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Dietary egg-white protein increases body protein mass and reduces body fat mass through an acceleration of hepatic β -oxidation in rats

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Abstract

Egg-white protein (EWP) is known to reduce lymphatic TAG transport in rats. In this study, we investigated the effects of dietary EWP on body fat mass. Male rats, 4 weeks old, were fed diets containing either 20% EWP or casein for 28 d. Carcass protein levels and gastrocnemius leg muscle weights in the EWP group were significantly higher than those in the casein group. In addition, carcass TAG levels and abdominal fat weights in the EWP group were significantly lower than those in the casein group; adipocyte size in abdominal fat in the EWP group was smaller than that in the casein group. To identify the involvement of dietary fat levels in the rats, one of two fat levels (5 or 10%) was added to their diet along with the different protein sources (EWP and casein). Abdominal fat weight and serum and hepatic TAG levels were significantly lower in the EWP group than in the casein group. Moreover, significantly higher values of enzymatic activity related to β -oxidation in the liver were observed in the EWP group compared with the casein group. Finally, abdominal fat weight reduction in the EWP group with the 10% fat diet was lower than that in the EWP group with the 5% fat diet. In conclusion, our results indicate that, in addition to the inhibition of dietary TAG absorption reported previously, dietary EWP reduces body fat mass in rats through an increase of body protein mass and the acceleration of β -oxidation in the liver.

Key words: Egg-white protein: Adipocyte size: Body composition: β -Oxidation: Rats

Proteins serve numerous nutritional functions in the body, contributing to essential structural components and enzymes, as well as providing the base materials for hormones and assisting in immunity⁽¹⁾. Importantly, protein deficiency has been reported to be one of the causes of sarcopenia⁽²⁾. Because there is a negative correlation between the number of sarcopenia patients and the amount of protein ingested in the diet⁽³⁾, it is important to consume foods high in protein to maintain and increase muscle. However, the ingestion of large amounts of protein for middle-aged and elderly individuals can be difficult. Furthermore, because proteins are used in different ways throughout the body depending on their source⁽⁴⁾, it is important to ingest protein that can contribute appropriately to muscle health.

Egg whites are a fat-free, high-protein food. Egg-white protein (EWP) contains many essential amino acids, having an amino acid score of 100, and is a high-quality source of protein compared with proteins derived from milk and soybeans⁽⁵⁾. EWP also has a higher net protein utilisation than these proteins and other proteins with an amino acid score of 100⁽⁶⁾. In this

study, we focused on EWP as one such source. Recently, because of changes in lifestyle habits, an increase in abdominal fat has become prevalent and has been found to lead to dyslipidaemia, hypertension and impaired glucose tolerance, eventually escalating to the metabolic syndrome, which can cause atherosclerotic disease⁽⁷⁾. Approximately 50% of adult Japanese men have the metabolic syndrome or are at an increased risk for this condition⁽⁸⁾. Aside from water, the human body primarily consists of proteins and lipids⁽⁹⁾. Therefore, it is hypothesised that increasing body protein can lower body fat proportion and abdominal fat levels and prevent or reduce the incidence of the metabolic syndrome.

Most studies reporting a decrease in abdominal fat because of the consumption of functional foods have identified physiologically active substances contributing to this reduction, such as catechins⁽¹⁰⁾ and polyphenols⁽¹¹⁾, and it has also been reported that medium-chain fatty acids have an abdominal-fat-lowering effect⁽¹²⁾. Importantly, a relationship between protein ingestion and visceral fat reduction is indicated by consumption

Abbreviation: EWP, egg-white protein.

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of lactoferrin, a protein fraction contained in milk, which has been shown to decrease visceral fat⁽¹³⁾. In addition, the soyabean protein fraction, β -conglycinin, has been reported to lower serum TAG concentrations⁽¹⁴⁾. Therefore, protein consumption can potentially lead to a decrease in visceral fat. However, in the nutritional protein research field, the effect of EWP on abdominal fat levels has not been investigated.

Here, we investigated the effect of EWP on body protein and body fat. In addition, we assessed the effect of EWP on abdominal fat accumulation and sought to identify the suppressive mechanism of EWP on abdominal fat accumulation.

Methods

Materials

EWP was obtained from the Kewpie Egg Corporation. Casein was purchased from Oriental Yeast Co. Ltd. The protein content was determined by the Dumas method (Association of Official Analytical Chemists official method 968.06) and calculated as the nitrogen content $\times 6.25$ ⁽¹⁵⁾. The measured protein content was 85.9% for the casein group and 86.1% for the EWP group. The amino acid composition of EWP and casein was analysed with an automatic amino acid analyser JLC-500/V (JEOL Ltd); results are shown in Table 1.

Animals and diets

Male Sprague–Dawley rats (Slc:SD), 4 weeks old, were obtained from Japan SLC Inc. The rats were housed individually in an air-conditioned room (22–24°C) at 50% humidity on a 12 h light–12 h dark cycle (light from 08.00–20.00 hours). Before the experiment, all rats were permitted free access to AIN-76 diets⁽¹⁶⁾ for 3 d. Experimental diets were also prepared according to the AIN-76 formulation. The compositions of experimental diets are shown in Table 2. Body weight and food intake of rats in Expts 1–3 were recorded every day.

