content and rate of protein synthesis as shown in Tables 6 and 7 were higher in NH,-N fed plant



**Fig. 1.** Effect of nitrogen source and mineral supply on nitrate reductase activity of rice plants.

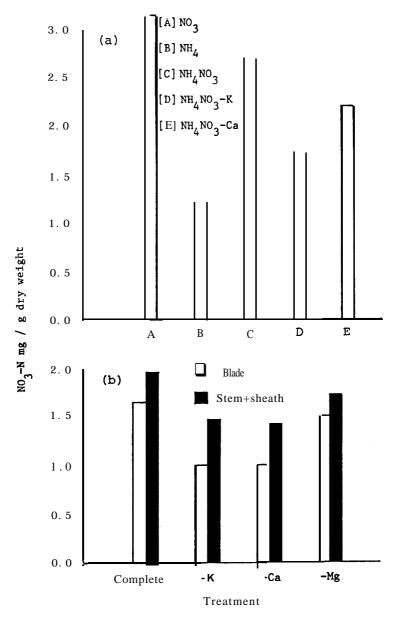


Fig. 2. Effect of nitrogen source and mineral supply on nitrate content of rice plants, (a) nitrate content in blade tissues was determined after 10 days of A, B, C, D and E treatment, (b) nitrate content in blade and stem+sheath tissues was determined after 12 days of complete, -K, -Ca and -Mg treatment (2 days of 18NO<sub>3</sub>-feeding).

Table 5.	Effect of	nitrogen	source	and	mineral	supply	on	the	concentration	
of free a	mino acio	ds of rice	leaves.							

		Ammo	nium-N			Nitra	ite-N	
Free amino acid	Com- plete	-K	-Ca	-Mg	Comple	ete -K	-Ca	-Mg
			μmo	1/ <sub>13</sub> . by	weight			
Aspartic acid Threonine Serine Asparagine Glutamic acid	15. 0 2. 6 7. 6 7. 3 20. 0	10.0 3.6 12.0 18.0 20.0	10.0 1.5 4.3 2.8 12.0	7. 8 3. 6 15: 0	14.0 2.3 5.7 1:1 22.0	9.9 2.0 3.8 2.0 17.0	8.9 1.5 3.5 1:0 16.0	12.0 1.8 3.9 0.7 19.0
Glutamine Proline Glycine Alanine Valine Isoleucine Leucine Tyrosine Phenylalanine	2.3 0.9 0.5 7.6 0.8 0.5 0.4 0.2 0.3	1.3 2.2 10.0 1.1 0.5 0.4 0.3 0.3	0.8 0.4 7.3 0.9 0.5 0.4 0.2 0.5	0.7.9 0.67.5 1.0 0.7 0.5 0.4 0.6	0.7 1.0 8.1 0.9 0.5 0.5 0.4 0.5	0.6 1.0 8.3 0.9 0.5 0.4 0.3 0.5	1. 0 0. 6 8. 7 0. 6 0. 4 0. 3 0. 3	0. 7 0. 5 8. 9 0. 7 0. 5 0. 4 0. 3 0. 5
Lysine Histidine Arginine	0. 3 0. 4 0. 1 0. 3	0. 8 0. 3 2. 8	0.3 0.2 0.3	0. 4 0. 2 0. 6	0. 3 0. 4 0. 2 0. 3	0. 5 0. 2 0. 4	0.3 0.1 0.2	0. 3 0. 2 0. 3
Total	66.8	89.0	44.3	57. 1	58.6	48. 3	43.9	50. 7
Ammonia	7.7	19.0	5. 4	4.9	9.0	8.7	5.6	5.5

Table 6. Effect of mineral supply on protein-15N content of rice plants.

