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<https://doi.org/10.5109/23667>

出版情報：九州大学大学院農学研究院紀要. 22 (4), pp.211-219, 1978-07. Kyushu University
バージョン：
権利関係：

Effect of Plant Hormones on Tobacco Mosaic Virus Concentration in Tobacco Tissue Culture

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(Received February 3, 1978)

Tobacco mosaic virus (TMV) concentrations in tobacco callus tissues grown on the media containing α -naphthaleneacetic acid (NAA) and kinetin at different concentrations were assayed periodically during successive cultures. Tobacco callus originally raised from the TMV-infected tobacco stem became green and compact when serially cultured on the medium containing lower concentration (0.01-0.1 mg/l) of NAA or higher concentration (2 mg/l) of kinetin. Translucent and soft callus was obtained at higher concentration (1-40 mg/l) of NAA or lower concentration (0-0.2 mg/l) of kinetin. Microscopic observations of the thin sectioned callus tissues revealed that the green colored compact tissue was composed of the closely associated small cells and many tracheid-like structures, while in the translucent and soft tissue, large cells were roughly arranged showing less association and no tracheid-like structure was found. TMV was maintained at high concentration in the compact callus during successive cultures, however, it was no longer detected beyond the third generation (90 days) in the translucent soft callus grown under the optimum hormone balances. Since both hormones do not have direct inactivation effect on TMV, the results suggest that TMV multiplication in the callus tissue closely corresponds with the cell arrangement of the tissue, which is considerably affected by hormone balances in the media.

INTRODUCTION

Previous investigation provided evidence that tobacco mosaic virus (TMV) concentrations in the growing tobacco callus tissue were different depending on the type of the tissue (Omura, 1978). Various calli different in appearance were known to be obtained by changing concentrations or balances of plant hormones in the medium. Kinetin promotes cell division of tobacco callus tissue (Miller *et al.*, 1955). Tobacco callus becomes compact if kinetin concentration is high in the medium, while it becomes soft or friable at lower concentrations (Fossard *et al.*, 1974; Linsmaier and Skoog, 1965; Nishiyama and Taira, 1966; Vasil and Hildebrandt, 1967). Generally, α -naphthaleneacetic acid (NAA) promotes cell enlargement in tobacco callus tissue. On the influence of these plant hormones upon virus infection and/or multiplication, many works have been carried out by using intact systemic and local lesion hosts (Aldwinckle, 1975; Aldwinckle and Selman, 1967; Cheo, 1969; Daft, 1965; Gondo, 1953; Hariharasubramanian, 1968; Kiraly *et al.*, 1968; Kiraly and Szirmai, 1964; Milo and Srivastava, 1969a; Nakagaki, 1971; Selman, 1964; Simons

et al., 1972). But these results will not be applicable in the case of callus tissue. By using TMV-infected tobacco callus tissue Kutsky and Rawlins (1950) and Kutsky (1952) reported that both NAA and indolebutyric acid markedly decreased TMV concentration. Milo and Srivastava (1969) reported that TMV multiplication in tobacco pith tissue culture was strongly inhibited by cytokinin at higher concentrations (20-200 $\mu\text{g/l}$) but increased at lower concentrations (0-2 $\mu\text{g/l}$).

The present experiment was undertaken to make clear the effect of NAA and kinetin on TMV multiplication in the growing tobacco tissue cultures with special reference to the growth habit, type and histological observation of the tissue.

MATERIALS AND METHODS

Culture media

Murashige and Skoog (1962) medium (free from Edamin, added with 20 g/l of sucrose and 6 g/l of agar) (basal medium) supplied with 1 mg/l of NAA and 0.2 mg/l of kinetin (stock medium) was used for stock culture. In order to test the effect of plant hormones, the media containing NAA and kinetin in different concentrations were prepared and pH was adjusted to 5.7-5.8 with either 1 N NaOH or 1 N HCl. Thirty ml of the medium was allotted to each 100 ml Erlenmeyer's flask, and was autoclaved at 120°C for 10 min.