For Expt 1, the rats were fed either the casein (*n* 10) or EWP diet (*n* 10) and provided feed containing 10% maize oil

(Table 2) for 28 d using a pair-feeding protocol. At the end of the final study day, the rats were subjected to 8-h fasting, anaesthetised with isoflurane (Intervet K.K.) and then killed via cervical dislocation and their carcasses were analysed.

For Expt 2, the rats were fed the casein (*n* 10) or EWP diet (*n* 9) and provided feed containing 10% maize oil (Table 2) for 28 d using a pair-feeding protocol. At the end of the final study day, the rats were subjected to 8-h fasting, anaesthetised with isoflurane and then killed by abdominal aortic exsanguination. The gastrocnemius and soleus leg muscles, and white adipose tissues (epididymal, mesenteric, perirenal + retroperitoneal and subcutaneous), were excised and weighed.

For Expt 3, rats were fed the casein or EWP diet and provided feed containing 5 or 10% maize oil (Table 2) for 28 d using a pair-feeding protocol (casein, maize oil 5%: *n* 6; EWP, maize oil 5%: *n* 6; casein, maize oil 10%: *n* 6; EWP, maize oil 10%: *n* 6). At the end of the final study day, the rats were subjected to 8-h fasting, anaesthetised with isoflurane and then killed by abdominal aortic exsanguination. The liver and white adipose tissues (epididymal, mesenteric, perirenal + retroperitoneal and subcutaneous) were excised and weighed.

The livers and white adipose tissues were kept at –20°C until they were analysed. Blood was kept at room temperature for 30 min and then centrifuged at 1600 *g* for 10 min to collect the serum. The serum was kept at –20°C until it was analysed. Samples for liver enzyme activity were prepared by homogenising the liver, centrifuging the homogenate at 500 *g* for 10 min, collecting the supernatant (a part of the supernatant such as the liver homogenate without nuclear fraction was stored at –80°C until it was analysed) and centrifuging the supernatant at 100 000 *g* for 1 h. The supernatant (the liver cytosolic fraction) from the last step was collected and stored at –80°C until it was analysed⁽¹⁷⁾.

All experiments were conducted under the guidelines for Animal experiments at Kewpie Corporation and Law no. 105

Table 1. Amino acid composition of casein and egg-white protein (EWP)

	Casein (%)	EWP (%)
Asp	6.54	9.91
Thr	4.00	4.43
Ser	5.08	6.39
Glu	18.8	12.1
Pro	9.93	3.39
Gly	1.69	3.39
Ala	4.96	7.04
Val	6.30	6.78
Cys	0.48	2.74
Met	2.78	3.91
Ile	4.96	5.22
Leu	8.84	8.21
Tyr	5.45	3.65
Phe	4.72	5.87
Lys	7.99	7.17
His	2.66	2.35
Trp	1.33	1.56
Arg	3.51	5.87
Amino acid score (%)	100	100

Table 2. Composition of experimental diets*

	Maize oil 5% (g/kg)		Maize oil 10% (g/kg)	
	Casein	EWP	Casein	EWP
Casein	200	–	200	–
Dried EWP	–	200	–	200
Maize starch	150	150	150	150
Cellulose	50	50	50	50
Maize oil	50	50	100	100
Mineral mixture (AIN-76)	35	35	35	35
Vitamin mixture (AIN-76)	10	10	10	10
DL-Met	3	3	3	3
Choline bitartrate	2	2	2	2
Sucrose	500	500	450	450
Energy (kJ/100 g diet)	1565	1565	1677	1677
Moisture (g/100 g diet)	4.45	3.75	4.41	3.71
Protein (g/100 g diet)	17.7	17.7	17.7	17.7
Carbohydrate (g/100 g diet)	65.4	65.4	60.4	60.5
Fat (g/100 g diet)	5.41	5.19	10.4	10.2
Ash (g/100 g diet)	1.99	2.81	1.99	2.81
Fibre (g/100 g diet)	5.00	5.00	5.00	5.00

EWP, egg-white protein.

* For Expts 1 and 2, the rats were fed the casein or EWP diet and provided feed containing 10% maize oil for 28 d. For Expt 3, the rats were fed the casein or EWP diet and provided feed containing 5 or 10% maize oil for 28 d.

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Analysis of morphometric and metabolic parameters

Serum TAG, phospholipids, NEFA and glucose levels were measured using commercial enzyme assay kits from Wako Pure Chemicals (TAG E-test; Phospholipid C-test, NEFA C-test and GlucoseCII-test). Serum insulin and leptin concentrations were determined using commercial ELISA kits (Morinaga Insulin ELISA Kit or Morinaga Leptin ELISA Kit; Morinaga Institute of Biological Science, Inc.). Creatinine was analysed with a Creatinine Assay Kit (Biovision Co.) and ketone bodies with a β -hydroxybutyric acid (ketone body) Assay Kit (Cayman Chemical Co.).

Total lipids in the carcass (Expt 1) and the liver (Expt 3) were extracted using the method described by Folch *et al.*⁽¹⁸⁾. Carcass protein content was analysed using the Dumas method⁽¹⁵⁾. The TAG and phospholipid content in the carcass and liver were determined using the methods described by Fletcher⁽¹⁹⁾ and Wootton⁽²⁰⁾, respectively.