T	Protein 15N (mg/g dry weight)						
Treatment	Root	Stem+sheath	Blade	Whole plant			
Ammonium-N							
C-K plete -Ca	2.37 a 3.61 4.01 a a	2.77 a b 3.171.80ab	2.80 a 3.742.08 a a	2.70 a 3.47 a			
-Mg	2.46 a	2.26 a b	2.66 a	2. 32 a 2.46 a			
Nitrate-Complete							
-K	1.49 2.15 a a	0.92 1.66 a b	2.18 1.07 b a	1. 95 a 1.09 b			
-Ca -Mg	1.76 a 1.86 a	0.72 b 1.61 a	1.37 b c 1.71 a c	1.19 b c 1.70 a c			

Within a column, means followed by the same letter are not significantly different at 5% level according to Duncan's multiple range test.

than in  $NO_3$ -N fed plant. Usually, the plants which have received NH,-N are clearly different in their nitrogenous composition from those which have received NO,-N (Clark, 1936; Takaki *et al.*, 1968) because  $NH_4$ + is directly incorporated into amino acids and then protein, but  $NO_3$ — must be reduced to  $NH_3$  before its incorporation to amino acids and protein. The present data show that the synthesis of amino acids from inorganic nitrogen may be more active in plants supplied with NH,-N than in those supplied with  $NO_3$ -N. There

<b>Table 7.</b> Effect of mineral sup	oly on t	the rate of	protein-15N to	total 15N
in each tissue of rice plants.				

_	Protein <sup>15</sup> N*(%)						
Treatment	Root	Stem+sheath	Blade	Whole plant			
Ammonium-N							
Complete - K -Ca -Mg	53. 3 a 59.3 a 88.6 b 49. 3 a	51.5 a 56.2 a 49.8 a 47.4 a	75. 4 b 72. 9 b 82: 6 c	56.5 a 63.5 b 66.9 b 58.7 a			
Nitrate-N							
Complete -K	36.2 a	39.8 a	70.8 a	47.0 a			
-Ca -Mg	30.2 34.6 a a 38.6 a	36.2b 27.9c 43.2d	70.0 71.1 a a <b>71. 2 a</b>	42.2 41.1 bb <b>49. 5 a</b>			

<sup>\*</sup> Protein <sup>15</sup>N was expressed as % of the total amount of <sup>15</sup>N in each part. Within a column, means followed by the same letter are not significantly different according to Waller and Duncan, using new LSD test for k=100.

is evidence that ammonium could accumulate only in very small amounts, and its change to amides or amino acids Was very quick, but nitrate <code>could</code> accumulate in large amounts because ammonium ion (NH,+) or its dissociation product (NH,) is toxic but nitrate ion is not toxic. Ammonium ion is "detoxified" by union with amino acids. In contrast, nitrate can be absorbed and stored in various tissues without damage to the plant (Hageman, 1980).

Total free amino acid concentrations of rice plants which were supplied with NH4-N were higher in -K treatment than those in the other treatment (Table 5). The increases in the amino acid concentration of -K treatment were 28 % as compared to control treatment. The stimulation of both amino acid and protein synthesis (Tables 5-7) in K-deficit plant supplied with NH<sub>4</sub>-N may be attributed to higher absorption of NH<sub>4</sub>-N, because excess ammonium must be in union with amino acid for detoxification of NH.+. Treatment of -Mg had no effect on the synthesis of amino acids and protein from both the nitrogen sources. It thus seems that in Mg-deficit plant the Mg concentration required for enzyme activation may be still adequate in plant tissues (Table 2). In NO,-N fed plant free amino acid and protein synthesis were decreased by -K and -Ca treatment (Tables 5-7). Total amino acids slightly decreased in K- and Ca-deficit plant supplied with NO<sub>3</sub>-N (Table 5). The inhibition of protein synthesis in K- and Ca-deficit plant was 58.7 % and 47.1% as compared to control plant, respectively (Table 6). Lower absorption of NO, and nitrate reductase activity are considered to be responsible for limited assimilation of NO,-N in K- and Ca-deficit plant. (Table 3 and Fig. 1).

In rice plant, conclusively, moderate K deficiency promotes the utilization of NH,-N through the enhanced absorption of NH,-N but moderate Ca and Mg deficiency had no effect on the utilization of NH,-N. Both K and Ca have important roles in the absorption, translocation and reduction of NO,-N in rice plant. Moderate Mg deficiency does not affect the utilization of NO,-N.

