九州大学学術情報リポジトリ Kyushu University Institutional Repository

Synthesis and Evaluation of 8-halogenated-7-deaza-2'-deoxy-guanosine as 8-oxo-2'-deoxy-guanosine analogues

尹, 貽貞

https://hdl.handle.net/2324/1500653

出版情報:九州大学, 2014, 博士(創薬科学), 課程博士

バージョン:

権利関係:やむを得ない事由により本文ファイル非公開(2)

氏 名	Yizhen Yin (尹 貽貞)
論 文 名	Synthesis and evaluation of
	8-halogenated-7-deaza-2'-deoxy-guanosine as
	8-oxo-2'-deoxy-guanosine analogues
論文調査委員	主查 九州大学大学院薬学府教授 佐々木茂貴
	副查 九州大学大学院薬学府教授 王子田彰夫
	副查 九州大学大学院薬学府准教授 麻生真理子
	副查 九州大学大学院薬学府准教授 谷口陽祐

論文審査の結果の要旨

Introduction. 8-Oxo-2'-deoxyguanosine (8-oxo-dG) is a representative nucleoside damage that is formed by oxidation of 2'-deoxyguanosine (dG) with reactive oxygen species (ROS), and its presence has been linked to aging, cancer, etc [1]. Unlike dG, 8-oxo-dG forms stable base pairs with both 2'-deoxycytidine (dC) and 2'-deoxyadenosine (dA). Based on the base-pairing properties of 8-oxo-dG, DNA polymerases incorporate 8-oxo-dGTP opposite dA and dATP opposite 8-oxo-dG, causing AT to CG and GC to TA transversion mutations. To suppress the genotoxicity of 8-oxo-dG and protect the genome integrity, hOGG1 can excise 8-oxo-dG from 8-oxo-dG:dC base pairs within duplex DNA. And hMYH provides the defense by removing dA opposite 8-oxo-dG. To prevent the incorporation of 8-oxo-dGTP into DNA, hMTH1 hydrolyzes 8-oxo-dGTP to 8-oxo-dGMP that is further hydrolyzed by nucleotidase. Recently, some DNA

repair enzymes such as DNA polymerase β and hOGG1 have been regarded as antitumor targets. Especially, hMTH1 is responsible for removing of oxidized nucleotides and required for survival of cancer cells [2]. In this study, 8-halogenated-7-deaza-dG

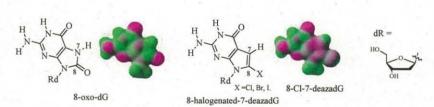


Figure 1. Structures of 8-oxo-dG and 8-halogenated-7-deazadG derivatives.

derivatives were designed as 8-oxo-dG analogues to elucidate the contributions of N7-H and C8-oxygen to the base pairing, replication and repair of 8-oxo-dG, and efforts have been devoted to find out functional inhibitors of DNA repair enzymes among the 8-halogenated-7-deaza-dG derivatives (Figure 1).

Results.

1. Synthesis and base pairing properties of 8-halogenated-7-deaza-dG derivatives.

The syntheses of 8-halogenated-7-deaza-dG derivatives were achieved via the reaction between acetylated 7-deaza-dG and N-halogenated succinimides. These compounds were incorporated into the central part of 13-mer oligonucleotides. The properties of these derivatives were investigated by computational, NMR and thermal denaturing studies. The significant upfield shift of the C-2' signals and characteristic downfield shift of H-2' signals indicated that 8-halogenated-7-deaza-dG derivatives prefer syn-conformation in DMSO solution similarly to 8-oxo-dG. It was shown that the base pair of 8-halogenated-7-deaza-dG with dC was destabilized compared with dG, supporting their preference for syn conformation. Unlike 8-oxo-dG, 8-halogenated-7-deaza-dG did not form a stable base pair with dA, most likely due to the lack of N7-H hydrogen bonding with dA. Therefore, the newly-designed 8-halogenated-7-deaza-dG derivatives resemble 8-oxo-dG in shape and preference for syn conformation, but they do not form Hoogsteen base pair with the opposite dA.