Adipocyte size was measured as described elsewhere^(21,22). In brief, white adipose tissue was rinsed with a saline solution, fixed in a 10% neutral formalin-buffered saline solution and embedded in paraffin. The tissue was cut into 10- μ m sections and stained with haematoxylin to measure the cell size (100 cells/rat). The National Institutes of Health *ImageJ* software was used to measure cell size⁽²³⁾. The DNA content of perirenal+retroperitoneal adipose tissue was extracted using proteinase K followed by phenol/chloroform. The quantity of the DNA obtained was assessed by UV spectrophotometry at 260 nm, as described elsewhere.

The enzymatic activities of carnitine palmitoyltransferase and acyl-CoA oxidase in the liver homogenates without the nuclear fraction were determined using the methods described by Ide *et al.*⁽¹⁷⁾. The enzymatic activities of fatty acid synthase, malic enzyme and glucose-6-phosphate dehydrogenase in the liver cytosolic fraction were determined using the methods described by Ide *et al.*⁽¹⁷⁾.

Statistical analysis

We expected a Δ body fat (%) about 5 (%) smaller in the EWP group than in the casein group based on the previous study by Ochiai & Matsuo⁽²⁴⁾. A SD of 1.5 of the higher value could be allowed. For a type 1 risk α of 0.05 and power (1- β) of 80%, the sample size required was more than three per group.

All results are shown as the means with their standard errors. Comparisons between two groups were performed using Student's *t* test (Table 3 of Expt 1, Table 4 and Fig. 1 of Expt 2). The effects of dietary protein source, dietary fat content and their interaction were analysed by two-way ANOVA (Table 5 of Expt 3). If an interaction was observed, it was analysed further by Tukey-Kramer multiple comparison *post hoc* test (Table 5 of Expt 3). Differences were considered significant at $P < 0.05$. Statistical analysis was performed using SPSS version 20 (IBM Japan Ltd).

Results

Effects of dietary egg-white protein on body fat and protein mass (Expt 1)

To assess the effect of EWP on body composition in rats, we analysed rat carcasses fed the casein or EWP diet. No significant differences in body weight increase, food intake and energy intake between the EWP and casein groups were observed (Table 3). The levels of protein and TAG were measured in the rat carcasses. The protein levels in the carcasses of the EWP group were significantly higher than that of the casein group (Table 3). In addition, the TAG levels in the EWP group were significantly lower than that of the casein group (Table 3). Finally, the sum of body protein and fat levels in carcasses was 28.1 (SE 0.5) and 28.1 (SE 0.7) g in the casein and EWP groups, respectively, which was not significantly different.

Effect of dietary egg-white protein on adipocyte size (Expt 2)

On the basis of the results of Table 3, we evaluated the effects of EWP on abdominal fat mass and adipocyte size in rats. No significant differences in body weight increase, food intake and energy intake between the EWP and casein groups were observed (Table 4). Perirenal+retroperitoneal and subcutaneous adipose tissue weights were significantly lower in the EWP group compared with the casein group. No significant differences in the epididymal and mesenteric adipose tissue weights between the EWP and casein groups were observed (Table 4).

The average adipocyte size in perirenal+retroperitoneal adipose tissue in the EWP group was smaller than that in the casein group. In addition, the number of small adipocytes was increased and the number of large adipocytes was decreased in the EWP group compared with the casein group (Table 4). The DNA content, used as an index of the number of cells in the perirenal+retroperitoneal adipose tissue, was significantly higher in the EWP group than in the casein group (Table 4). However, the liver TAG concentration was significantly lower in the EWP group than in the casein group (Table 4).

Table 3. Growth parameters and carcass contents of rats fed the casein or egg-white protein (EWP) diet (Expt 1)
(Mean values with their standard errors; *n* 10/group)

	Casein		EWP	
	Mean	SEM	Mean	SEM
Growth parameter (g/d)				
Body weight gain	7.24	0.09	7.29	0.18
Food intake	17.2	0.1	16.7	0.4
Energy intake (kJ/d)	288	2	280	5
Carcass contents (g/100 g body weight)				
Protein	18.6	0.2	20.9	0.5*
TAG	9.57	0.72	7.18	0.63*
Phospholipids	0.683	0.045	0.641	0.026

Mean values were significantly different from those of the casein group: * $P < 0.05$.

Table 4. Growth and morphometric parameters, adipocyte size and DNA contents in adipose tissue of rats fed the casein or egg-white protein (EWP) diet (Expt 2) (Mean values with their standard errors; *n* 9–10/group)

	Casein		EWP	
	Mean	SEM	Mean	SEM
Body weight gain (g/d)	6.74	0.25	6.98	0.16
Food intake (g/d)	17.8	0.2	17.7	0.2
Energy intake (kJ/d)	298	6	298	3
Gastrocnemius leg muscle (g/100 g body weight)	1.08	0.02	1.15*	0.02
Soleus leg muscle (g/100 g body weight)	0.0663	0.0027	0.0644	0.0033
Fat mass (g/100 g body weight)				
Epididymal	1.55	0.10	1.38	0.08
Mesenteric	0.944	0.044	0.856	0.053
Perirenal + retroperitoneal	2.38	0.14	2.02*	0.12
Total (abdominal fat)	4.87	0.24	4.25*	0.23
Subcutaneous	2.59	0.22	2.06*	0.20
Adipocyte size (μm^2)				
Perirenal + retroperitoneal	4634	395	3209*	213
DNA ($\mu\text{g/g}$ tissue)				
Perirenal + retroperitoneal	238	21	332*	18

*Mean value was significantly different from that of the casein group ($P < 0.05$).