Tobacco tissue culture

Tobacco tissue cultures raised from TMV (ordinary strain)-infected tobacco stem (cv. Bright Yellow) have been transferred monthly on the stock medium for 70 generations in our laboratory. The dark green colored compact callus maintaining TMV at high concentration was used for the first transfer in each experiment. The pieces about 100 mg each from translucent soft tissue of a callus were monthly transferred on the media containing NAA and kinetin at different concentrations and were incubated under continuous 3,000 lux white fluorescent light at ca. 25°C. For growth studies, 18 calli were cultured in each experiment, and fresh weight of 10 calli selected at random were recorded at the end of each generation.

Infectivity assay

Nicotiana glutinosa was used for bioassay of TMV concentration as reported in the previous paper (Omura, 1978). The plants grown in 12 cm pots in greenhouse for 60-80 days were moved to a greenhouse air-conditioned at 25°C, a week before inoculation, and the tip and lower leaves were cut off leaving four fully expanded leaves three days before inoculation. Small piece from the peripheral tissue of each callus was weighed and homogenized with mortar and pestle and was appropriately diluted with distilled water. Five hundred-fold (W/V) diluted suspension was commonly inoculated. Inoculation was made by cotton swabs on half leaves of *N. glutinosa* dusted with 400 mesh carborandum. Purified TMV, the concentrations of which were ex-

pected to produce 50-150 local lesions on a half-leaf, was simultaneously inoculated on the opposite half-leaves as a control. Eight leaves were used for each specimen. Local lesions were counted three days after inoculation. All the experiments were repeated three times.

RESULTS

Effect of plant hormones on the growth and appearance of tobacco tissue culture

The tissue pieces obtained aseptically from the peripheral part of a stock callus were transferred on the test media containing NAA 0.01-40 mg/l and kinetin 0-2 mg/l in different combinations. Table 1 shows the color and average fresh weight of ten calli grown on each medium during each generation (30 days). The tissues grew well on the media containing 1 and 10 mg/l of NAA and the best growth was resulted with the combination of NAA 1-10 mg/l and kinetin 0.2 mg/l. Most part of the callus proliferated into green or brown and became compact at 2 mg/l of kinetin irrespective of NAA concentrations or at relatively low concentrations of NAA irrespective of kinetin concentrations. The inner tissue of these calli was rather translucent. The green and compact peripheral tissue was able to separate easily from the soft translucent inner tissue when calli were grown at the combination of 1 mg/l of NAA and 0.02 or 0.2 mg/l of kinetin. At relatively high concentrations of NAA with relatively low concentrations of kinetin, the callus having translucent peripheral tissue with greenish inner tissue grew. Translucent tissue gradually became whitish in later stage of 30 days incubation period. In general, the green or brown tissue was compact and the translucent tissue was soft and friable.

Table 1. Growth and color of tobacco tissue cultures on the media containing different combinations of plant hormones¹⁾.

NAA (mg/l)	Kinetin (mg/l)				
	0	0.002	0.02	0.2	2
0.01	0.2 ²⁾ G ³⁾	0.3 T, G ⁴⁾	0.3 B	0.2 G	0.4 G, B
0.1	0.5 G, T	0.7 G, T	0.7 T, G	0.6 G	0.8 G
1	1.5 G, T	G, T	T, G	4.2 T, G	3.2 G
10	2.2 G, T	1.6 B, T	2.5 G, T	6.3 G, T	1.0 G
40	0.6 B, T	0.4 B, T	0.4 G, T	0.9 G, T	0.7 G

¹⁾ Basal medium: Murashige and Skoog (1962) medium (free from Edamin, added with 20 g/l of sucrose and 6 g/l of agar). ²⁾ Mean fresh weight of callus at the end of the first generation (generation: 30 days). ³⁾ Capital letters show the characteristics of callus. B; brown and compact, G; green and compact, T ; translucent and soft. ⁴⁾ Of two capitals written side by side, the left indicates the characteristics of inner tissue and the right indicates that of peripheral tissue.

From above results, two series of the combination of NAA and kinetin concentrations, i. e., 0.01-40 mg/l NAA added with 0.2 mg/l kinetin, and 0-2 mg/l kinetin added with 1 mg/l NAA were employed. These series were considered to be sufficient to elucidate the effect of hormones on the nature of the tissue and TMV multiplication.