#### REFERENCES

- Bange, G. G. J., J. Tromp and S. Henkes 1965 Interactions in the absorption of potassium, sodium, and ammonium ions in excised barley roots. *Acta Bot. Neerl.*, 14: 116-130
- Beevers, L., L. E. Schrader, D. Flesher and R. H. Hageman 1965 The role of light and nitrate in the induction of nitrate reductase in radish cotyledons and maize seedlings. *Plant Physiol.*, 40: 691-698
- Ben-Zioni, A., Y. Vaadia and S. H. Lips 1970 Correlations between nitrate reduction, protein synthesis and malate accumulation. *Physiol. Plant.*, 23: 1039-1047
- Ben-Zioni, A., Y. Vaadia and S. H. Lips 1971 Nitrate uptake by roots as regulated by nitrate reduction products of the shoot. *Physiol. Plant.*, 24: 288-290
- Besford, R. T. 1975 Effect of potassium nutrition on leaf protein concentration and growth of young tomato plants. *Plant Soil*, 42: 441-451
- Blevins, D. G., N. M. Barnett and W. B. Frost 1978 Role of potassium and malate in nitrate uptake and translocation by wheat seedlings. *Plant Physiol.*, *62:* 784-788
- Blevins, D. G., A. J. Hiatt and R. H. Lowe 1974 The influence of nitrate and chloride uptake on expressed sap pH, organic acid synthesis and potassium accumulation in higher plants. *Plant Physiol.*, 54: 82-87
- Burstrom, H. 1954 Studies on growth and metabolism of roots. X. Investigation of the calcium effects. *Physiol. Plant.*, 7: 332-343
- Cataldo, D. A., M. Haroon L. E. Schrader and V. L. Youngs 1975 Rapid colorimetric determination of nitrate in plant tissue by nitration of salicylic acid. Soil Sci. Plant Anal., 6: 71-80
- Clark, H. E. 1936 The effect of ammonium and nitrate nitrogen on the composition of the tomato plant. *Plant Physiol.*, 11: 5-24
- Cox, W. J. and H. M. Reisenauer 1973 Growth and ion uptake by wheat supplied nitrogen as nitrate, or ammonium, or both. *Plant Soil*, 38: 363-383
- Dijkshoorn, W. and M. Ismunadji 1972 Nitrogen nutrition of rice plants measured by growth and nutrient content in pot experiments. 2- Uptake of ammonium and nitrate from a water-logged soil. **Neth.** J. Agrc. Sci., 20: 44-57
- Draycott, A. P. and M. J. Durrant 1971 Plant and soil magnesium in relation to response of sugar beet to magnesium application. *J. Int. Inst.* Sugar *Beet Res.*, *5:* 129-135
- Duncan, D. B. 1955 Multiple range and multiple F tests. Biometrics, 11: 1-42
- Fageria, N. K. 1976 Influence of potassium concentration on growth and potassium uptake by rice plants. *Plant Soil, 44: 567-573*
- Fiedler, R. and G. Proksch 1975 The determination of nitrogen-15 by emission and mass spectrometry in biochemical analysis, a review. *Anal. Chim. Acta*, 78: 1-62
- Fried, M., F. Zsoldes, P. B. Vose and I. L. Shatokhin 1965 Characterizing the NO, and NH, uptake process of rice roots by use of <sup>15</sup>Nlabelled NH<sub>4</sub>NO<sub>3</sub>. **Physiol. Plant.**, 18: 313-320
- Frost, W. B., D. G. Blevins and N. M. Barnett 1978 Cation pretreatment effects on nitrate uptake, xylem exudate, and malate levels in wheat seedlings. *Plant Physiol.*, *61:* 323-326
- Haeder, H. E. and K. Mengel 1969 The absorption of potassium and sodium in dependence on the nitrogen nutrition level of the plant. Landw. Forsch., 23: 53-60
- Hageman, R. H. 1980 Effect of form of nitrogen on plant growth. In "Nitrification Inhibitors-Potentials and Limitations" ed. by J. J. Meisinger, J. J. Randall and M. L. Vitosh, Amer. Soc. Agron., Madison, pp. 47-62.
- Hageman, R. H., D. Flesher and A. Gitter 1961 Diurnal variation and other light effects influencing the activity of nitrate reductase and nitrogen metabolism in corn.