Effect of dietary egg-white protein on lipid metabolism in the liver (Expt 3)

To identify the involvement of dietary fat levels in the rats, one of two fat levels (5 or 10%) was added to their diet along with the different protein sources (EWP and casein). No significant difference in food intake and energy intake among the four groups was observed. The increase in body weight was significantly higher in rats given 10% maize oil than rats given 5% maize oil. Mesentery adipose tissue weight trended to being lower in the EWP group compared with the casein group ($P = 0.06$), and in the 5% maize oil group compared with the 10% maize oil group ($P = 0.06$). The perirenal adipose tissue weight was significantly lower in the EWP group compared with the casein group, whereas no significant difference between the 5 and 10% maize oil groups was observed. The testicular adipose tissue weight trended to being lower in the EWP group compared with the casein group ($P = 0.06$), and these values for the 5% maize oil group were significantly lower than in the 10% maize oil group. The total white adipose tissue weight was significantly lower for the EWP group compared with the casein group, and the 5% maize oil group trended to having lower values than the 10% maize oil group (Table 5).

Serum TAG and NEFA concentrations were significantly lower following EWP consumption than after casein consumption. Serum insulin concentration was significantly higher after EWP consumption than after casein consumption, whereas significantly lower values were observed in the 5% maize oil group than in the 10% maize oil group. In addition, significantly lower leptin values were noted after EWP consumption than after casein consumption, whereas significantly higher values were observed in the 5% maize oil group than in the 10% maize oil group. Serum creatinine and β -hydroxybutyric acid concentrations were significantly lower after EWP consumption than after casein consumption, whereas the creatinine concentration was significantly lower in the 10% maize oil group

than in the 5% maize oil group. No significant differences were observed for the serum phospholipids or glucose concentrations among the four groups (Table 5).

No significant differences in liver phospholipid concentration among the four groups were observed (Table 5). The liver TAG concentration was significantly lower in rats that were on the EWP diet compared with rats on the casein diet. In addition, significantly higher values for carnitine palmitoyltransferase and acyl-CoA oxidase activity in the liver were observed after EWP consumption than after casein consumption, and significantly lower malic enzyme activity was observed after EWP consumption than after casein consumption. Finally, with a 10% maize oil diet, glucose-6-phosphate dehydrogenase activity was significantly lower after EWP consumption than after casein consumption. However, no effect of EWP ingestion on fatty acid synthase activity was observed (Table 5).

Discussion

The results of this study indicate that the consumption of an EWP-based diet increases body protein by 12% and gastrocnemius leg muscle weight by 6%, while decreasing body TAG levels by 25%, abdominal fat by 12–18%, subcutaneous fat by 20% and adipocyte size by 31% when compared with a casein-based diet (Tables 3–5, Fig. 1). The increase in the level of body protein following EWP consumption appears to be due to effects from its fractions, the amino acids that compose the proteins and the EWP component as a whole. The effect of EWP is likely to be because of its net protein utilisation of approximately 95%, which is much higher than that of casein, which is 70%^(6,25). It has been reported previously that ovalbumin and ovotransferrin from EWP fractions promote the growth of cultured muscle cells (C2C12 cells)⁽²⁶⁾. In addition, the effects of the amino acids within EWP are likely because of the presence of branched amino acids in egg whites, which increase the branched amino acid concentrations in the blood when ingested⁽²⁷⁾. After exercise, the concentrations of these amino



Table 5. Serum and hepatic lipid, and hepatic enzyme activity in rats fed casein and egg-white protein (EWP) on changing dietary fat level (5 or 10%) (Expt 3) (Mean values with their standard errors; *n* 6/group)