Effect of NAA on the growth of callus and TMV concentration

The fresh weight of each callus grown on the medium containing different concentrations of NAA is given in Table 2. The growth patterns were much different depending on the concentrations of NAA. At 0.01 and 0.1 mg/l of NAA, the growth of callus during the first generation (30 days) reduced to about 1/10 and 1/5 respectively of that of stock callus and low level of growth lasted till the 6th generation. At 1, 10 and 40 mg/l of NAA, however, the callus was quickly adapted to the medium and good growth was resulted after the second generation. The calli grown on the medium containing 0.01 or 0.1 mg/l NAA were bright green colored and compact and they sometimes produced leaf primordia. At 10 and 40 mg/l of NAA, translucent soft tissue developed from the peripheral part of calli and inner part remain-

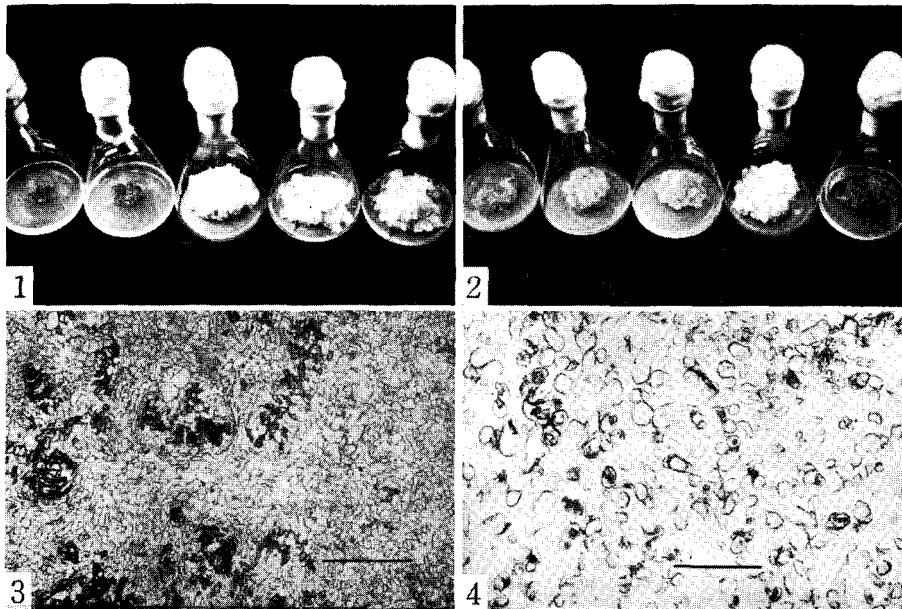


Fig. 1. Calli at the end of the 6th generation on each concentration of NAA added with 0.2mg/l of kinetin. From the left, 0.01, 0.1, 1, 10 and 40 mg/l of NAA.

Fig. 2. Calli at the end of the 6th generation on each concentration of kinetin added with 1mg/ of NAA. From the left, 0, 0.002, 0.02, 0.2 and 2 mg/l of kinetin.

Fig. 3. Section of peripheral tissue of stock callus. Scale indicates 500 μ .

Fig. 4. Section of a callus grown for 6 generations on the media containing 40mg/l of NAA and 0.2 mg/l of kinetin. Scale indicates 500 μ .

Table 2. Effect of NAA on the growth of tobacco tissue culture¹⁾.

NAA conc. (mg/l)	Generation ²⁾					
	1	2	3	4	5	6
0.01	0.43 ³⁾	0.4	0.4	0.5	0.6	0.8
	0.8	0.9	1.0		1.1	1.5
0.1	3.9	6.6	8.7	9.9	9.7	9.7
10	6.5	8.9	9.5	9.3	8.9	9.4
40	0.9	9.8	9.1	8.9	9.4	8.9

¹⁾ Kinetin concentration in the medium was 0.2 mg/l.

²⁾ About 100 mg of translucent tissue was transferred every 30 days.

³⁾ Mean fresh weight (g) of callus.

Table 3. Effect of NAA on TMV concentration in tobacco tissue culture¹⁾.