- Crop Sci., 1: 201-204
- Hageman, R. H. and D. P. Hucklesby 1971 Nitrate reductase from higher plants. In "Methods in Enzymology" Vol 23, ed. by A. San Pietro. Academic Press Inc., New York, pp. 491-503.
- Heimer, Y. M. and P. Filner 1971 Regulation of nitrate assimilation pathway in cultured tobacco cells. III. The nitrate uptake system. Biochim. Biophys. Acta, 230: 362-372
- Hsiao, T. C., R. H. Hageman and E. H. Tyner 1970 Effects of potassium nutrition on protein and total free amino acids in Zea mays L., Crop Sci., 10: 78-82
- Kedenburg, C. P. 1971 A lithium buffer system for accelerated single-column amino acid analysis in physiological fluids. Anal. Biochem., 40: 35-42
- Koch, K. and K. Mengel 1972 Effect of varied potassium nutrition on the uptake and incorporation of labelled nitrate by young tobacco plants (Nicotiana tabacum L.). J.Sci. Food Agr., 23: 1107-1112
- Koch, K. and K. Mengel 1974 The influence of the level of potassium supply to young tobacco plants (*Nicotiana tabacum* L.) on short-term uptake and utilization of nitrate nitrogen (15N). *J. Sci. Food Agr.*, 25: 465-471
- Loneragan, J. F. and K. Snowball 1969 Calcium requirements of plants. Aust. J. Agr. Res., 20: 465-478
- Matsumoto, H., K. Teraoka and T. Kawasaki 1980 Repression of nitrate reductase in cucumber leaves caused by calcium deficiency. *Plant Cell Physiol.*, *21: 183-191*
- Minotti, P. L., D. C. Williams and W. A. Jackson 1968 Nitrate uptake and reduction as affected by calcium and potassium. Soil *Sci. Soc. Amer. Proc.*, 32: 692-698
- Muhammad, S. and K. Kumazawa 1974 Assimilation and transport of nitrogen in rice. II. <sup>15</sup>N-labelled nitrate nitrogen. *Plant Cell Physiol.*, 15: 759-766
- Oji, Y. and G. Izawa 1970 Studies on the absorption and assimilation of inorganic nitrogen in intact plants (part 2). Physiological characteristics in absorption and assimilation of nitrate-N and ammonium-N in young rice seedling. *J. Sci. Soil Manure Japan, 41: 31-36* (in Japanese).
- Orebamjo, T. 0. and G. R. Stewart 1975a Ammonium repression of nitrate reductase formation in *Lemnu minor* L.. *Planta*, 122: 27-36
- Orebamjo, T. O. and G. R. Stewart 1975 b Ammonium inactivation of nitrate reductase in Lemnu *minor* L.. *Planta*, 122: 37-44.
- Paulsen, G. H. and J. E. Harper 1968 Evidence for a role of calcium in nitrate assimilation in wheat seedlings. *Plant Physiol.*, 43: 775-780
- Rosen, C. J. and R. M. Carison 1984 Characterization of K+ and NH,+ absorption by myrobalan plum and tomato: Influence of plant potassium status and solution concentrations of K+ and NH,+. J. Amer. Soc. Hort. Sci., 109: 552-559
- Sasakawa, H. and Y. Yamamoto 1977 Nitrate assimilation in rice seedlings as affected by mineral nutrition. *Plant Cell Physiol.*, 18: 215-224
- Schrader, L. E. and R. H. Hageman 1967 Regulation of nitrate reductase activity in corn (Zeu mays L.) seedlings by endogenous metabolites. *Plant Physiol.*, 42: 1750-1756
- Takaki, H., M. Ikeda, Y. Yamada and T. Harada 1968 Occurrence of glucosamine in higher plants. Soil Sci. Plant Nutr., 14: 56-61
- Waller, R. A. and D. B. Duncan 1969 A Bayes rule for the symmetric multiple comparisons problem. *J. Amer. Statist. Ass.*, 64: 1484-1503
- Webster, G. C. 1961 Protein synthesis. Ann. Rev. Plant Physiol., 12: 113-132
- Woodson, W. R., F. B. Negm and J. W. Boodley 1984 Relationship between nitrate reductase, nitrogen accumulation, and nitrogen partitioning in chrysanthemum. *J. Amer. Soc. Hort.* Sci., 109: 491-494