	Maize oil 5%				Maize oil 10%				Two-way ANOVA (<i>P</i>)		
	Casein		EWP		Casein		EWP		Protein	Oil	Interaction
	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM			
Growth parameters (g/d)											
Body weight gain	6.15	0.17	6.28	0.23	6.73	0.22	6.78	0.38	NS	NS	NS
Food intake	17.6	0.0	17.4	0.4	17.2	0.3	16.7	0.6	NS	NS	NS
Energy intake (kJ/d)	275	1	273	6	289	5	280	10	NS	NS	NS
Fat mass (g/100 g body weight)											
Epididymal	1.42	0.07	1.27	0.13	1.80	0.16	1.44	0.12	0.0570	<0.05	NS
Mesenteric	1.02	0.04	0.904	0.036	1.15	0.08	1.02	0.08	0.0641	0.0641	NS
Perirenal + retroperitoneal	0.748	0.099	0.652	0.089	0.880	0.071	0.654	0.036	0.0504	NS	NS
Total (abdominal fat)	3.19	0.19	2.82	0.22	3.82	0.28	3.12	0.21	<0.05	0.0521	NS
Serum parameters											
TAG (mg/100 ml)	135	12	79.8	18.1	101	14	50.9	4.9	<0.05	NS	NS
Phospholipids (mg/100 ml)	158	2	159	13	149	11	138	14	NS	NS	NS
NEFA (mEq/l)	0.738	0.021	0.584	0.066	0.758	0.068	0.519	0.062	<0.05	NS	NS
Glucose (mg/100 ml)	92.8	5.4	104	6	104	5	108	7	NS	NS	NS
Insulin (ng/ml)	0.787	0.107	1.26	0.18	0.286	0.138	0.532	0.075	<0.05	<0.05	NS
Leptin (ng/ml)	3.24	0.33	2.03	0.48	3.58	0.46	1.68	0.28	<0.05	<0.05	NS
Serum parameters											
Creatinine (nmol/ml)	9.94	0.42	6.93	0.29	6.66	0.52	5.32	0.27	<0.05	<0.05	NS
β-Hydroxybutyric acid (mg/ml)	0.459	0.031	0.373	0.039	0.451	0.022	0.361	0.027	<0.05	NS	NS
Hepatic lipids (mg/g liver)											
TAG	49.5	4.9	28.0	1.9	38.6	3.5	26.5	2.6	<0.05	NS	NS
Phospholipids	30.6	0.6	29.3	1.0	29.5	0.6	30.5	0.7	NS	NS	NS
Hepatic enzyme activity (nmol/min per mg protein)											
CPT	2.89	0.51	4.79	0.52	2.89	0.34	3.95	0.26	<0.05	NS	NS
ACO	2.35	0.35	3.95	0.33	3.32	0.49	4.62	0.33	<0.05	NS	NS
FAS	7.21	0.72	7.19	0.41	7.49	0.50	7.70	0.32	NS	NS	NS
G6PDH	54.8	4.8 ^a	31.4	3.3 ^b	27.1	3.3 ^b	26.4	2.7 ^b	<0.05	<0.05	<0.05
ME	30.9	3.1	24.3	1.3	24.0	1.4	14.8	2.2	<0.05	<0.05	NS

CPT, carnitine palmitoyltransferase; ACO, acyl-CoA oxidase; FAS, fatty acid synthase; G6PDH, glucose-6-phosphate dehydrogenase; ME, malic enzyme.

^{a,b} Mean values within a row with unlike superscript letters were significantly different (*P* < 0.05) by Tukey–Kramer multiple comparison *post hoc* test.

Egg-white protein improves body composition

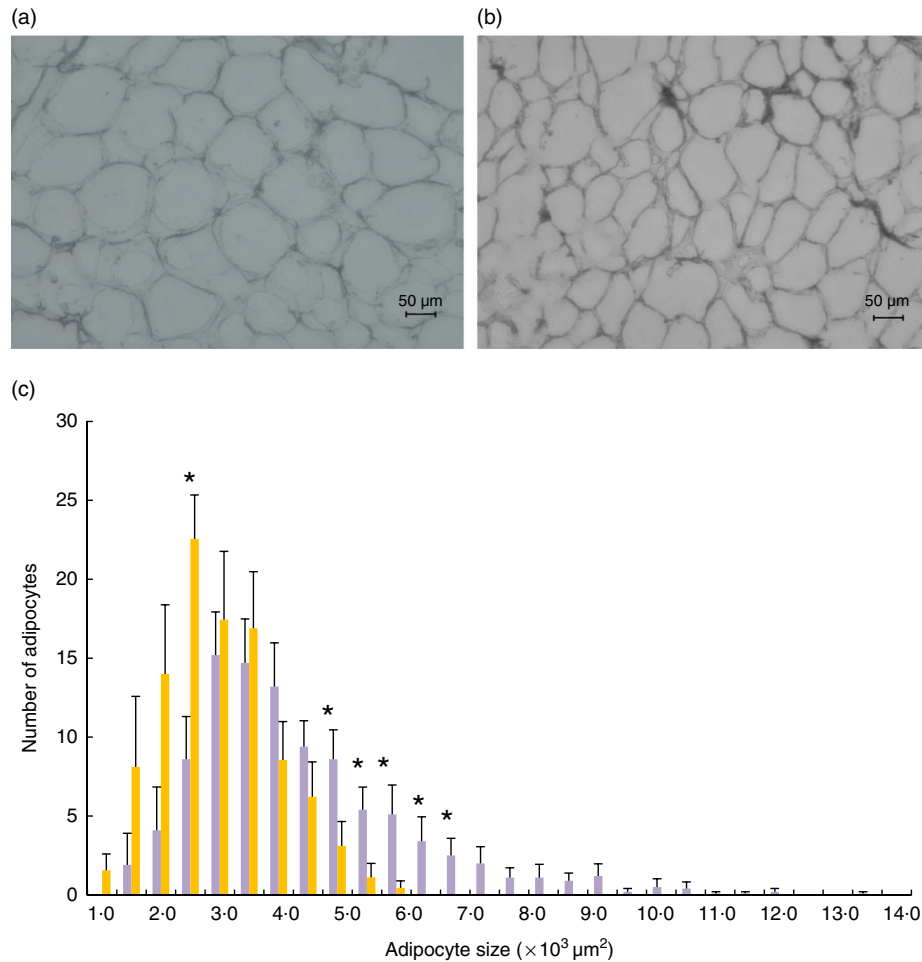


Fig. 1. Effects of dietary egg-white protein (EWP) on cell size in white adipose tissue. Adipocytes are shown in paraffin sections of perirenal+retroperitoneal adipose tissue (scale bar: 50 μm). Casein group (a), EWP group (b). Profile of the distribution of cell size for adipocytes from perirenal+retroperitoneal adipose tissue (c). Values are means (n 9–10 rats/group), with their standard errors. c: \square , Casein; \blacksquare , EWP. Student's t test between the casein and EWP groups: * $P < 0.05$.