2. Recognition and excision of 8-halogenated-7-deazadG in DNA duplex by 8-oxo-dG glycosidase.

The recognition and excision of 8-halogenated-7-deazadG derivatives in DNA duplex were investigated using Fpg and hOGG1. 8-Halogenated-7-deazadG derivatives, especially 8-Cl-7-deazadG, were good glycosidase substrates for Fpg. However, 8-halogenated-7-deazadG derivatives were slightly excised by hOGG1. Kinetic properties of the reaction were analyzed by quartz crystal microbalance (QCM). In the case of Fpg, the association rate constant (k_{on}) for dG or 7-deaza-dG was smaller than that for 8-oxo-dG and 8-halogenated-7-deazadG, suggesting that introducing C8-oxygen or C8-halogen help to the recognition by Fpg. Interestingly, the dissociation rate constants (k_{off}) for 7-deaza-dG derivatives were similar to 8-oxo-dG, implying the importance of the presence of hydrogen at 7-position. In the case of hOGG1, 8-oxo-dG exhibited much lower k_{off} value than the other compounds, probably arising from the strong hydrogen bonding between 7-NH with Gly42 in the active site of hOGG1. Although 8-Cl- and 8-Br-7-deazadG had lower k_{off} value than 8-oxo-dG, they exhibited higher k_{on} resulting in the similar dissociate constant to 8-oxo-dG. Accordingly, it has been demonstrated that 8-halogenated-7-deaza-dG containing duplexes are competitive inhibitors for the glycosidase activity of hOGG1 to excise 8-oxo-dG in duplex DNA.

3. Synthesis and evaluation of 8-halogenated-7-deaza-dGTP.

The triphosphate derivatives, 7-deaza-dGTP, 8-Cl-, 8-Br- and 8-I-7-deaza-dGTP, were synthesized and tested for the reactivity for hMTH1. It should be noted that 8-Halogenated- 7-deaza-dGTP derivatives were hardly hydrolyzed by hMTH1, but showed competitive inhibitory activity against 8-oxo-dGTP hydrolysis by hMTH1. Therefore, it is expected that 7-deazadGTP and 8-halogenated-7-deazadGTP would show antitumor activity by targeting hMTH1. It was found that 8-halogenated-7-deazadGTP were only slightly incorporated into DNA to pair with dC and hardly incorporated to pair with dA by KF-exo and human polymerase β. Moreover, 8-halogenated-7-deazadG derivatives in duplex DNA were tested to be difficult to pair with dA during replication process. Therefore, 8-halogenated-7-deazadGTP derivatives are expected to have little side effects, further supporting their potentials as antitumor agents.

Conclusion. 8-Halogenated-7-deaza-dG derivatives were designed and synthesized as 8-oxo-dG analogues, and their chemical and biological properties were evaluated. 8-Halogenated-7-deaza-dG derivatives resemble 8-oxo-dG in shape and preference for syn-conformation, but they do not form Hoogsteen base pair with the opposite dA. Interestingly, 8-halogenated-7-deaza-dG derivatives in duplex DNA, especially 8-Cl-7-deaza-dG, were good glycosidase substrates for Fpg and strong binders to hOGG1. Accordingly, 8-halogenated-7-deaza-dG derivatives in duplex DNA demonstrated competitive inhibition for the glycosidase activity of hOGG1 to excise 8-oxo-dG in duplex DNA. Remarkable finding was that 8-halogenated-7-deazadGTP exhibited strong inhibition against hMTH1 at nanomolar concentrations, suggesting their potentials as antitumor agents.

Thus, this study has been approved to deserve a doctorate degree.

References

- 1. Burrows, C. J.; Muller, J. G. Chem. Rev., 1998, 98, 1109-1152.
- Gad H.; Koolmeister T.; Jemth A.S.; Eshtad S.; Jacques S.A.; Ström C.E. et al. Nature, 2014, 508, 215-221; Huber K.V.; Salah E.; Radic B.; Gridling M.; Elkins J.M.; Stukalov A.; Jemth A.S. et al. Nature, 2014, 508, 222-227.

Publications

- 1. <u>Yin, Y.</u>; Taniguchi, Y.; Sasaki, S. Synthesis of 8-halogenated-7-deaza-2'-deoxyguanosine as an 8-oxo-2'-deoxyguanosine analogue and evaluation of its base pairing properties. *Tetrahedron*, **2014**, 70, 2040-2047.
- Yin, Y.; Sasaki, S.; Taniguchi, Y. Recognition and excision properties of 8-Halogenated-7-deaza-2'-deoxyguanosine as 8-oxo-2'-deoxyguanosine analogue by Fpg and hOGG1. Submitted.
- 3. Yin, Y.; Sasaki, S.; Taniguchi, Y. The triphosphate derivatives, 7-deaza-dGTP, 8-Cl-, 8-Br- and 8-I-7-deaza-dGTP, as competitive inhibitors for hMTH1, in preparation.