NAA conc. (mg/l)	Exp. no.	Generation ²⁾			
		0	2	4	6
0.01	1		1053 ³⁾	120	77
	2		106	102	143
	3		47	88	42
0.1	1		125	117	70
	2		104	64	21
	3		81	70	16
1	1	100	140	14	1
	2	100	73	13	2
	3	100		4	0
10	1		2	5	3
	2		4	1	1
	3		0	3	0
40	1		5	2	1
	2		0	1	1
	3		1	0	0

¹⁾ See Table 2. ²⁾ See Table 2. ³⁾ Relative TMV concentration at the end of each generation as compared to that in stock callus.

ed greenish and relatively compact. Fig. 1 shows the calli grown on the media containing different concentrations of NAA at the end of the 6th generation.

TMV concentrations in the calli were assayed every two transfers. As shown in Table 3, TMV activity diminished markedly during successive generations correlating with NAA concentrations, *i. e.*, it was maintained at high level for long period at 0.01 and 0.1 mg/l of NAA, but it fell rapidly at 1, 10 and 40 mg/l of NAA. Although TMV concentration fell most rapidly at 10 and 40 mg/l of NAA, it was maintained in detectable level in some callus tissues even at the end of 6 th generation.

Effect of kinetin on the growth of callus and **TMV** concentration

Growth of the callus on the medium containing kinetin at different con-

centrations of 0.2 mg/l with 1 mg/l of NAA is given in Table 4. During the first generation on the medium free from kinetin, the growth reduced to about 1/3 of that of stock callus. The best growth of the callus was observed in every generations at 0.2 mg/l of kinetin. The type of calli was markedly affected by concentration of kinetin. The calli grown at 0 and 0.002 mg/l of kinetin were composed of translucent and soft peripheral layer with greenish and relatively compact inner tissue at the end of the first generation and it became completely translucent and friable at the end of the 6 th generation (Fig. 2). The calli grown at 0.02 and 0.2 mg/l of kinetin were composed of translucent and soft inner tissue with green and compact peripheral tissue and whole body became translucent and soft when inner tissue was used for every transfer. At 2 mg/l of kinetin, the callus was green and compact in appearance till the 6 th generation even though the softest tissue was used for every transfer.

Table 4. Effect of kinetin on the growth of tobacco tissue culture¹⁾.

Kinetin conc. (mg/l)	Generation ²⁾					
	1	2	3	4	5	6
0	1.7 ³⁾	2.1	2.6	4.4	5.0	5.2
0.002	3.2	3.5	4.6	5.2	5.6	4.8
0.02	3.6	4.4	5.4	5.9	6.0	5.8
0.2	4.2	7.0	8.3	9.7	9.6	9.7
2	3.3	3.8	4.4	5.1	5.4	5.5

¹⁾ NAA concentration in the medium was 1 mg/l. ²⁾³⁾ See Table 2.

Table 5. Effect of kinetin on TMV concentration in tobacco tissue culture¹⁾.

Kinetin conc. (mg/l)	Exp. no.	Generation ²⁾			
		0	2	4	6
0	1		0 ³⁾	0	0
	2		8	0	0
	3		10	0	0
0.002	a		29	0	0
			0	0	0
	3		4	0	0
0.02	1		25	1	0
	2		13	0	0
	3		14	2	0
0.2	1	100	75		1
	2	100	103	10	0
				20	0
2	1		149	190	187
	2		120	145	104
	3		143	88	88

¹⁾ See Table 4. ²⁾ See Table 2. ³⁾ See Table 3.

TMV concentration in callus tissue was assayed every two transfers. Table 5 shows the trends of TMV concentration in the tissues cultured on the media containing different concentrations of kinetin (0-2 mg/l) added with 1 mg/l of NAA. TMV concentration decreased rapidly at lower concentrations of kinetin. TMV concentration in the tissue grown on the medium containing kinetin less than 0.002 mg/l fell markedly with increase of generations and no longer was detected beyond the 4 th generation. At 2 mg/l of kinetin, however, no decreasing effect was observed.