acids decrease⁽²⁷⁾, and it is possible that after exercise the branched amino acids in EWP are used for muscle repair and hypertrophy. Consistent with this, the combination of strength training and EWP ingestion has been found to increase muscle mass and strength⁽²⁸⁾, and egg whites are thus considered to be an effective source of protein for body building.

An increase in body protein in rat carcasses following EWP consumption with a high-fat diet has also been reported by Ochiai & Matsuo⁽²⁴⁾. They also noted that EWP ingestion decreased body fat, although the effect was not as clear. In their study, the food intake amount was significantly lower in rats fed EWP compared with rats fed casein. Because body fat is affected by the overall amount of food and lipids consumed, the decrease in body fat expected in that study would be because of the reduction in energy and lipid ingestion, and thus could not be attributed to EWP consumption. In our experiments, we used a pair-feeding strategy to match the food intake amount. Therefore, as equivalent food intake was accounted for, it was possible to determine that EWP consumption was responsible for increasing body protein and decreasing body fat. In addition, the combined weight of body protein and fat were roughly

equivalent for the casein and EWP groups in this study, lending further support that EWP can promote a gain in body protein and reduction in body fat.

Previously, it was reported that daily ingestion of 15 g of EWP by male subjects performing a mild resistance exercise led to an increase in their muscle mass⁽²⁸⁾. Although muscles typically do not increase in size unless they are exercised, muscle mass increased in our study for an unknown reason. For example, the consumption of EWP led to an increase in gastrocnemius muscle mass, when compared with consumption of casein. It is possible that the protein increase provided by the diets in this study, particularly the EWP diet, led to a decrease in abdominal fat because protein contributes heavily to the increase in muscle mass, and muscles are the site of fat combustion. However, serum creatinine levels, which are related to muscle energy metabolism, were not elevated in this study even though this would be expected if fat combustion in the muscles was increased⁽²⁹⁾. In contrast, the levels of β -hydroxybutyric acid, a ketone body, were significantly decreased in the rats fed EWP compared with the rats fed casein (Table 5). Serum ketone bodies levels reflect the balance between the rate of hepatic

ketogenesis and the rate of ketone body utilisation in extra-hepatic tissues and organs, such as skeletal muscle, heart and brain⁽³⁰⁾. Therefore, the increase of protein in skeletal muscle could have led to increased utilisation of ketone bodies followed by a decrease in serum β -hydroxybutyric acid levels. In addition, our findings indicate that there is an increase in the activity of enzymes related to β -oxidation in the liver and a decrease in liver TAG levels following consumption of an EWP-based diet (Table 5). Therefore, the decrease in liver TAG might cause an increase in the activity of liver β -oxidation enzymes after the ingestion of EWP. In the future, it will be necessary to examine whether liver β -oxidation is regulated in this manner, using gene analysis or other methods.

Another possible mechanism through which EWP may have exerted its abdominal-fat-lowering effect is the suppression of lipid absorption into abdominal fat cells. We previously reported that EWP can suppress lipid absorption and lymphatic lipid transport 20% more than casein does⁽³¹⁾, which is important because food ingredients effective at suppressing TAG absorption, such as indigestible dextrin and catechin, have been shown to reduce abdominal fat^(32–36). The mechanism underlying the difference in inhibition is likely that the water-holding capacity, settling volume and viscosity are higher in the degradation products of EWP pepsin hydrolysate than in those of casein pepsin hydrolysate⁽³¹⁾. However, there are likely other mechanisms involved, because the suppression of TAG absorption was higher than the suppression of cholesterol absorption.

An additional possible mechanism that could have contributed to the abdominal-fat-lowering effect we observed surrounds the EWP ovalbumin. It is known that ovalbumin binds to fatty acids⁽³⁷⁾ and that unheated ovalbumin is minimally degraded by pepsin⁽³⁸⁾, and therefore it can reach the small intestine largely intact. Moreover, EWP inhibit lipase activity⁽³⁹⁾ and a small amount of NEFA produced by the action of lipase suppress lipid absorption by binding to ovalbumin. In this study, we used unheated egg white, but ovalbumin in heated egg whites is known to be more susceptible to degradation by pepsin⁽³⁸⁾. For this reason, confirmation of the abdominal-fat-lowering effect we observed here using heated egg whites would help to clarify that the observed effect was due to an inhibition of lipid absorption.

We also observed an abdominal fat mass reduction of approximately 10–20% (Tables 2 and 3). Therefore, abdominal fat mass appears to be highly influenced by the inhibition of lipid absorption. However, the suppression of lipid absorption is insufficient to explain the reduction in the size of abdominal adipocytes we observed, because the ingestion of EWP caused a decrease in the area of abdominal adipocytes by approximately 30% in comparison with casein (Fig. 1). Therefore, the reduction in the size of abdominal adipocytes following the ingestion of EWP is likely to be due to the suppression of lipid absorption, an increase of body protein mass and enhancement of liver β -oxidation. The reduction of abdominal adipocytes may also occur because of cellular events caused by direct action of amino acids and/or peptides. For example, thiazolidinedione derivatives have been reported to reduce the size of abdominal adipocytes by inducing expression of the *PPAR γ* gene⁽⁴⁰⁾. This would be an interesting avenue for future investigation that we did not perform here, because we could

not perform an assay of gene expression within abdominal fat in this study.

The effective dose of EWP in humans is unknown. It has been reported that daily ingestion of 5 g of β -conglycinin leads to a reduction of abdominal fat⁽¹⁴⁾. Given that EWP (i.e. ovalbumin, ovotransferrin and lysozyme) reduce abdominal fat⁽³¹⁾, daily ingestion of approximately 8 g of EWP, equivalent to a total of 5 g of these three fractions, should successfully reduce abdominal fat. This may not be possible in practice, because it is known that adverse effects, including the development of allergies and biotin deficiency, can occur. However, we believe that avidin likely undergoes degradation by pepsin, because the avidin band disappears in SDS-PAGE after a reaction with pepsin⁽³⁸⁾, and that there is unlikely to be a problem unless biotin is ingested in conjunction with egg white. Moreover, low-allergen egg white free of ovomucoid is available, so this type of egg white might exert the abdominal-fat-lowering effect at a lower amount than regular egg white.

Increase of abdominal fat is a risk factor for arteriosclerotic diseases, because it induces symptoms such as dyslipidaemia, glucose intolerance and hypertension⁽⁷⁾. To gain the benefits of EWP, it may be preferable to ingest only egg whites in processed foods, because it allows one to ingest only egg white as opposed to the whole egg. However, products using egg whites are readily available in the market, as egg white is utilised in most foods manufactured today. Ultimately, the successful clarification of an abdominal-fat-lowering effect of EWP will lead to the successful utilisation of EWP as a nutraceutical that could prevent the metabolic syndrome.

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References

1. Kido T, Shizuka F, Shimomura Y, *et al.* (2013) Dietary reference intake for Japanese 2010: protein. *J Nutr Sci Vitaminol* **59**, 836–843.
2. Rosenberg IH (1997) Sarcopenia: origins and clinical relevance. *J Nutr* **127**, 990S–991S.
3. Houston DK, Nicklas BJ, Harris TB, *et al.* (2008) Dietary protein intake is associated with lean mass change in order,