Histological observation of the callus

Frozen sections of the calli grown at different hormonal conditions were made at the end of the 6 th generation to compare with those of stock callus. Most area of the peripheral tissue of stock callus consisted of compactly arranged small cells, and many tracheid-like structures were observed (Fig. 3). The callus tissues grown on 0.01 and 0.1 mg/l of NAA added with 0.2 mg/l of kinetin were composed of the cells much different in size, associating tightly with each other in some areas. In most cases, the callus grown at 40 mg/l of NAA and 0.2 mg/l of kinetin was composed of loosely arranged large cells (Fig. 4) as in the case of the tissues grown at 10 mg/l of NAA with 0.2 mg/l of kinetin.

The callus cultured for 6 generations at 0, 0.002, 0.02 or 0.2 mg/l kinetin added with 1 mg/l of NAA was composed of cells larger and less associated than those in the callus grown at 40 mg/l of NAA and 0.2 mg/l of kinetin (Fig. 4). On the other hand, the callus grown for 6 generations at 2 mg/l of kinetin with 1 mg/l of NAA was composed of the cells varied in size tightly associating in most areas, which was similar to those grown at 0.01 and 0.1 mg/l of NAA added with 0.2 mg/l of kinetin.

Effect of NAA on the growth of callus and TMV concentration under the kinetin-free conditions

Since the maintenance of TMV activity in tobacco callus was highly correlated with kinetin concentration in the medium (Table 5), growth of the callus and change of TMV concentration were examined on purpose with kinetin free medium.

At 0.01 mg/l of NAA, tissue became brown and did not grow beyond the 3 rd transfer. At 0.1 mg/l of NAA, watery soft tissue developed at first, but grew poor after that, and finally they became brown at the end of each generation. At 1, 10 and 40 mg/l of NAA, tissue proliferated rapidly to make friable soft callus after the 2 nd or 3rd transfer. TMV concentration fell sharply during the first and 2 nd generation and no longer be detected beyond the 3 rd or 4 th generation in the friable tissues.

DISCUSSION

Tobacco callus tissue infected with TMV was reported to be consisted of a mixture of the infected and healthy cells (Hansen and Hildebrandt, 1966).

According to the previous work carried out by fluorescent antibody method, TMV was unevenly distributed in the infected callus corresponding with compactness of the tissue or distribution frequency of tracheid-like structures (Omura, 1978). TMV concentration in these tissues usually ran parallel with the frequency of the infected cells (Omura, 1978). Furthermore, it was also elucidated that TMV concentration was maintained for many generations by successive transfers of green compact tissues, while it decreased rapidly if translucent soft tissues were successively transferred (Omura, 1978). The medium containing either lower level of kinetin or higher level of NAA made callus tissue translucent and soft, while opposite hormonal conditions made it green and compact (Table 1), coinciding with the results obtained by previous workers (Fossard *et al.*, 1974; Linsmaier and Skoog, 1965; Nishiyama and Taira, 1966; Vasil and Hildebrandt, 1967). TMV concentration was decreased rapidly in the callus grown under the former condition, whereas it was maintained for many years under the latter condition (Table 3, 5). The most rapid decrease was observed in the callus grown on kinetin-free medium (Table 5).

The reasons why TMV concentration decreases so rapidly in the translucent soft tissue grown on the media containing hormones at optimum concentrations are yet obscure. Milo and Srivastava (1969b) reported that TMV multiplication in tobacco callus was strongly inhibited at 0.2 mg/l of kinetin. In our experiment, however, TMV concentration was maintained at a constant level for over 80 generations in stock callus on the medium containing 0.2 mg/l of kinetin and 1 mg/l of NAA so far as green compact tissue was used for every transfer. Furthermore, TMV concentration in the originally transplanted pith tissue did not decrease at all at the end of the first generation. These results suggest that direct inactivation effect of plant hormones on TMV will be negligible.

As reported previously, TMV is unevenly distributed in tobacco callus tissue, correlating with compactness of cell arrangement (Omura, 1978). The effect of plant hormones on TMV concentration in callus tissue seems not to be caused by direct manner but by shifting the compactness of cell arrangement in the tissue.

For the purpose of eliminating virus from virus-infected plant tissue through callus tissue, the successive transfers of translucent tissue on the medium containing higher concentration (10, 40 mg/l) of NAA with lower concentration (0.02 mg/l) of kinetin should be recommended.

ACKNOWLEDGMENTS

The authors are grateful to Dr. Z. Hidaka, for his encouragement during this work.

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