- community-dwelling adults: the health, aging, and body composition (Health ABC) study. *Am J Clin Nutr* **87**, 150–155.
4. Lin CP & Huang PC (1986) Comparison of control diets containing various protein levels for determining net protein utilization by rats. *J Nutr* **116**, 216–222.
5. World Health Organization (1985) *Energy and Protein Requirements. Report of a Joint FAO/WHO/UNU Expert Consultation. WHO Technical Report Series*, no. 724. Geneva: WHO.
6. Sheffner AL, Eckfeldt GA & Spector H (1956) The pepsin-digest-residue (PDR) amino acid index of net protein utilization. *J Nutr* **60**, 105–120.
7. Alberti KG & Zimmet PZ (1998) Definition, diagnosis and classification of diabetes mellitus and its complications, part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. *Diabet Med* **15**, 539–553.
8. Kato Y, Hamasaki T, Sato S, *et al.* (2014) Relationship between the attitudes towards improving eating habits and prevalence of metabolic syndrome. Data from the 2005 National Health and Nutrition Survey and the comprehensive survey of living conditions. *Nihon Kosbu Eisei Zasshi* **61**, 385–395.
9. Frisch RE, Hegsted DM & Yoshinaga K (1977) Carcass components at first estrus of rats on high-fat and low-fat diets: body water, protein, and fat. *Proc Natl Acad Sci U S A* **74**, 379–383.
10. Nagao T, Hase T & Tokimitsu I (2007) A green tea extract high in catechins reduces body fat and cardiovascular risks in humans. *Obesity (Silver Spring)* **15**, 1473–1483.
11. Akazome Y, Kametani N, Kanda T, *et al.* (2010) Evaluation of safety of excessive intake and efficacy of long-term intake of beverages containing apple polyphenols. *J Oleo Sci* **59**, 321–338.
12. Takeuchi H, Sekine S, Kojima K, *et al.* (2008) The application of medium-chain fatty acids: edible oil with suppressing effect on body fat accumulation. *Asia Pac J Clin Nutr* **17**, 320–323.
13. Ono T, Murakoshi M, Suzuki N, *et al.* (2010) Potent anti-obesity effect of enteric-coated lactoferrin: decrease in visceral fat accumulation in Japanese men and women with abdominal obesity after 8-week administration of enteric coated lactoferrin tablets. *Br J Nutr* **104**, 1688–1695.
14. Kohno M, Hirotsuka M, Kito M, *et al.* (2006) Decreases in serum triglycerol and visceral fat mediated by dietary soybean beta-conglycinin. *J Atheroscler Thromb* **13**, 247–255.
15. Sticher JE, Jolliff CR & Hill RM (1969) Comparison of dumas and kjeldahl methods for determination of nitrogen in faces. *Clin Chem* **15**, 248–254.
16. American Institute of Nutrition (1977) Report of the American Institute of Nutrition *ad hoc* committee on standards for nutritional studies. *J Nutr* **107**, 1340–1348.
17. Ide T, Murata M & Sugano M (1996) Stimulation of the activities of hepatic fatty acid oxidation enzyme by dietary fat rich in α -linolenic acid in rat. *J Lipid Res* **37**, 448–463.
18. Folch J, Lee M & Stanley GHS (1957) A simple method for the isolation and purification of total lipids for animal tissue. *J Biol Chem* **226**, 497–509.
19. Fletcher MJ (1968) A calorimetric method for estimating serum triglycerides. *Clin Chim Acta* **22**, 393–397.
20. Wootton DG (1977) Semiautomated spectrophotometry of total phospholipids in plasma. *Clin Chem* **23**, 1766–1768.
21. Sato M, Uzu K, Yoshida T, *et al.* (2008) Effects of milk fermented by *Lactobacillus gasseri* SBT2055 on adipocyte size in rats. *Br J Nutr* **99**, 1013–1017.
22. Caiou B, Harmelen KV, Duran-Sandoval D, *et al.* (2006) The farnesoid X receptor modulates adipose and peripheral insulin sensitivity in mice. *J Biol Chem* **281**, 11039–11049.
23. Tong LT, Katakura Y, Kawamura S, *et al.* (2010) Effects of Kurozu concentrated liquid on adipocyte size in rats. *Lipids Health Dis* **9**, 134.
24. Ochiai M & Matsuo T (2014) Effect of egg white and its hydrolysate on stearoyl-CoA desaturase index and fat accumulation in rat tissue. *Int J Food Sci Nutr* **65**, 948–952.
25. Tagle MA & Donoso G (1965) Net protein utilization determined in short- and long-term experiments with rats. *J Nutr* **87**, 173–178.
26. Mizunoya W, Tashima A, Sato Y, *et al.* (2015) The growth-promoting activity of egg white proteins in the C2C12 myoblast cell line. *Anim Sci J* **86**, 194–199.
27. Kato Y, Numao S, Miyauchi R, *et al.* (2010) Effect of intermittent blood volume fluctuation of light resistance exercise after ingestion of the high protein snacks on plasma branched chain amino acid concentrations in young adults. *J Nutr Sci Vitaminol* **56**, 255–259.
28. Kato Y, Sawada A, Numao S, *et al.* (2011) Chronic effects of light resistance exercise after ingestion of a high-protein snack on increase of skeletal muscle mass and strength in young adults. *J Nutr Sci Vitaminol* **57**, 233–238.
29. Patel SS, Molnar MZ, Tayek JA, *et al.* (2013) Serum creatinine as a marker of muscle mass in chronic kidney disease: results of a cross-sectional study and review of literature, L. *J Cachexia Sarcopenia Muscle* **4**, 19–29.
30. Fabbrini E & Magkos F (2015) Hepatic steatosis as a marker of metabolic dysfunction. *Nutrients* **7**, 4995–5019.
31. Matsuoka R, Shirouchi B, Kawamura S, *et al.* (2014) Dietary egg white protein inhibits lymphatic lipid transport in thoracic lymph duct-cannulated rats. *J Agric Food Chem* **62**, 10694–10700.
32. Ikeda I, Tsuda Y, Suzuki Y, *et al.* (2005) Tea catechins with a galloyl moiety suppress postprandial hypertriglycerolemia by delaying lymphatic transport of dietary fat in rats. *J Nutr* **135**, 155–159.
33. Unno T, Tago M, Suzuki Y, *et al.* (2005) Effect of tea catechins on postprandial plasma lipid responses in human subjects. *Br J Nutr* **93**, 543–547.
34. Kajimoto O, Kajimoto Y, Yabune M, *et al.* (2005) Tea catechins with a galloyl moiety reduce body weight and fat. *J Health Sci* **51**, 161–171.
35. Kametani N, Asakura R, Mitsuda H, *et al.* (2009) Effects of mixing tea beverage containing resistant maltodextrin on postprandial serum triglyceride and safety evaluation on long-term or excessive intake of the beverage. *Jpn J Food Chem Safety* **16**, 20–27.
36. Kishimoto Y, Wakabayashi S & Tokunaga K (2000) Effects of long-term administration of indigestible dextrin on visceral fat accumulation. *J Jpn Assoc Dietary Fiber Res* **4**, 59–65.
37. Handa A, Gennadios MA, Hanna CL, *et al.* (1999) Physical and molecular properties of egg-white lipid films. *J Food Sci* **64**, 860–864.
38. Sakai K, Ushiyama Y & Manabe S (1999) Peptic and pancreatic digestibility of raw and heat-treated hen's egg white protein. *Jpn J Pediatr Allergy Clin Immunol* **13**, 36–42.
39. Gargouri Y, Julien R, Sugihara A, *et al.* (1984) Inhibition of pancreatic and microbial lipase by proteins. *Bioclim Biophys Acta* **795**, 326–331.
40. Okuno A, Tamemoto H, Tobe K, *et al.* (1998) Troglitazone increases the number of small adipocytes without the change of white adipose tissue mass in obese Zucker rats. *J Clin Invest* **101**, 1354–1361.